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(54) SOLUBLE RECEPTOR BR43X2 AND METHODS OF USING THEM FOR THERAPY

LÖSLICHE REZEPTOREN BR43X2 UND VERFAHREN ZU DEREN THERAPEUTISCHEN
VERWENDUNG

RECEPTEURS SOLUBLES BR43x2 ET PROCEDES D'UTILISATION

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(56) References cited:
**EP-A- 0 869 180 WO-A-98/18921
WO-A-98/39361**

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- **G-U VON BÜLOW AND R J BRAM: "NF-AT activation induced by a CAML-interacting member of the tumor necrosis factor receptor superfamily" SCIENCE., vol. 278, 3 October 1997 (1997-10-03), pages 138-141, XP002140938 AAAS. LANCASTER, PA., US cited in the application**
- **J A GROSS ET AL.: "TACI and BCMA are receptors for a TNF homologue implicated in B-cell autoimmune disease" NATURE., vol. 404, 27 April 2000 (2000-04-27), pages 995-999, XP002140939 MACMILLAN JOURNALS LTD. LONDON., GB ISSN: 0028-0836**

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Description**BACKGROUND OF THE INVENTION**

- [0001]** Cellular interactions which occur during an immune response are regulated by members of several families of cell surface receptors, including the tumor necrosis factor receptor (TNFR) family. The TNFR family consists of a number of integral membrane glycoprotein receptors many of which, in conjunction with their respective ligands, regulate interactions between different hematopoietic cell lineages (Smith et al., The TNF Receptor Superfamily of Cellular and Viral Proteins: Activation, Costimulation and Death, 76:959-62, 1994; Cosman, Stern Cells 12:440-55, 1994).
- [0002]** One such receptor is TACI, transmembrane activator and CAML-interactor (von Bülow and Bram, Science 228:138-41, 1997 and WIPO Publication WO 98/39361). TACI is a membrane bound receptor having an extracellular domain containing two cysteine-rich pseudo-repeats, a transmembrane domain and a cytoplasmic domain that interacts with CAML (calcium-modulator and cyclophilin ligand), an integral membrane protein located at intracellular vesicles which is a co-inducer of NF-AT activation when overexpressed in Jurkat cells. TACI is associated with B cells and a subset of T cells. von Bülow and Bram (*ibid.*) report that the ligand for TACI is not known.
- [0003]** The polypeptides of the present invention, a TACI isoform having only one cysteine-rich pseudo-repeat (BR43x2), TACI and a related B cell protein, BCMA (Gras et al., Int. Immunol. 17:1093-106, 1995) were found to bind to the TNF ligand, ztnf4, now known as neutrokinin α (WIPO Publication, WO 98/18921), BLyS (Moore et al., Science, 285:260-3, 1999), BAFF (Schneider et al., J. Exp. Med. 189:1747-56, 1999), TALL-1 (Shu et al., J. Leukoc. Biol. 65: 680-3, 1999) or THANK (Mukhopadhyay et al., J. Biol. Chem. 274:15978-81, 1999). As such, BR43x2, TACI, and BCMA would be useful to regulate the activity of ztnf4 in particular, the activation of B cells.
- [0004]** Towards this end, the present invention provides protein therapeutics for modulating the activity of ztnf4 or other BR43x2, TACI or BCMA ligands, related compositions and methods as well as other uses that should be apparent to those skilled in the art from the teachings herein.
- [0005]** Accordingly, in a first aspect the invention provides use of a compound in the manufacture of a medicament for inhibiting ztnf4 activity in a mammal, wherein the compound is selected from the group consisting of: a) a polypeptide comprising the extracellular domain of BR43x2; b) a soluble polypeptide comprising the extracellular domain of TACI; c) a polypeptide comprising the extracellular domain of BCMA; d) a polypeptide comprising the sequence of SEQ ID NO:10; e) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:2; f) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:4; g) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:6; h) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:8; i) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:10; k) a polypeptide of SEQ ID NO:4; l) amino acid residues 1-166 of SEQ ID NO:6; and m) amino acid residues 1-150 of SEQ ID NO:8.
- [0006]** Within one embodiment the compound is a fusion protein consisting of a first portion and a second portion joined by a peptide bond, said first portion comprising a polypeptide selected from the group consisting of: a) a polypeptide comprising the sequence of SEQ ID NO:8; b) a polypeptide comprising amino acid residues 25-58 of SEQ ID NO:2; c) a polypeptide comprising amino acid residues 34-66 of SEQ ID NO:6; d) a polypeptide comprising amino acid residues 71-104 of SEQ ID NO:6; e) a polypeptide comprising amino acid residues 25-104 of SEQ ID NO:6; f) a polypeptide comprising amino acid residues 8-37 of SEQ ID NO:8; g) a polypeptide comprising amino acid residues 41-88 of SEQ ID NO:8; h) a polypeptide comprising amino acid residues 8-88 of SEQ ID NO:8; and said second portion comprising another polypeptide. Within another embodiment the first portion further comprises a polypeptide selected from the group consisting of: a) amino acid residues 59-120 of SEQ ID NO:2; b) amino acid residues 105-166 of SEQ ID NO:6; and c) amino acid residues 89-150 of SEQ ID NO:8. Within another embodiment the first portion is selected from the group consisting of: a) a polypeptide comprising the extracellular domain of BR43x2; b) a polypeptide comprising the extracellular domain of TACI; and c) a polypeptide comprising the extracellular domain of BCMA. Within a related embodiment the first portion is selected from the group consisting of: a) a polypeptide of SEQ ID NO:4; b) amino acid residues 1-154 of SEQ ID NO:6; and c) amino acid residues 1-48 of SEQ ID NO:8. Within another related embodiment the second portion is an immunoglobulin heavy chain constant region.
- [0007]** Within another embodiment the antibody or antibody fragment is selected from the group consisting of: a) polyclonal antibody; b) murine monoclonal antibody; c) humanized antibody derived from b); and d) human monoclonal antibody. Within a related embodiment the antibody fragment is selected from the group consisting of F(ab'), Fab', Fab, Fv, scFv. Within another embodiment the mammal is a primate.
- [0008]** Within another embodiment the ztnf4 activity is associated with B lymphocytes. Within another related embodiment the ztnf4 activity is associated with activated B lymphocytes. Within yet another embodiment the ztnf4 activity is associated with resting B lymphocytes. Within another embodiment the ztnf4 activity is associated with antibody production. Within a related embodiment the antibody production is associated with an autoimmune disease. Within a related embodiment the said autoimmune disease is systemic lupus erythematosus, myasthenia gravis, multiple scl-

rosis, or rheumatoid arthritis. Within another embodiment the ztnf4 activity is associated with asthma, bronchitis or emphysema. Within still another embodiment the ztnf4 activity is associated with end stage renal failure. Within yet another embodiment the ztnf4 activity is associated with renal disease. Within a related embodiment the renal disease is glomerulonephritis, vasculitis, nephritis or pyronephritis. Within yet another embodiment the renal disease is associated with renal neoplasms, multiple myelomas, lymphomas, light chain neuropathy or amyloidosis. Within another embodiment the ztnf4 activity is associated with effector T cells. Within a related embodiment the ztnf4 activity is associated with moderating immune response. Within yet another embodiment the activity is associated with immunosuppression. Within yet another embodiment immunosuppression is associated with graft rejection, graft verses host disease or inflammation. Within another embodiment the activity is associated with autoimmune disease. Within a related embodiment the autoimmune disease is insulin dependent diabetes mellitus or Crohn's Disease. Within another embodiment the ztnf4 activity is associated with inflammation. Within a related embodiment the inflammation is associated with joint pain, swelling, anemia, or septic shock. Within another aspect the invention provides a method for the manufacture of a medicament for inhibiting BR43x2, TACI or BCMA receptor-ztnf4 engagement including an amount of a compound as described above. Within another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with B lymphocytes. Within another related embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with activated B lymphocytes. Within yet another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with resting B lymphocytes.

[0009] Within another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with antibody production. Within a related embodiment the antibody production is associated with an autoimmune disease. Within a related embodiment the said autoimmune disease is systemic lupus erythematosus, myasthenia gravis, multiple sclerosis, or rheumatoid arthritis. Within another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with asthma, bronchitis or emphysema. Within still another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with end stage renal failure. Within yet another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with renal disease. Within a related embodiment the renal disease is glomerulonephritis, vasculitis, nephritis or pyronephritis. Within yet another embodiment the renal disease is associated with renal neoplasms, multiple myelomas, lymphomas, light chain neuropathy or amyloidosis. Within another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with effector T cells. Within a related embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with moderating immune response. Within yet another embodiment the activity is associated with immunosuppression. Within yet another embodiment immunosuppression is associated with graft rejection, graft verses host disease or inflammation. Within another embodiment the activity is associated with autoimmune disease. Within a related embodiment the autoimmune disease is insulin dependent diabetes mellitus or Crohn's Disease. Within another embodiment the BR43x2, TACI or BCMA receptor-ligand engagement is associated with inflammation. Within a related embodiment the inflammation is associated with joint pain, swelling, anemia, or septic shock.

[0010] Within another aspect the invention provides an isolated polynucleotide molecule encoding a polypeptide of SEQ ID NO:2. Also provided is an isolated polynucleotide molecule of SEQ ID NO:1. Within a related embodiment is provided an expression vector comprising the following operably linked elements: a transcription promoter; a polynucleotide molecule as described above, and a transcription terminator. Within another embodiment the expression vector further comprises a secretory receptor-ligand engagement sequence operably linked to said polynucleotide molecule. Also provided is a cultured cell into which has been introduced an expression vector as described above, wherein said cultured cell expresses said polypeptide encoded by said polynucleotide segment. The invention further provides a method of producing a polypeptide comprising: culturing a cell into which has been introduced an expression vector as described above; whereby said cell expresses said polypeptide encoded by said polynucleotide molecule; and recovering said expressed polypeptide. The invention also provides an isolated polypeptide having the sequence of SEQ ID NO:2. Within a related embodiment the polypeptide is in combination with a pharmaceutically acceptable vehicle.

BRIEF DESCRIPTION OF THE DRAWING

[0011]

Figure 1 shows a multiple amino acid sequence alignment between BR43x2, TACI (von Bülow and Bram, *ibid.*) (SEQ ID NO:8), BCMA (Gras et al., *ibid.*) (SEQ ID NO:6) and BR43x1 (SEQ ID NO:9). The cysteine-rich pseudo repeats and transmembrane domain are noted.

Figure 2 shows a Scatchard plot analysis of soluble $\text{I}^{125}\text{-ztnf4}$ binding to TACI and BCMA expressed by stable BHK transfecants.

Figure 3A shows ztnf4 co-activating human B lymphocytes to proliferate and secrete immunoglobulin.

Figure 3B shows levels of IgM and IgG measured in supernatants obtained from B cells stimulated with soluble

ztnf4 in the presence of IL4 or IL4+IL5 after 9 days in culture.

Figure 4 shows human peripheral blood B cells stimulated with soluble ztnf4 or control protein (ubiquitin) in the presence of IL-4 for 5 days *in vitro*. Purified TACI-Ig, BCMA-Ig and control Fc were tested for inhibition of ztnf4 specific proliferation.

5 Figure 5A shows results from ztnf4 transgenic animals that have developed characteristics of SLE.

Figure 5B shows lymph node, spleen and thymus cells from ztnf4 transgenic animals stained with antibodies to CD5, CD4 and CD8.

Figure 5C shows total IgM, IgG and IgE levels in serum from transgenic ztnf4 animals ranging from 6 to 23 weeks of age.

10 Figure 5D shows amyloid deposition and thickened mesangium of the glomeruli identified in kidney sections from ztnf4 transgenic animals.

Figure 5E shows effector T cells in ztnf4 transgenic mice.

Figures 6A and B show elevated ztnf4 levels in serum obtained from ZNBWF1 mice and MRL/lpr/lpr mice that correlates with development of SLE.

15 Figure 7 shows the percentage of ZNBWF1 mice that develop proteinuria over the course of the study.

Figure 8 shows anti-dsDNA levels by ELISA from ztnf4 transgenic mice and control litter mates compared to serum from ZNBWF1 and MRL/lpr/lpr mice.

[0012] These and other aspects of the invention will become evident upon reference to the following detailed description.

DETAILED DESCRIPTION OF THE INVENTION

[0013] Prior to setting forth the invention, it may be helpful to an understanding thereof to set forth definitions of certain terms to be used hereinafter:

Affinity tag: is used herein to denote a polypeptide segment that can be attached to a second polypeptide to provide for purification or detection of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract, protein A (Nilsson et al., *EMBO J.* 4:1075, 1985; Nilsson et al., *Methods Enzymol.* 198:3, 1991), glutathione S transferase (Smith and Johnson, *Gene* 67:31, 1988), Glu-Glu affinity tag (Grussenmeyer et al., *Proc. Natl. Acad. Sci. USA* 82:7952-4, 1985), substance P, Flag™ peptide (Hopp et al., *Biotechnology* 6:1204-10, 1988), streptavidin binding peptide, or other antigenic epitope or binding domain. See, in general, Ford et al., *Protein Expression and Purification* 2: 95-107, 1991. DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

Allelic variant: Any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (i.e., no change in the encoded polypeptide), or may encode polypeptides having altered amino acid sequence. The term "allelic variant" is also used herein to denote a protein encoded by an allelic variant of a gene. Also included are the same protein from the same species which differs from a reference amino acid sequence due to allelic variation. Allelic variation refers to naturally occurring differences among individuals in genes encoding a given protein.

Amino-terminal and carboxyl-terminal: are used herein to denote positions within polypeptides and proteins. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide or protein to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a reference sequence within a protein is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete protein.

Complement/anti-complement pair: Denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions. For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair. Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the complement/anti-complement pair preferably has a binding affinity of <10⁻⁹ M.

Contig: Denotes a polynucleotide that has a contiguous stretch of identical or complementary sequence to another polynucleotide. Contiguous sequences are said to "overlap" a given stretch of polynucleotide sequence either in their entirety or along a partial stretch of the polynucleotide. For example, representative contigs to the polynucleotide sequence 5'-ATGGCTTAGCTT-3' are 5'-TAGCTTgagtct-3' and 3'-gtcgacTACCGA-5'.

Complements of polynucleotide molecules: Denotes polynucleotide molecules having a complementary base se-

quence and reverse orientation as compared to a reference sequence. For example, the sequence 5' ATGCACGG 3' is complementary to 5' CCCGTGCAT 3'.

Degenerate Nucleotide Sequence or Degenerate Sequence: Denotes a sequence of nucleotides that includes one or more degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

Expression vector: A DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments may include promoter and terminator sequences, and optionally one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, and the like. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

Isoform: refers to different forms of a protein that may be produced from different genes or from the same gene by alternate splicing. In some cases, isoforms differ in their transport activity, time of expression in development, tissue distribution, location in the cell or a combination of these properties.

Isolated polynucleotide: denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free of other extraneous or unwanted coding sequences; and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of associated regions will be evident to one of ordinary skill in the art (see for example, Dynan and Tijan, Nature 316:774-78, 1985).

Isolated polypeptide or protein: is a polypeptide or protein that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other polypeptides, particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably greater than 99% pure. When used in this context, the term "isolated" does not exclude the presence of the same polypeptide in alternative physical forms, such as dimers or alternatively glycosylated or derivatized forms.

Operably linked: As applied to nucleotide segments, the term "operably linked" indicates that the segments are arranged so that they function in concert for their intended purposes, e.g., transcription initiates in the promoter and proceeds through the coding segment to the terminator.

Ortholog: Denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different species. Sequence differences among orthologs are the result of speciation.

Polynucleotide: denotes a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules. Sizes of polynucleotides are expressed as base pairs (abbreviated "bp"), nucleotides ("nt"), or kilobases ("kb"). Where the context allows, the latter two terms may describe polynucleotides that are single-stranded or double-stranded. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs". It will be recognized by those skilled in the art that the two strands of a double-stranded polynucleotide may differ slightly in length and that the ends thereof may be staggered as a result of enzymatic cleavage; thus all nucleotides within a double-stranded polynucleotide molecule may not be paired. Such unpaired ends will in general not exceed 20 nt in length.

Polypeptide: Is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides".

Promoter: Denotes a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

Protein: is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-peptidic components, such as carbohydrate groups. Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

Receptor: A cell-associated protein, or a polypeptide subunit of such protein, that binds to a bioactive molecule (the "ligand") and mediates the effect of the ligand on the cell. Binding of ligand to receptor results in a change in the receptor (and, in some cases, receptor multimerization, i.e., association of identical or different receptor subunits) that causes interactions between the effector domain (s) of the receptor and other molecule(s) in the cell. These interactions in turn lead to alterations in the metabolism of the cell. Metabolic events that are linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, cell proliferation, in-

creases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. BR43x2 has characteristics of TNF receptors, as discussed in more detail herein.

5 Secretory signal sequence: A DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized. The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

10 Soluble receptor: A receptor polypeptide that is not bound to a cell membrane. Soluble receptors are most commonly ligand-binding receptor polypeptides that lack transmembrane and cytoplasmic domains. Soluble receptors can comprise additional amino acid residues, such as affinity tags that provide for purification of the polypeptide or provide sites for attachment of the polypeptide to a substrate. Many cell-surface receptors have naturally occurring, soluble counterparts that are produced by proteolysis or translated from alternatively spliced mRNAs. Receptor polypeptides are said to be substantially free of transmembrane and intracellular polypeptide segments when they lack sufficient portions of these segments to provide membrane anchoring or signal transduction, respectively.

15 [0014] Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to $\pm 10\%$.

20 [0015] All references cited herein are incorporated by reference in their entirety.

25 [0016] The present invention is based in part upon the discovery of a 1192 bp DNA sequence (SEQ ID NO:1) and corresponding polypeptide sequence (SEQ ID NO:2) which is an isoform of the receptor TACI. The isoform has been designated BR43x2. A soluble form of BR43x2 is disclosed in SEQ ID NO:4, the polynucleotide encoding the soluble receptor in SEQ ID NO:3. As is described in more detail herein, the BR43x2 receptor-encoding polynucleotides and polypeptides of the present invention were initially identified by signal trap cloning using a human RPMI 1788 library and the N- or C-terminally FLAG-tagged, biotin- or FITC-labeled tumor necrosis factor ligand ztnf4, now known as neutrokine α (WIPO WO98/18921), BLyS (Moore et al., *ibid.*), BAFF (Schneider et al., *ibid.*), TALL-1 (Shu et al., *ibid.*) or THANK (Mukhopadhyay et al., *ibid.*). Positive pools were identified by ligand binding, broken down to single clones, the cDNA isolated and sequenced. A comparison of the BR43x2 deduced amino acid sequence (as represented in SEQ ID NO:2) with known tumor necrosis factor receptors indicated that BR43x2 is an isoform of TACI, having a single, poorly conserved, cysteine-rich pseudo-repeat.

30 [0017] Structurally, the TNF receptor family is characterized by an extracellular portion composed of several modules called, historically, "cysteine-rich pseudo-repeats". A prototypical TNFR family member has four of these pseudo-repeats, each about 29-43 residues long, one right after the other. A typical pseudo-repeat has 6 cysteine residues. They are called pseudo-repeats because, although they appear to originate from a common ancestral module, they do not repeat exactly: pseudo-repeats #1, #2, #3 and #4 have characteristic sequence features which distinguish them from one another. The crystal structure of the p55 TNF receptor revealed that each pseudo-repeat corresponds to one folding domain, and that all four pseudo-repeats fold into the same tertiary structure, held together internally by disulfide bonds.

40 [0018] TACI contains two cysteine-rich pseudo-repeats (von Bülow and Bram, *ibid.*), the first is conserved in structure with other members of the TNF receptor family, the second is less conserved. The BR43x2 isoform of the present invention lacks the first TACI cysteine-rich pseudo-repeat, retaining only the second, less conserved repeat.

45 [0019] Sequence analysis of a deduced amino acid sequence of BR43x2 as represented in SEQ ID NO:2 indicates the presence of a mature protein having an extracellular domain (residues 1-120 of SEQ ID NO:2) which contains one cysteine-rich pseudo-repeat (residues 25-58 of SEQ ID NO:2), a transmembrane domain (residues 121-133 of SEQ ID NO:2) and a cytoplasmic domain (residues 134-247 of SEQ ID NO:2). The cysteine-rich pseudo-repeat of BR43x2 has 6 conserved cysteine residues (residues 25, 40, 43, 47, 54 and 58 of SEQ ID NO:2), a conserved aspartic acid residue (residue 34 of SEQ ID NO:2) and two conserved leucine residues (residues 36 and 37 of SEQ ID NO:2) and shares 46% identity with the first cysteine-rich pseudo-repeat of TACI (SEQ ID NO:6) and 35% identity with the cysteine-rich pseudo-repeat of BCMA (SEQ ID NO:8) (Figure 1). The cysteine-rich pseudo-repeat can be represented by the following motif:

55 CX [QEK] [QEKNRDHS] [QE] X{0-2} [YFW] [YFW] DXLLX{2}C [IMLV] XCX{3}
CX{6-8}CX{2}[YF]C (SEQ ID NO:10),

wherein C represents the amino acid residue cysteine, Q glutamine, E glutamic acid, K lysine, N asparagine, R

arginine, D aspartic acid, H histidine, S serine, Y tyrosine, F phenylalanine, W tryptophan, L leucine, I isoleucine, V valine and X represents any naturally occurring amino acid residue except cysteine. Amino acid residues in square brackets "[]" indicate the allowed amino acid residue variation at that position. The number in the braces "{}" indicates the number of allowed amino acid residues at that position.

5 [0020] The present invention also provides soluble polypeptides of from 32 to 40 amino acid residues in length as provided by SEQ ID NO:10.

[0021] The soluble BR43x2 receptor, as represented by residues 1-120 of SEQ ID NO:4, contains one cysteine-rich pseudo-repeat (residues 25-58 of SEQ ID NO:4) and lacks the transmembrane and cytoplasmic domains of BR43x2 as described in SEQ ID NO:2.

10 [0022] Those skilled in the art will recognize that these domain boundaries are approximate, and are based on alignments with known proteins and predictions of protein folding. These features indicate that the receptor encoded by the DNA sequences of SEQ ID NOs:1 and 3 is a member of the TNF receptor family.

15 [0023] Northern blot and Dot blot analysis of the tissue distribution of the mRNA corresponding to nucleotide probes to BR43x1 which are predicted to detect BR43x2 expression showed expression in spleen, lymph node, CD19⁺ cells, weakly in mixed lymphocyte reaction cells, Daudi and Raji cells. Using reverse transcriptase PCR BR43x1 was detected in B cells only and not in activated T cells as had been reported for TACI (von Bülow and Bram, *ibid.*). Using a BR43x2 probe that overlaps 100% with the corresponding TACI sequence, TACI and BR43x2 were detected in spleen, lymph node and small intestine, stomach, salivary gland, appendix, lung, bone marrow, fetal spleen, CD 19⁺ cells, and Raji cells.

20 [0024] Using Northern Blot analysis BCMA was detected in small intestine, spleen, stomach, colon, appendix, lymph node, trachea, and testis. BCMA was also detected in adenolymphoma, non-Hodgkins lymphoma, and parotid tumor, detected faintly in CD 8⁺, CD 19⁺, MLR cells, Daudi, Raji and Hut 78 cells.

[0025] Northern blot analysis was also done using murine ztnf4 (SEQ ID NO:19) and like human TACI, BCMA, and BR43x2, murine ztnf4 expression was detected predominately in spleen and thymus. Murine ztnf4 was also expressed in lung and faint expression was detected in skin and heart.

25 [0026] The present invention also provides polynucleotide molecules, including DNA and RNA molecules, that encode the BR43x2 polypeptides disclosed herein. Those skilled in the art will readily recognize that, in view of the degeneracy of the genetic code, considerable sequence variation is possible among these polynucleotide molecules. SEQ ID NO:11 is a degenerate DNA sequence that encompasses all DNAs that encode the soluble BR43x2 polypeptide of SEQ ID NO:4. Similarly, SEQ ID NO:12 is a degenerate DNA sequence that encompasses all DNAs that encode the BR43x2 polypeptide of SEQ ID NO:2. Those skilled in the art will recognize that the degenerate sequence of SEQ ID NO:12 also provides all RNA sequences encoding SEQ ID NO:4 by substituting U for T. Thus, BR43x2 polypeptide-encoding polynucleotides comprising nucleotide 1 to nucleotide 360 of SEQ ID NO:11, nucleotide 1 to 741 of SEQ ID NO:12 and their RNA equivalents are contemplated by the present invention. Table 1 sets forth the one-letter codes used within SEQ ID NOs:11 and 12 to denote degenerate nucleotide positions. "Resolutions" are the nucleotides denoted by a code letter. "Complement" indicates the code for the complementary nucleotide(s). For example, the code Y denotes either C or T, and its complement R denotes A or G, A being complementary to T, and G being complementary to C.

TABLE 1

Nucleotide	Resolution	Complement	Resolution
A	A	T	T
C	C	G	G
G	G	C	C
T	T	A	A
R	A G	Y	C T
Y	C T	R	A G
M	A C	K	G T
K	G T	M	A C
S	C G	S	C G
W	A T	W	A T
H	A C T	D	A G T
B	C G T	V	A C G
V	A C G	B	C G T
D	A G T	H	A C T

TABLE 1 (continued)

Nucleotide	Resolution	Complement	Resolution
N	A C G T	N	A C G T

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[0027] The degenerate codons used in SEQ ID NOs:11 and 12, encompassing all possible codons for a given amino acid, are set forth in Table 2.

TABLE 2

	Amino Acid	One Letter Code	Codons	Degenerate Codon
10	Cys	C	TGC TGT	TGY
	Ser	S	AGC AGT TCA TCC TCG TCT	WSN
	Thr	T	ACA ACC ACG ACT	ACN
	Pro	P	CCA CCC CCG CCT	CCN
	Ala	A	GCA GCC GCG GCT	GCN
	Gly	G	GGA GGC GGG GGT	GGN
	Asn	N	AAC AAT	AAY
	Asp	D	GAC GAT	GAY
	Glu	E	GAA GAG	GAR
	Gln	Q	CAA CAG	CAR
15	His	H	CAC CAT	CAY
	Arg	R	AGA AGG CGA CGC CGC CGT	MGN
	Lys	K	AAA AAG	AAR
	Met	M	ATG	ATG
	Ile	I	ATA ATC ATT	ATH
	Leu	L	CTA CTC CTG CTT TTA TTG	YTN
	Val	V	GTA GTC GTG GTT	GTN
	Phe	F	TTC TTT	TTY
	Tyr	Y	TAC TAT	TAY
	Trp	W	TGG	TGG
20	Ter	.	TAA TAG TGA	TRR
	Asn Asp	B		RAY
	Glu Gln	Z		SAR
	Any	X		

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[0028] One of ordinary skill in the art will appreciate that some ambiguity is introduced in determining a degenerate codon, representative of all possible codons encoding each amino acid. For example, the degenerate codon for serine (WSN) can, in some circumstances, encode arginine (AGR), and the degenerate codon for arginine (MGN) can, in some circumstances, encode serine (AGY). A similar relationship exists between codons encoding phenylalanine and leucine. Thus, some polynucleotides encompassed by the degenerate sequence may encode variant amino acid sequences, but one of ordinary skill in the art can easily identify such variant sequences by reference to the amino acid sequences of SEQ ID NOs:2 and 4. Variant sequences can be readily tested for functionality as described herein.

[0029] One of ordinary skill in the art will also appreciate that different species can exhibit "preferential codon usage." In general, see, Grantham, et al., *Nuc. Acids Res.* 8:1893-912, 1980; Haas, et al. *Curr. Biol.* 6:315-24, 1996; Wain-Hobson, et al., *Gene* 13:355-64, 1981; Grosjean and Fiers, *Gene* 18:199-209, 1982; Holm, *Nuc. Acids Res.* 14:3075-87, 1986; Ikemura, *J. Mol. Biol.* 158:573-97, 1982. As used herein, the term "preferential codon usage" or "preferential codons" is a term of art referring to protein translation codons that are most frequently used in cells of a certain species, thus favoring one or a few representatives of the possible codons encoding each amino acid (See Table 2). For example, the amino acid threonine (Thr) may be encoded by ACA, ACC, ACG, or ACT, but in mammalian cells ACC is the most commonly used codon; in other species, for example, insect cells, yeast, viruses or bacteria, different Thr codons may be preferential. Preferential codons for a particular species can be introduced into the polynucleotides of the present invention by a variety of methods known in the art. Introduction of preferential codon sequences into recombinant DNA can, for example, enhance production of the protein by making protein translation more efficient within a particular cell type or species. Therefore, the degenerate codon sequences disclosed in SEQ ID NOs:11 and 12 serve as a template

for optimizing expression of polynucleotides in various cell types and species commonly used in the art and disclosed herein. Sequences containing preferential codons can be tested and optimized for expression in various species, and tested for functionality as disclosed herein.

[0030] The highly conserved amino acids in the cysteine-rich pseudo-repeat of BR43x2 can be used as a tool to identify new family members. For instance, reverse transcription-polymerase chain reaction (RT-PCR) can be used to amplify sequences encoding the extracellular ligand-binding domain, described above, from RNA obtained from a variety of tissue sources or cell lines. In particular, highly degenerate primers designed from the BR43x2 sequences are useful for this purpose.

[0031] Within preferred embodiments of the invention, isolated polynucleotides will hybridize to similar sized regions of SEQ ID NO:3, or to a sequence complementary thereto, under stringent conditions. In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typical stringent conditions are those in which the salt concentration is up to about 0.03 M at pH 7 and the temperature is at least about 60°C.

[0032] As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for isolating DNA and RNA are well known in the art. It is generally preferred to isolate RNA from RPMI 1788 cells, PBMNCs, resting or activated transfected B cells or tonsil tissue, although DNA can also be prepared using RNA from other tissues or isolated as genomic DNA. Total RNA can be prepared using guanidine HCl extraction followed by isolation by centrifugation in a CsCl gradient (Chirgwin et al., *Biochemistry* 18:52-94, 1979). Poly (A)⁺ RNA is prepared from total RNA using the method of Aviv and Leder (*Proc. Natl. Acad. Sci. USA* 69:1408-12, 1972). Complementary DNA (cDNA) is prepared from poly(A)⁺ RNA using known methods. Polynucleotides encoding BR43x2 polypeptides are then identified and isolated by, for example, hybridization or PCR.

[0033] Those skilled in the art will recognize that the sequences disclosed in SEQ ID NOs:1 and 3 represent a single allele of the human gene, and that allelic variation and alternative splicing is expected to occur. Allelic variants of the DNA sequences shown in SEQ ID NOs:1 and 3, including those containing silent mutations and those in which mutations result in amino acid sequence changes, are within the scope of the present invention, as are proteins which are allelic variants of SEQ ID NOs:2 and 4. Allelic variants and splice variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals or tissues according to standard procedures known in the art.

[0034] The receptor polypeptides of the present invention, including full-length receptor polypeptides, soluble receptors polypeptides, polypeptide fragments, and fusion polypeptides, can be produced in genetically engineered host cells according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook et al., *Molecular Cloning: A Laboratory Manual, Second Edition*, Cold Spring Harbor, NY, 1989; and Ausubel et al., eds., *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., NY, 1987.

[0035] In general, a DNA sequence encoding a BR43x2 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

[0036] To direct a BR43x2 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a signal sequence, leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of the BR43x2 polypeptide, or may be derived from another secreted protein (e.g., t-PA) or synthesized *de novo*. The secretory signal sequence is joined to the BR43x2 DNA sequence in the correct reading frame and positioned to direct the newly synthesized polypeptide into the secretory pathway of the host cell. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

[0037] Cultured mammalian cells are suitable hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection (Wigler et al., *Cell* 14:725, 1978; Corsaro and Pearson, *Somatic Cell Genetics* 7:603, 1981; Graham and Van der Eb, *Virology* 52:456, 1973), electroporation (Neumann et al., *EMBO J.* 1:841-45, 1982), DEAE-dextran mediated transfection (Ausubel et al., *ibid.*), and liposome-mediated transfection (Hawley-Nelson et al., *Focus* 15:73, 1993; Ciccarone et al., *Focus* 15:80, 1993). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.

S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham et al., *J. Gen. Virol.* 36:59-72, 1977), Jurkat (ATCC No. CRL-8129) , BaF3 (an interleukin-3 dependent pre-lymphoid cell line derived from murine bone marrow. See, Palacios and Steinmetz, *Cell* 41: 727-34, 1985; Mathey-Prevot et al., *Mol. Cell. Biol.* 6: 4133-5, 1986) and Chinese hamster ovary (e.g., CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978 and the adenovirus major late promoter.

[0038] Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems may also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g., hygromycin resistance, multi-drug resistance, puromycin acetyltransferase) can also be used. Alternative markers that introduce an altered phenotype, such as green fluorescent protein, or cell surface proteins such as CD4, CD8, Class I MHC, placental alkaline phosphatase may be used to sort transfected cells from untransfected cells by such means as FACS sorting or magnetic bead separation technology.

[0039] Other higher eukaryotic cells can also be used as hosts, including plant cells, insect cells and avian cells. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore)* 11:47-58, 1987. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, commonly derived from *Autographa californica nuclear polyhedrosis virus* (AcNPV). See, King and Possee, *The Baculovirus Expression System: A Laboratory Guide*, London, Chapman & Hall; O'Reilly et al., *Baculovirus Expression Vectors: A Laboratory Manual*, New York, Oxford University Press., 1994; and Richardson, Ed., *Baculovirus Expression Protocols. Methods in Molecular Biology*, Totowa, NJ, Humana Press, 1995. A second method of making recombinant BR43x2 baculovirus utilizes a transposon-based system described by Luckow (Luckow, et al., *J Virol* 67:4566-79, 1993). This system, which utilizes transfer vectors, is sold in the Bac-to-Bac™ kit (Life Technologies, Rockville, MD). This system utilizes a transfer vector, pFastBacI™ (Life Technologies) containing a Tn7 transposon to move the DNA encoding the BR43x2 polypeptide into a baculovirus genome maintained in *E. coli* as a large plasmid called a "bacmid." See, Hill-Perkins and Possee, *J. Gen. Virol.* 71:971-6, 1990; Bonning, et al., *J. Gen. Virol.* 75:1551-6, 1994; and, Chazenbalk, and Rapoport, *J. Biol. Chem.* 270:1543-9, 1995. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope tag at the C- or N-terminus of the expressed BR43x2 polypeptide, for example, a Glu-Glu epitope tag (Grussenmeyer et al., *Proc. Natl. Acad. Sci.* 82:7952-4, 1985). Using a technique known in the art, a transfer vector containing BR43x2 is transformed into *E. coli*, and screened for bacmids which contain an interrupted lacZ gene indicative of recombinant baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g. Sf9 cells. Recombinant virus that expresses BR43x2 is subsequently produced. Recombinant viral stocks are made by methods commonly used the art.

[0040] The recombinant virus is used to infect host cells, typically a cell line derived from the fall armyworm, *Spodoptera frugiperda*. See, in general, Glick and Pasternak, *Molecular Biotechnology: Principles and Applications of Recombinant DNA*, ASM Press, Washington, D.C., 1994. Another suitable cell line is the High FiveO™ cell line (Invitrogen) derived from *Trichoplusia ni* (U.S. Patent #5,300,435). Commercially available serum-free media are used to grow and maintain the cells. Suitable media are Sf900 II™ (Life Technologies) or ESF 921™ (Expression Systems) for the Sf9 cells; and Ex-cellO405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™ (Life Technologies) for the *T. ni* cells. The cells are grown up from an inoculation density of approximately 2-5 x 10⁵ cells to a density of 1-2 x 10⁶ cells at which time a recombinant viral stock is added at a multiplicity of infection (MOI) of 0.1 to 10, more typically near 3. Procedures used are generally described in available laboratory manuals (King and Possee, *ibid.*; O'Reilly, et al., *ibid.*; Richardson, *ibid.*). Subsequent purification of the BR43x2 polypeptide from the supernatant can be achieved using methods described herein.

[0041] Fungal cells, including yeast cells, can also be used within the present invention. Yeast species of particular interest in this regard include *Saccharomyces cerevisiae*, *Pichia pastoris*, and *Pichia methanolica*. Methods for transforming *S. cerevisiae* cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by,

for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in *Saccharomyces cerevisiae* 5 is the *POT1* vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago maydis*, *Pichia pastoris*, 10 *Pichia methanolica*, *Pichia guillermorandi* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., *J. Gen. Microbiol.* 132:3459-65, 1986 and Cregg, U.S. Patent No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* 15 are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

[0042] For example, the use of *Pichia methanolica* as host for the production of recombinant proteins is disclosed by Raymond, U.S. Patent No. 5,716,808, Raymond, U.S. Patent No. 5,736,383, Raymond et al., *Yeast* 14:11-23, 1998, and in international publication Nos. WO 97/17450, WO 97/17451, WO 98/02536, and WO 98/02565. DNA molecules for use in transforming *P. methanolica* will commonly be prepared as double-stranded, circular plasmids, which are 20 preferably linearized prior to transformation. For polypeptide production in *P. methanolica*, it is preferred that the promoter and terminator in the plasmid be that of a *P. methanolica* gene, such as a *P. methanolica* alcohol utilization gene (*AUG1* or *AUG2*). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. To facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. A preferred selectable marker for use in *Pichia methanolica* is a *P. methanolica* *ADE2* gene, which encodes phosphoribosyl-5-aminoimidazole carboxylase (AIRC; EC 4.1.1.21), which allows *ade2* host cells to grow in the absence of adenine. For 25 large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol utilization genes (*AUG1* and *AUG2*) are deleted. For production of secreted proteins, host cells deficient in vacuolar protease genes (*PEP4* and *PRB1*) are preferred. Electroporation is used to facilitate the introduction of a plasmid containing DNA encoding a polypeptide of interest into *P. methanolica* cells. It is preferred to transform *P. methanolica* cells by electroporation using an exponentially decaying, pulsed electric field having a field strength of 30 from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (t) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

[0043] Prokaryotic host cells, including strains of the bacteria *Escherichia coli*, *Bacillus* and other genera are also 35 useful host cells within the present invention. Techniques for transforming these hosts and expressing foreign DNA sequences cloned therein are well known in the art (see, e.g., Sambrook et al., *ibid.*). When expressing a BR43x2 polypeptide in bacteria such as *E. coli*, the polypeptide may be retained in the cytoplasm, typically as insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured 40 polypeptide can then be refolded and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

[0044] Transformed or transfected host cells are cultured according to conventional procedures in a culture medium 45 containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for 50 example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell. *P. methanolica* cells are cultured in a medium comprising adequate sources of carbon, nitrogen and trace nutrients at a temperature of about 25°C to 35°C. Liquid cultures are provided with sufficient aeration by conventional means, such as shaking of small flasks or sparging of fermentors. A preferred culture medium for *P. methanolica* is YEPD (2% D-glucose, 2% Bacto™ Peptone (Difco Laboratories, Detroit, MI), 1% Bacto™ yeast extract (Difco Laboratories), 0.004% adenine and 0.006% L-leucine).

[0045] Expressed recombinant BR43x2 polypeptides (or chimeric or fusion BR43x2 polypeptides) can be purified 55 using fractionation and/or conventional purification methods and media. It is preferred to provide the proteins or polypeptides of the present invention in a highly purified form, i.e. greater than 95% pure, more preferably greater than

99% pure. Ammonium sulfate precipitation and acid or chaotrope extraction may be used for fractionation of samples. Exemplary purification steps may include hydroxyapatite, size exclusion, FPLC and reverse-phase high performance liquid chromatography. Suitable anion exchange media include derivatized dextrans, agarose, cellulose, polyacrylamide, specialty silicas, and the like. PEI, DEAE, QAE and Q derivatives are preferred, with DEAE Fast-Flow Sepharose (Pharmacia, Piscataway, NJ) being particularly preferred. Exemplary chromatographic media include those media derivatized with phenyl, butyl, or octyl groups, such as Phenyl-Sepharose FF (Pharmacia), Toyopearl butyl 650 (Toso Haas, Montgomeryville, PA), Octyl-Sepharose (Pharmacia) and the like; or polyacrylic resins, such as Amberchrom CG 71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-linked polyacrylamide resins and the like that are insoluble under the conditions in which they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling chemistries include cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling chemistries. These and other solid media are well known and widely used in the art, and are available from commercial suppliers. Methods for binding receptor polypeptides to support media are well known in the art. Selection of a particular method is a matter of routine design and is determined in part by the properties of the chosen support. See, for example, *Affinity Chromatography: Principles & Methods*, Pharmacia LKB Biotechnology, Uppsala, Sweden, 1988.

[0046] The polypeptides of the present invention can be isolated by exploitation of their physical properties. For example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich proteins including those comprising polyhistidine tags. Briefly, a gel is first charged with divalent metal ions to form a chelate (Sulkowski, *Trends in Biochem.* 3:1-7, 1985). Histidine-rich proteins will be adsorbed to this matrix with differing affinities, depending upon the metal ion used, and will be eluted by competitive elution, lowering the pH, or use of strong chelating agents. Other methods of purification include purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography (Methods in Enzymol., Vol. 182, "Guide to Protein Purification", M. Deutscher, (ed.), Acad. Press, San Diego, 1990, pp. 529-39). Within additional embodiments of the invention, a fusion of the polypeptide of interest and an affinity tag (e.g., maltose-binding protein, FLAG-tag (Asp Tyr Lys Asp Asp Asp Asp Lys (SEQ ID NO:13)), Glu-Glu tag (Glu Glu Tyr Met Pro Met Glu (SEQ ID NO:14)), an immunoglobulin domain) may be constructed to facilitate purification.

[0047] Protein refolding (and optionally reoxidation) procedures may be advantageously used. It is preferred to purify the protein to >80% purity, more preferably to >90% purity, even more preferably >95%, and particularly preferred is a pharmaceutically pure state, that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins and nucleic acids, and free of infectious and pyrogenic agents. Preferably, a purified protein is substantially free of other proteins, particularly other proteins of animal origin.

[0048] BR43x2 polypeptides may also be prepared through chemical synthesis. Exemplary BR43x2 polypeptides include polypeptides of from 32-40 residues in length having an amino acid sequence conforming to the motif: XXCX [QEK][QEKNRDHS][QE]X{0-2}[YFW][YFW]DXLLX{2} C[IMLV]XCX{3}CX{6-8}CX{2}[YF]CXX (SEQ ID NO:10), and subject to the limitations described herein.

[0049] BR43x2 polypeptides can be synthesized by exclusive solid phase synthesis, partial solid phase methods, fragment condensation or classical solution synthesis. The polypeptides are preferably prepared by solid phase peptide synthesis.

[0050] The present invention further provides a variety of other polypeptide fusions and related multimeric proteins comprising one or more polypeptide fusions. A soluble BR43x2, TACI or BCMA polypeptide can be expressed as a fusion with an immunoglobulin heavy chain constant region, typically an F_c fragment, which contains two constant region domains and lacks the variable region. Methods for preparing such fusions are disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Such fusions are typically secreted as multimeric molecules wherein the Fc portions are disulfide bonded to each other and two non-Ig polypeptides are arrayed in close proximity to each other. Immunoglobulin-BR43x2 (TACI or BCMA) polypeptide fusions can be expressed in genetically engineered cells to produce a variety of multimeric BR43x2 analogs. Auxiliary domains can be fused to BR43x2 (TACI or BCMA) polypeptides to target them to specific cells, tissues, or macromolecules. Fusions may also be made using toxins as discussed herein. In this way, polypeptides and proteins can be targeted for therapeutic or diagnostic purposes. A BR43x2 polypeptide can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between domains. See, Tuan et al., *Connect. Tiss. Res.* 34:1-9, 1996.

[0051] The invention also provides soluble BR43x2 receptors and polypeptide fragments used to form fusion proteins with affinity tags or labels. Soluble BR43x2-affinity tag fusion proteins are used, for example, to identify the BR43x2 ligands, as well as agonists and antagonists of the natural ligand. Using labeled, soluble BR43x2, cells expressing the ligand, agonists or antagonists are identified by fluorescence immunocytometry or immunohistochemistry. The soluble fusion proteins are useful in studying the distribution of the ligand on tissues or specific cell lineages, and to provide

insight into receptor/ligand biology.

[0052] To purify ligand, agonists or antagonists, a BR43x2-Ig fusion protein is added to a sample containing the ligand, agonist or antagonist under conditions that facilitate receptor-ligand binding (typically near-physiological temperature, pH, and ionic strength). The receptor-ligand complex is then separated by the mixture using protein A, which is immobilized on a solid support (e.g., insoluble resin beads). The ligand, agonist, antagonist is then eluted using conventional chemical techniques, such as with a salt or pH gradient. In the alternative, the fusion protein itself can be bound to a solid support, with binding and elution carried out as above,

[0053] The resulting media will generally be configured in the form of a column, and fluids containing ligand are passed through the column one or more times to allow ligand to bind to the receptor polypeptide. The ligand is then eluted using changes in salt concentration, chaotropic agents ($MnCl_2$), or pH to disrupt ligand-receptor binding.

[0054] To direct the export of the soluble receptor from the host cell, the soluble receptor DNA is linked to a second DNA segment encoding a secretory peptide, such as a t-PA secretory peptide. To facilitate purification of the secreted receptor domain, an N- or C-terminal extension, such as an affinity tag or another polypeptide or protein for which an antibody or other specific binding agent is available, can be fused to the receptor polypeptide.

[0055] Scatchard plot analysis for soluble ^{125}I -ztnf4 binding to TACI and BCMA is shown in Figure 2 and compared with the binding constants of other members of the TNFR family in Table 7.

Table 7

Ligand	Kd M	Cell source	Reference
TNF _a high	7.14E-11	HL-60	a
TNF _a low	3.26E-10	HEP-2	a
TNF _a high	2.00E-10	HL-60	b
CD27L	3.70E-10	MP-1	c
CD27L	8.30E-09	MP-1	c
CD40L	5.00E-10	EL40.5	d
CD40L	1.00E-09	EBNA	d
(125I-CD40)			
4-1BBL	1.16E-09	Biacore	e
anti 41BBmab	4.14E-10	Biacore	e
ztnf4 sol.	1.11E-09	TACI-BHK	
ztnf4 sol.	1.25E-09	BCMA-BHK	

a Hohmann et al., *J. Biol. Chem.* 264:14927-34, 1989

b Manna and Aggarwal, *J. Biol. Chem.* 273:33333-41, 1998

c Goodwin et al., *Cell* 73:447-56, 1993

d Armitage et al., *Nature* 357:80-82, 1992

e Shuford et al., *J. Exp. Med.* 186:47-55, 1997

[0056] Ztnf4 (5 ng/ml) was found to bind to BR43x2 (SEQ ID NO:2), TACI (SEQ ID NO:6), BCMA (SEQ ID NO:8) and BR43x1 (SEQ ID NO:9), by FACS analysis (Flow Cytometry and Sorting, Melamed et al. eds. Wiley-Liss, 1990 and Immunofluorescence and Cell Sorting, Current Protocols in Immunology, Volume 1, Coligan et al. eds. John Wiley & Son, 1997). FITC-tagged, soluble ztnf4 was also shown to bind specifically to, among other things, B lymphocytes in PBMNCs, tonsil cells, to B cell lymphoma cell lines (Raji, Burkitt's human lymphoma, ATCC CCL86), Ramos (Burkitt's lymphoma cell line, ATCC CRL-1596), Daudi (Burkitt's human lymphoma, ATCC CCL213) and RPMI 1788 (a B lymphocyte cell line, ATCC CCL-156) using FACS analysis. No binding was seen with HL-60, (ATCC a promyelocytic cell line, ATCC CCL-240). Specificity for binding to B cells from PBMNC and tonsil cells was confirmed by co-staining with

antibodies to B cell specific molecules including CD19, IgD, IgM, and CD20. Similarity of ztnf4 to CD40L suggested a broader tissue distribution than was seen. Affinity of ztnf4 was tested on monocytes, dendritic cells, and purified T cells using cytokine proliferation and T cell proliferation assays, for example, and could not detect binding of ztnf4 or any other biological effect on any other type of cell tested. Therefore, the specificity for B cells by the ligand and receptor suggests that they are useful for the study and treatment of autoimmunity, B cell cancers, immunomodulation, IBD and any antibody-mediated pathologies, e.g. ITCP, myasthenia gravis and the like, renal diseases, indirect T cell immune response, graft rejection, graft versus host disease.

[0057] Ztnf4 has been shown to activate B cells resulting in B cell proliferation, antibody production and up-regulation of activation markers *in vitro* (see examples below). These affects may require co-stimulation via IL-4 or other cytokines or stimulation through the B cell antigen receptor or other cell surface receptors which activate B cells, i.e., CD40. Other tumor necrosis factor ligands, such as gp39 and TNF β , also stimulate B cell proliferation. Thus the polypeptides of the current invention can be targeted to specifically regulate B cell responses, inhibiting activated B cells, during the immune response without affecting other cell populations which is advantageous in the treatment of disease. Additionally, the polypeptides of the present invention could be used to modulate B cell development, development of other cells, antibody production and cytokine production. BR43x2 polypeptides can also find use in inducing apoptosis and/or anergy within cells. Polypeptides of the present invention could also modulate T and B cell communication by neutralizing the proliferative effects of ztnf4. Bioassays and ELISAs are available to measure cellular response to ztnf4 in the presence of soluble BR43x2, TACI and/or BCMA. Other assays include those which measure changes in cytokine production as a measure of cellular response (see for example, *Current Protocols in Immunology* ed. John E. Coligan et al., NIH, 1996). Assays to measure other cellular responses, including antibody isotype, monocyte activation, NK cell formation, antigen presenting cell function, apoptosis.

[0058] BR43x2 polypeptides of the present invention would be useful in the preparation of a medicament to neutralize the effects of ztnf4 for treating pre-B or B-cell leukemias, such as plasma cell leukemia, chronic or acute lymphocytic leukemia, myelomas such as multiple myeloma, plasma cell myeloma, endothelial myeloma and giant cell myeloma; and lymphomas such as non-Hodgkins lymphoma, for which an increase in ztnf4 polypeptides is associated. Soluble BR43x2 would be a useful component in a medicament for inhibiting tumor progression and survival.

[0059] Northern blot analysis showed ztnf4 is expressed in CD8 $^{+}$ cells, monocytes, dendrocytes, activated monocytes. This suggests that in some autoimmune disorders, cytotoxic T-cells might stimulate B-cell production through excess production of ztnf4. Immunosuppressant proteins that selectively block the action of B-lymphocytes would be of use in treating disease. Autoantibody production is common to several autoimmune diseases and contributes to tissue destruction and exacerbation of disease. Autoantibodies can also lead to the occurrence of immune complex deposition complications and lead to many symptoms of systemic lupus erythematosis, including kidney failure, neuralgic symptoms and death. Modulating antibody production independent of cellular response would also be beneficial in many disease states. B cells have also been shown to play a role in the secretion of arthritogenic immunoglobulins in rheumatoid arthritis, (Korganow et al., *Immunity* 10:451-61, 1999). As such, inhibition of ztnf4 antibody production would be beneficial in treatment of autoimmune diseases such as myasthenia gravis and rheumatoid arthritis. Immunosuppressant therapeutics such as soluble BR43x2 that selectively block or neutralize the action of B-lymphocytes would be useful for such purposes. To verify these capabilities in BR43x2 soluble receptor polypeptides of the present invention, such BR43x2 polypeptides are evaluated using assays known in the art and described herein.

[0060] The invention provides methods employing BR43x2, TACI or BCMA polypeptides, fusions, antibodies, agonists or antagonists for selectively blocking or neutralizing the actions of B-cells in association with end stage renal diseases, which may or may not be associated with autoimmune diseases. Such methods would also be useful for treating immunologic renal diseases. Such methods would be useful for treating glomerulonephritis associated with diseases such as membranous nephropathy, IgA nephropathy or Berger's Disease, IgM nephropathy, Goodpasture's Disease, post-infectious glomerulonephritis, mesangiproliferative disease, minimal-change nephrotic syndrome. Such methods would also serve as therapeutic applications for treating secondary glomerulonephritis or vasculitis associated with such diseases as lupus, polyarteritis, Henoch-Schonlein, Scleroderma, HIV-related diseases, amyloidosis or hemolytic uremic syndrome. The methods of the present invention would also be useful as part of a therapeutic application for treating interstitial nephritis or pyelonephritis associated with chronic pyelonephritis, analgesic abuse, nephrocalcinosis, nephropathy caused by other agents, nephrolithiasis, or chronic or acute interstitial nephritis.

[0061] The methods of the present invention also include use of BR43x2, TACI or BCMA polypeptides, fusions, antibodies, agonists or antagonists in the treatment of hypertensive or large vessel diseases, including renal artery stenosis or occlusion and cholesterol emboli or renal emboli.

[0062] The present invention also provides methods for diagnosis and treatment of renal or urological neoplasms, multiple myelomas, lymphomas, light chain neuropathy or amyloidosis.

[0063] The invention also provides methods for blocking or inhibiting activated B cells using BR43x2, TACI, or BCMA polypeptides, fusions, antibodies, agonists or antagonists for the treatment of asthma and other chronic airway diseases

such as bronchitis and emphysema.

[0064] Also provided are methods for inhibiting or neutralizing an effector T cell response using BR43x2, TACI, or BCMA polypeptides, fusions, antibodies, agonists or antagonists for use in immunosuppression, in particular for such therapeutic use as for graft-versus-host disease and graft rejection. Additional use would be found in regulation of the immune response, in particular the activation and regulation of lymphocytes. BR43x2, TACT, or BCMA polypeptides, fusions, antibodies, agonists or antagonists would be useful in therapies for treating immunodeficiencies. BR43x2, TACI, or BCMA polypeptides, fusions, antibodies, agonists or antagonists would be useful in therapeutic protocols for treatment of such autoimmune diseases as insulin dependent diabetes mellitus (IDDM) and Crohn's Disease. Methods of the present invention would have additional therapeutic value for treating chronic inflammatory diseases, in particular to lessen joint pain, swelling, anemia and other associated symptoms as well as treating septic shock.

[0065] The effect of soluble BR43x2, TACI, or BCMA polypeptides and fusion proteins on immune response can be measured by administering the polypeptides of the present invention to animals immunized with antigen followed by injection of ztnf4 and measuring antibody isotype production and B and T cell responses including delayed type hypersensitivity and *in vitro* proliferation and cytokine production according the methods known in the art.

[0066] The present invention therefore provides use of a compound in the manufacture of a medicament for inhibiting ztnf4 activity on a mammal, wherein the compound is selected from the group consisting of: a) a polypeptide of SEQ ID NO:4; b) a polypeptide of SEQ ID NO:8; c) a fusion protein; d) a polypeptide of SEQ ID NO:6 from amino acid residue 1 to residue 166; e) a polypeptide of SEQ ID NO:8 from amino acid residue 1 to residue 150; f) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:4; and g) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:10. Examples of fusion proteins include fusions of soluble BR43x2 (SEQ ID NO:4), TACI (from amino acid residue 1 to residue 166 of SEQ ID NO:6) or BCMA (from amino acid residue 1 to residue 150 of SEQ ID NO:8) with another polypeptide, preferably an immunoglobulin heavy chain constant region Fc fragment. The invention similarly provides a method to prepare a medicament for inhibiting BR43x2, TACI or BCMA receptor-ligand engagement.

[0067] Such methods would be particularly useful where ztnf4 activity is associated with activated B lymphocytes and for treating pre-B cell or B-cell cancers. Such methods would also be useful where ztnf4 activity is associated with antibody production. In particular, antibody production associated with autoimmune diseases such as systemic lupus erythematosus, myasthenia gravis or rheumatoid arthritis.

[0068] The adenovirus system can also be used for protein production *in vitro*. By culturing adenovirus-infected non-293 cells under conditions where the cells are not rapidly dividing, the cells can produce proteins for extended periods of time. For instance, BHK cells are grown to confluence in cell factories, then exposed to the adenoviral vector encoding the secreted protein of interest. The cells are then grown under serum-free conditions, which allows infected cells to survive for several weeks without significant cell division. Alternatively, adenovirus vector infected 293S cells can be grown in suspension culture at relatively high cell density to produce significant amounts of protein (see Garnier et al., *Cytotechnol.* 15:145-55, 1994). With either protocol, an expressed, secreted heterologous protein can be repeatedly isolated from the cell culture supernatant. Within the infected 293S cell production protocol, non-secreted proteins may also be effectively obtained.

[0069] Well established animal models are available to test *in vivo* efficacy of soluble BR43x2, TACI, or BCMA polypeptides of the present invention in certain disease states.

[0070] Immune response in animals subjected to a regular antigen challenge (for example, ovalbumin or collagen) followed by administration of BR43x2, TACI or BCMA polypeptides or soluble Ig-fusions can be done to measure effect on B cell response.

[0071] Pharmacokinetic studies can be used in association with radiolabeled, soluble BR43x2, TACI or BCMA polypeptides or fusions to determine the distribution and half life of such polypeptides *in vivo*. Additionally animal models can be used to determine the effects of soluble BR43x2, TACI or BCMA on tumors and tumor development *in vivo*.

[0072] Also provided is the use of BR43x2, TACI or BCMA polypeptides as surrogate markers for autoimmune diseases, kidney diseases, B and T cell diseases. Such patients can be bled and BR43x2, TACI or BCMA soluble receptors and their ligands can be detected in the blood.

[0073] The invention also provides uses of antibody in the preparation of medicaments. Antibodies to BR43x2 or peptides having an amino acid sequence of SEQ ID NO:8, can be obtained, for example, using as an antigen the product of an expression vector containing the polypeptide of interest, or a polypeptide isolated from a natural source. Particularly useful antibodies "bind specifically" with BR43x2 or peptides having an amino acid sequence of SEQ ID NO:10. Antibodies are considered to be specifically binding if the antibodies bind to a BR43x2 polypeptide or a polypeptide of SEQ ID NO:8, peptide or epitope with a binding affinity (K_a) of $10^6 M^{-1}$ or greater, preferably $10^7 M^{-1}$ or greater, more preferably $10^8 M^{-1}$ or greater, and most preferably $10^9 M^{-1}$ or greater. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, *Ann. NY Acad. Sci.* 51:660, 1949). Suitable antibodies include antibodies that bind with BR43x2, in particular the extracellular domain of BR43x2 (amino acid residues 1-120 of SEQ ID NO:2) and those that bind with polypeptides having an amino acid

sequence of SEQ ID NO:10.

[0074] Anti-BR43x2 antibodies can be produced using antigenic BR43x2 epitope-bearing peptides and polypeptides. Antigenic epitope-bearing peptides and polypeptides of the present invention contain a sequence of at least nine, preferably between 15 to about 30 amino acids contained within SEQ ID NO:2. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of the invention, containing from 30 to 50 amino acids, or any length up to and including the entire amino acid sequence of a polypeptide of the invention, also are useful for inducing antibodies that bind with BR43x2. It is desirable that the amino acid sequence of the epitope-bearing peptide is selected to provide substantial solubility in aqueous solvents (*i.e.*, the sequence includes relatively hydrophilic residues, while hydrophobic residues are preferably avoided). Hydrophilic peptides can be predicted by one of skill in the art from a hydrophobicity plot, see for example, Hopp and Woods (*Proc. Nat. Acad. Sci. USA* 78:3824-8; 1981) and Kyte and Doolittle (*J. Mol. Biol.* 157: 105-142, 1982). Moreover, amino acid sequences containing proline residues may be also be desirable for antibody production.

[0075] Polyclonal antibodies to recombinant BR43x2 protein or to BR43x2 isolated from natural sources can be prepared using methods well-known to those of skill in the art. See, for example, Green et al., "Production of Polyclonal Antisera," in *Immunochemical Protocols* (Manson, ed.), pages 1-5 (Humana Press 1992), and Williams et al., "Expression of foreign proteins in *E. coli* using plasmid vectors and purification of specific polyclonal antibodies," in *DNA Cloning 2: Expression Systems*, 2nd Edition, Glover et al. (eds.), page 15 (Oxford University Press 1995). The immunogenicity of a BR43x2 polypeptide can be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of BR43x2 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like," such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

[0076] Although polyclonal antibodies are typically raised in animals such as horses, cows, dogs, chicken, rats, mice, rabbits, hamsters, guinea pigs, goats or sheep, an anti-BR43x2 antibody of the present invention may also be derived from a subhuman primate antibody. General techniques for raising diagnostically and therapeutically useful antibodies in baboons may be found, for example, in Goldenberg et al., international patent publication No. WO 91/11465, and in Losman et al., *Int. J. Cancer* 46:310, 1990. Antibodies can also be raised in transgenic animals such as transgenic sheep, cows, goats or pigs, and may be expressed in yeast and fungi in modified forms as will as in mammalian and insect cells.

[0077] Alternatively, monoclonal anti-BR43x2 antibodies can be generated. Rodent monoclonal antibodies to specific antigens may be obtained by methods known to those skilled in the art (see, for example, Kohler et al., *Nature* 256: 495, 1975, Coligan et al. (eds.), *Current Protocols in Immunology*, Vol. 1, pages 2.5.1-2.6.7 (John Wiley & Sons 1991), Picksley et al., "Production of monoclonal antibodies against proteins expressed in *E. coli*," in *DNA Cloning 2: Expression Systems*, 2nd Edition, Glover et al. (eds.), page 93 (Oxford University Press 1995)).

[0078] Briefly, monoclonal antibodies can be obtained by injecting mice with a composition comprising a BR43x2 gene product, verifying the presence of antibody production by removing a serum sample, removing the spleen to obtain B-lymphocytes, fusing the B-lymphocytes with myeloma cells to produce hybridomas, cloning the hybridomas, selecting positive clones which produce antibodies to the antigen, culturing the clones that produce antibodies to the antigen, and isolating the antibodies from the hybridoma cultures.

[0079] In addition, an anti-BR43x2 antibody of the present invention may be derived from a human monoclonal antibody. Human monoclonal antibodies are obtained from transgenic mice that have been engineered to produce specific human antibodies in response to antigenic challenge. In this technique, elements of the human heavy and light chain locus are introduced into strains of mice derived from embryonic stem cell lines that contain targeted disruptions of the endogenous heavy chain and light chain loci. The transgenic mice can synthesize human antibodies specific for human antigens, and the mice can be used to produce human antibody-secreting hybridomas. Methods for obtaining human antibodies from transgenic mice are described, for example, by Green et al., *Nat. Genet.* 7:13, 1994, Lonberg et al., *Nature* 368:856, 1994, and Taylor et al., *Int. Immun.* 6:579, 1994.

[0080] Monoclonal antibodies can be isolated and purified from hybridoma cultures by a variety of well-established techniques. Such isolation techniques include affinity chromatography with Protein-A Sepharose, size-exclusion chromatography, and ion-exchange chromatography (see, for example, Coligan at pages 2.7.1-2.7.12 and pages 2.9.1-2.9.3; Baines et al., "Purification of Immunoglobulin G (IgG)," in *Methods in Molecular Biology*, Vol. 10, pages 79-104 (The Humana Press, Inc. 1992)).

[0081] For particular uses, it may be desirable to prepare fragments of anti-BR43x2 antibodies. Such antibody fragments can be obtained, for example, by proteolytic hydrolysis of the antibody. Antibody fragments can be obtained by pepsin or papain digestion of whole antibodies by conventional methods. As an illustration, antibody fragments can be produced by enzymatic cleavage of antibodies with pepsin to provide a 5S fragment denoted F(ab')₂. This fragment can be further cleaved using a thiol reducing agent to produce 3.5S Fab' monovalent fragments. Optionally, the cleavage

reaction can be performed using a blocking group for the sulphydryl groups that result from cleavage of disulfide linkages. As an alternative, an enzymatic cleavage using pepsin produces two monovalent Fab fragments and an Fc fragment directly. These methods are described, for example, by Goldenberg, U.S. patent No. 4,331,647, Nisonoff et al., Arch Biochem. Biophys. 89:230, 1960, Porter, Biochem. J. 73:119, 1959, Edelman et al., in Methods in Enzymology Vol. 1, page 422 (Academic Press 1967), and by Coligan, ibid.

[0082] Other methods of cleaving antibodies, such as separation of heavy chains to form monovalent light-heavy chain fragments, further cleavage of fragments, or other enzymatic, chemical or genetic techniques may also be used, so long as the fragments bind to the antigen that is recognized by the intact antibody.

[0083] For example, Fv fragments comprise an association of V_H and V_L chains. This association can be noncovalent, as described by Inbar et al., Proc. Natl. Acad. Sci. USA 69:2659, 1972. Alternatively, the variable chains can be linked by an intermolecular disulfide bond or cross-linked by chemicals such as gluteraldehyde (see, for example, Sandhu, Crit. Rev. Biotech. 12:437, 1992).

[0084] The Fv fragments may comprise V_H and V_L chains which are connected by a peptide linker. These single-chain antigen binding proteins (scFv) are prepared by constructing a structural gene comprising DNA sequences encoding the V_H and V_L domains which are connected by an oligonucleotide. The structural gene is inserted into an expression vector which is subsequently introduced into a host cell, such as *E. coli*. The recombinant host cells synthesize a single polypeptide chain with a linker peptide bridging the two V domains. Methods for producing scFvs are described, for example, by Whitlow et al., Methods: A Companion to Methods in Enzymology 2:97, 1991, also see, Bird et al., Science 242:423, 1988, Ladner et al., U.S. Patent No. 4,946,778, Pack et al., Bio/Technology 11:1271, 1993, and Sandhu, ibid.

[0085] As an illustration, a scFv can be obtained by exposing lymphocytes to BR43x2 polypeptide *in vitro*, and selecting antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled BR43x2 protein or peptide). Genes encoding polypeptides having potential BR43x2 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known in the art (Ladner et al., U.S. Patent No. 5,223,409, Ladner et al., U.S. Patent No. 4,946,778, Ladner et al., U.S. Patent No. 5,403,484, Ladner et al., U.S. Patent No. 5,571,698, and Kay et al., Phage Display of Peptides and Proteins (Academic Press, Inc. 1996)) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA), and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the BR43x2 sequences disclosed herein to identify proteins which bind to BR43x2.

[0086] Another form of an antibody fragment is a peptide coding for a single complementarity-determining region (CDR). CDR peptides ("minimal recognition units") can be obtained by constructing genes encoding the CDR of an antibody of interest. Such genes are prepared, for example, by using the polymerase chain reaction to synthesize the variable region from RNA of antibody-producing cells (see, for example, Larrick et al., Methods: A Companion to Methods in Enzymology 2:106, 1991), Courtenay-Luck, "Genetic Manipulation of Monoclonal Antibodies," in Monoclonal Antibodies: Production, Engineering and Clinical Application, Ritter et al. (eds.), page 166 (Cambridge University Press 1995), and Ward et al., "Genetic Manipulation and Expression of Antibodies," in Monoclonal Antibodies: Principles and Applications, Birch et al., (eds.), page 137 (Wiley-Liss, Inc. 1995)).

[0087] Alternatively, an anti-BR43x2 antibody may be derived from a "humanized" monoclonal antibody. Humanized monoclonal antibodies are produced by transferring mouse complementary determining regions from heavy and light variable chains of the mouse immunoglobulin into a human variable domain. Typical residues of human antibodies are then substituted in the framework regions of the murine counterparts. The use of antibody components derived from humanized monoclonal antibodies obviates potential problems associated with the immunogenicity of murine constant regions. General techniques for cloning murine immunoglobulin variable domains are described, for example, by Orlandi et al., Proc. Natl. Acad. Sci. USA 86:3833, 1989. Techniques for producing humanized monoclonal antibodies are described, for example, by Jones et al., Nature 321:522, 1986, Carter et al., Proc. Natl. Acad. Sci. USA 89:4285, 1992, Sandhu, Crit. Rev. Biotech. 12:437, 1992, Singer et al., J. Immun. 150:2844, 1993, Sudhir (ed.), Antibody Engineering Protocols (Humana Press, Inc. 1995), Kelley, "Engineering Therapeutic Antibodies," in Protein Engineering: Principles and Practice, Cleland et al. (eds.), pages 399-434 (John Wiley & Sons, Inc. 1996), and by Queen et al., U. S. Patent No. 5,693,762 (1997).xxx

[0088] Polyclonal anti-idiotype antibodies can be prepared by immunizing animals with anti-BR43x2 antibodies or antibody fragments, using standard techniques. See, for example, Green et al., "Production of Polyclonal Antisera," in Methods In Molecular Biology: Immunochemical Protocols, Manson (ed.), pages 1-12 (Humana Press 1992). Also,

see Coligan, *ibid.* at pages 2.4.1-2.4.7. Alternatively, monoclonal anti-idiotype antibodies can be prepared using anti-BR43x2 antibodies or antibody fragments as immunogens with the techniques, described above. As another alternative, humanized anti-idiotype antibodies or subhuman primate anti-idiotype antibodies can be prepared using the above-described techniques. Methods for producing anti-idiotype antibodies are described, for example, by Irie, U.S. Patent No. 5,208,146, Greene, et al., U.S. Patent No. 5,637,677, and Varthakavi and Minocha, *J. Gen. Virol.* 77:1875, 1996.

[0089] Antibodies or polypeptides herein can also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. For instance, polypeptides or antibodies of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule (receptor or antigen, respectively, for instance). More specifically, BR43x2 polypeptides or anti-BR43x2 antibodies, or bioactive fragments or portions thereof, can be coupled to detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule.

[0090] Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/anti-complementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/anticomplementary pair.

[0091] Soluble BR43x2 polypeptides or antibodies to BR43x2 can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for the preparation of therapeutic compositions for use in *vivo*. For instance, polypeptides or antibodies of the present invention can be used to treat tissues or organs that express a corresponding anti-complementary molecule (receptor or antigen, respectively, for instance). More specifically, BR43x2 polypeptides or anti-BR43x2 antibodies, or bioactive fragments or portions thereof, can be coupled to cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule.

[0092] Suitable cytotoxic molecules can be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or antibodies can also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a cytotoxic molecule, the cytotoxic molecule can be conjugated with a member of a complementary/anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/anticomplementary pair.

[0093] Such polypeptide-toxin fusion proteins or antibody/fragment-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein including only the targeting domain can be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of interest. In instances where the domain only fusion protein includes a complementary molecule, the anti-complementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle for cell/tissue-specific delivery of generic anti-complementary/cytotoxic molecule conjugates. The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously, intraarterially or intraductally, or may be introduced locally at the intended site of action.

[0094] Antibodies can be made to soluble, BR43x2 polypeptides which are His or FLAG™ tagged. Antibodies can also be prepared to *E. coli* produced MBP-fusion proteins. Alternatively, such polypeptides could include a fusion protein with Human Ig. In particular, antiserum containing polypeptide antibodies to His-tagged, or FLAG™-tagged soluble BR43x2 can be used in analysis of tissue distribution of BR43x2 by immunohistochemistry on human or primate tissue. These soluble BR43x2 polypeptides can also be used to immunize mice in order to produce monoclonal antibodies to a soluble human BR43x2 polypeptide. Monoclonal antibodies to a soluble human BR43x2 polypeptide can also be used to mimic ligand/receptor coupling, resulting in activation or inactivation of the ligand/receptor pair. For instance, it has been demonstrated that cross-linking anti-soluble CD40 monoclonal antibodies provides a stimulatory signal to B cells that have been sub-optimally activated with anti-IgM or LPS, and results in proliferation and immunoglobulin production. These same monoclonal antibodies act as antagonists when used in solution by blocking activation of the receptor. Monoclonal antibodies to BR43x2 can be used to determine the distribution, regulation and biological interaction of the BR43x2/BR43x2-ligand pair on specific cell lineages identified by tissue distribution studies.

[0095] Pharmaceutically effective amounts of BR43x2, TACI, or BCMA polypeptides of the present invention can be formulated with pharmaceutically acceptable carriers for parenteral, oral, nasal, rectal, topical, transdermal administration or the like, according to conventional methods. Formulations may further include one or more diluents, fillers, emulsifiers, preservatives, buffers, excipients, and the like, and may be provided in such forms as liquids, powders, emulsions, suppositories, liposomes, transdermal patches and tablets, for example. Slow or extended-release delivery systems, including any of a number of biopolymers (biological-based systems), systems employing liposomes, and polymeric delivery systems, can also be utilized with the compositions described herein to provide a continuous or long-term source of the BR43x2 polypeptide or antagonist. Such slow release systems are applicable to formulations, for example, for oral, topical and parenteral use. The term "pharmaceutically acceptable carrier" refers to a carrier medium which does not interfere with the effectiveness of the biological activity of the active ingredients and which is not toxic to the host or patient. One skilled in the art may formulate the compounds of the present invention in an appropriate manner, and in accordance with accepted practices, such as those disclosed in Remington: The Science and Practice of Pharmacy, Gennaro, ed., Mack Publishing Co., Easton PA, 19th ed., 1995.

[0096] As used herein a "pharmaceutically effective amount" of a BR43x2, TACI, or BCMA polypeptide, agonists or antagonist is an amount sufficient to induce a desired biological result. The result can be alleviation of the signs, symptoms, or causes of a disease, or any other desired alteration of a biological system. For example, an effective amount of a BR43x2, TACI, or BCMA polypeptide is that which provides either subjective relief of symptoms or an objectively identifiable improvement as noted by the clinician or other qualified observer. For example, such an effective amount of a BR43x2, TACI, or BCMA polypeptide or soluble fusion would provide a decrease in B cell response during the immune response, inhibition or decrease in autoantibody production, inhibition of diminution of symptoms associated with SLE, MG or RA. Effective amounts of BR43x2, TACI, or BCMA will decrease the percentage of B cells in peripheral blood. Effective amounts of the BR43x2, TACI, or BCMA polypeptides can vary widely depending on the disease or symptom to be treated. The amount of the polypeptide to be administered and its concentration in the formulations, depends upon the vehicle selected, route of administration, the potency of the particular polypeptide, the clinical condition of the patient, the side effects and the stability of the compound in the formulation. Thus, the clinician will employ the appropriate preparation containing the appropriate concentration in the formulation, as well as the amount of formulation administered, depending upon clinical experience with the patient in question or with similar patients. Such amounts will depend, in part, on the particular condition to be treated, age, weight, and general health of the patient, and other factors evident to those skilled in the art. Typically a dose will be in the range of 0.1-100 mg/kg of subject. Doses for specific compounds may be determined from *in vitro* or *ex vivo* studies in combination with studies on experimental animals. Concentrations of compounds found to be effective *in vitro* or *ex vivo* provide guidance for animal studies, wherein doses are calculated to provide similar concentrations at the site of action.

[0097] The invention is further illustrated by the following examples.

EXAMPLES

Example 1

Identification of BR43x2

[0098] The TACI isoform was cloned from RPMI array library using secretion trap approach. An RPMI 1788 (activated B-cell line) library was arrayed using twenty 96-well plates. Each well contained about 100 *E. coli* colonies, with each colony containing one cDNA clone. DNA minipreps were prepared in 96-well format using the TomTech Quadra 9600. The isolated DNA was then pooled into 120 pools which represent 1600 clones each. These pools were transfected into Cos-7 cells and plated into 12-well plates. Three microliters of pool DNA and 5 µl LipofectAMINE were mixed in 92 µl serum-free DMEM media (55 mg sodium pyruvate, 146 mg L-glutamine, 5 mg transferrin, 2.5 mg insulin, 1 µg selenium and 5 mg fetuin in 500 ml DMEM), incubated at room temperature for 30 minutes, followed by addition of 400 µl serum-free DMEM media. The DNA-LipofectAMINE mix was added onto 220,000 Cos-7 cells/well plated on 12-well tissue culture plates and incubated for 5 hours at 37°C. Following incubation, 500 µl of 20% FBS DMEM media (100 ml FBS, 55 mg sodium pyruvate and 146 mg L-glutamine in 500 ml DMEM) was added to each well and the cells were incubated overnight.

[0099] The secretion trap screen was performed using biotinylated, FLAG-tagged ztnf4. The cells were rinsed with PBS and fixed for 15 minutes with 1.8% formaldehyde in PBS. The cells were then washed with TNT (0.1 M Tris-HCl, 0.15 M NaCl, and 0.05% Tween-20 in H₂O). Cells were permeated with 0.1% Triton-X in PBS for 15 minutes followed by a wash in TNT. The cells were blocked for 1 hour with TNB (0.1 M Tris-HCl, 0.15 M NaCl and 0.5% Blocking Reagent) using a NEN Renaissance® TSA-Direct Kit (NEN, Boston, MA) according the manufacturer's instruction. The cells were washed with TNT and blocked for 15 minutes with avidin and then biotin (Vector Labs Cat# SP-2001) washing in-between with TNT. The cells were incubated for 1 hour with 1 µg/ml ztnf4/Flag/Biotin in TNB followed by a TNT

wash. The cells were then incubated for one hour with a 1:300 dilution of streptavidin-HRP (NEN) in TNB, and washed with TNT. Hybridizations were detected with fluorescein tyramide reagent diluted 1:50 in dilution buffer (NEN) and incubated for 4.4 minutes and washed with TNT. Cells were preserved with Vectashield Mounting Media (Vector Labs, Burlingame, CA) diluted 1:5 in TNT.

[0100] The cells were visualized by fluorescent microscopy using a FITC filter. Twelve pools were positive for ztnf4 binding. Pool D8 (representing 1600 clones) was broken down and a single clone (D8-1), positive for ztnf4 binding, was isolated. Sequencing analysis revealed clone, D8-1, contained a polypeptide sequence which encoded an isoform of TACI, in which the Phe21-Arg67 first cysteine-rich pseudo repeat of TACI was replaced by a single amino acid residue, tryptophan. This isoform was designated BR43x2, the polynucleotide sequence of which is presented in SEQ ID NO:1.

Example 2

Localization of BR43x1 in Lymphocytes and Monocytes

[0101] Reverse transcriptase PCR was used to localize BR43x1 expression in T and B cells and monocytes. Oligonucleotide primers ZC19980 (SEQ ID NO:15) and ZC19981 (SEQ ID NO:16) were used to screen CD19⁺, CD3⁺ and monocyte cDNA for BR43. The reverse transcriptase reaction was carried out at 94°C for 3 minutes, followed by 30 cycles at 94°C for 30 seconds, 68°C for 2 minutes and 72°C for 1 minute, followed by a 7 minute extension at 72°C. A band of the expected size, 720 bp, was detected in B cells only and not in activated T cells as had been reported for TACT using antibodies (von Bülow and Bram, *ibid.*).

Example 3

B cell Proliferation Assay using the BR43 Ligand Ztnf4

[0102] A vial containing 1 x 10⁸ frozen, apheresed peripheral blood mononuclear cells (PBMCs) was quickly thawed in 37°C water bath and resuspended in 25 ml B cell medium (Iscove's Modified Dulbecco's Medium, 10% heat inactivated fetal bovine serum, 5% L-glutamine, 5% Pen/Strep) in a 50 ml tube. Cells were tested for viability using Trypan Blue (GIBCO BRL, Gaithersburg, MD). Ten milliliters of Ficoll/Hypaque Plus (Pharmacia LKB Biotechnology Inc., Piscataway, NJ) was layered under cell suspension and spun for 30 minutes at 1800 rpm and allowed to stop with the brake off. The interphase layer was then removed and transferred to a fresh 50 ml tube, brought up to a final volume of 40 ml with PBS and spun for 10 minutes at 1200 rpm with the brake on. The viability of the isolated B cells was tested using Trypan Blue. The B cells were resuspended at a final concentration of 1 x 10⁶ cells/ml in B cell medium and plated at 180 µl/well in a 96 well U bottom plate (Falcon, VWR, Seattle, WA).

[0103] To the cells were added one of the following stimulators to bring the final volume to 200 ml/well:

[0104] Soluble, FLAG-tagged ztnf-4sCF or ztnf-4sNF, at 10 fold dilutions from 1 mg-1 ng/ml either alone, with 10 µg/ml anti-IgM (goat anti Human IgM) diluted in NaH₂CO₃, pH 9.5, (Southern Biotechnology Associates, Inc., Birmingham, AL); or with 10 µg/ml anti-IgM, and 10 ng/ml recombinant human IL4 (diluted in PBS and 0.1% BSA). Additionally, other cytokines such as IL-3 and IL-6 as well as a soluble CD40 (sCD40) antibody (Pharmingen, San Diego, CA) were tested as well. As a control the cells incubated with 0.1% bovine serum albumen (BSA) and PBS, 10 µg/ml anti-IgM or 10 µg/ml anti-IgM and 10 ng/ml IL4 (or other cytokines). The cells were then incubated at 37°C in a humidified incubator for 72 hours. Sixteen hours prior to harvesting, 1 µCi ³H thymidine was added to all wells. The cells were harvested into a 96 well filter plate (UniFilter GF/C, Packard, Meriden, CT) where they were harvested using a cell harvester (Packard) and collected according to manufacturer's instructions. The plates were dried at 55°C for 20-30 minutes and the bottom of the wells were sealed with an opaque plate sealer. To each well was added 0.25 ml of scintillation fluid (Microscint-O, Packard) and the plate was read using a TopCount Microplate Scintillation Counter (Packard).

[0105] To measure induction of IgG production in response to various B cell mitogens following stimulation of purified B cells, cells were prepared as described and incubated for 9 days. The cell supernatant was collected to determine IgG production.

[0106] To measure cell surface marker activation in response to various B cell mitogens following stimulation of purified B cells, cells were prepared as described above but incubated only 48 hours. Cell surface markers were measured by FACS analysis.

[0107] Proliferation of human purified B cells stimulated with the various B cell mitogens is summarized in Table 5:

Table 5

Stimulus	Proliferative Index
ztnf4	1.5
ztnf4 + IL4	9.9
ztnf4 + anti-IgM + IL4	15.8

[0108] A synergistic effect of ztnf4 with IL4, IL3 (10 µg/ml) and IL6 (10 µg/ml) was seen on B cell proliferation. A two fold increase in B cell signaling was seen when using sCD40.

[0109] Induction of IgG production (ng/ml) in response to various B cell mitogens following stimulation of purified B cells is summarized in Table 6.

Table 6

Stimulus	Control	Ztnf4
anti-IgM	3	7.5
anti-IgM + IL-4	13	32
anti-IgM + IL-4 + IL-5	10	

[0110] An increase in cell surface activation markers after stimulation of purified B cells with ztnf4 alone, or with anti-IgM or anti-IgM + IL-4 was seen. There was no effect on the proliferation of PBMNCs in the presence of optimal or suboptimal T cell mitogens. Also, no effect on TNF α production was seen in purified monocytes in response to LPS stimulation.

[0111] Figure 3 shows soluble ztnf4 co-activation of human B lymphocytes to proliferate and secrete immunoglobulin. Figure 3A shows purified human peripheral blood B cells proliferation in response to stimulation with soluble ztnf4 (25 ng/ml) in the presence of IL-4 alone, and IL-4 with anti-IgM, anti-CD40, or anti-CD19, after five days in culture. Figure 3B shows the levels of IgM and IgG measured in the supernatants obtained from human B cells stimulated with soluble ztnf4 in the presence of IL-4 or IL-4 + IL-5, after nine days in culture.

[0112] These results suggest that soluble ztnf4 is a B cell activation molecule which acts in concert with other B cell stimuli and weakly by itself. Soluble ztnf4 promotes B cell proliferation and Ig production. The up regulation of adhesion molecules, costimulatory molecules and activation receptors suggests a role for promoting APC function of B cells.

[0113] Figure 4 shows stimulation of human peripheral blood B cells with soluble ztnf4 (25 ng/ml) or a control protein (ubiquitin) in the presence of 10 ng/ml IL-4 for 5 days *in vitro*. Purified TACI-Ig, BCMA-Ig, or control Fc were tested for inhibition of soluble ztnf4 specific proliferation.

Example 4

Selecting TACI and BCMA Transformed BHK Cells using Ztnf4 Binding

[0114] BHK cells expressing a high level of TACI protein were selected by dilution cloning of a transfectant pool. Transfectant cells (2×10^5) were incubated on ice for 30 minutes with biotinylated ztnf4 at 1 µg/ml in binding buffer (PBS, 2% BSA, 0.02% NaN₃). Cells were washed 2X with binding buffer, then incubated with SA-PE (Caltag) (1:1000 dilution in binding buffer) on ice for 30 minutes. Cells were then washed 2X in binding buffer, resuspended in binding buffer, and read by FACS (FACS Vantage, Becton Dickinson). Clones with the highest binding of TNF4 are selected.

[0115] BHK cells expressing a high level of BCMA protein were selected by surface labeling the BCMA-expressing transfectant pool with biotinylated ztnf4. This was followed by streptavidin-Phyco-Erythrin (SA-PE Caltag Burlingame, CA) and sterile sorting for bright cells in FL2 on the FACS Vantage (Becton Dickinson). The single colonies were then screened for ztnf4 binding.

Example 5

Tissue Distribution

[0116] Human Multiple Tissue Northern Blots (MTN I, MTN II and MTN III; Clontech) were probed to determine the tissue distribution of human BR43x2 and TACI expression. An approximately 500 bp PCR derived probe (SEQ ID NO: 21) was amplified using BR43x2 (SEQ ID NO:1) as templates and oligonucleotide ZC20061 (SEQ ID NO:22) and

ZC20062 (SEQ ID NO:23) as primers. This sequence is identical to the homologous region of TACI. The amplification was carried out as follows: 1 cycle at 94°C for 1.0 minutes, 30 cycles of 94°C for 30 seconds, 60°C for 30 seconds and 72°C for 30 seconds, followed by 1 cycle at 72°C for 10 minutes. The PCR products were visualized by agarose gel electrophoresis and the 500 bp PCR product was purified using a Gel Extraction Kit (Qiagen, Chatsworth, CA) according to manufacturer's instructions. The probe was radioactively labeled using the MULTIPRIME DNA labeling kit (Amersham, Arlington Heights, IL) according to the manufacturer's instructions. The probe was purified using a NUCTRAP push column (Stratagene). EXPRESSHYB (Clontech) solution was used for prehybridization and as a hybridizing solution for the Northern blots. Hybridization took place overnight at 65°C using 10⁶ cpm/ml of labeled probe. The blots were then washed in 2X SSC and 0.1% SDS at room temp, followed by 2 washes in 0.1X SSC and 0.1% SDS at 50°C. A transcript of approximately 1.5 kb was detected in spleen, lymph node and small intestine.

[0117] Human Multiple Tissue Northern Blots (MTN I, MTN II and MTN III; Clontech) were probed to determine the tissue distribution of human BCMA expression. An approximately 257 bp PCR derived probe (SEQ ID NO:24) was amplified using Daudi cell cDNA as a template and oligonucleotide ZC21065 (SEQ ID NO:25) and ZC21067 (SEQ ID NO:26) as primers. The amplification was carried out as follows: 1 cycle at 94°C for 1.0 minutes, 35 cycles of 94°C for 30 seconds, 60°C for 30 seconds and 72°C for 30 seconds, followed by 1 cycle at 72°C for 10 minutes. The PCR products were visualized by agarose gel electrophoresis and the 257 bp PCR product was purified using a Gel Extraction Kit (Qiagen, Chatsworth, CA) according to manufacturer's instructions. The probe was radioactively labeled using the MULTIPRIME DNA labeling kit (Amersham, Arlington Heights, IL) according to the manufacturer's instructions. The probe was purified using a NUCTRAP push column (Stratagene). EXPRESSHYB (Clontech) solution was used for prehybridization and as a hybridizing solution for the Northern blots. Hybridization took place overnight at 65°C using 10⁶ cpm/ml of labeled probe. The blots were then washed in 2X SSC and 0.1% SDS at room temp, followed by 2 washes in 0.1X SSC and 0.1% SDS at 50°C. A transcript of approximately 1.2 kb was detected in stomach, small intestine, lymph node, trachea, spleen and testis.

[0118] RNA Master Dot Blots (Clontech) that contained RNAs from various tissues that were normalized to 8 house-keeping genes was also probed with either the TACI probe (SEQ ID NO:21) or the BCMA probe (SEQ ID NO:24) and hybridized as described above. BR43x2/TACI expression was seen in spleen, lymph node, small intestine, stomach, salivary gland, appendix, lung, bone marrow and fetal spleen. BCMA expression was detected in small intestine, spleen, stomach, colon, lymph node and appendix.

[0119] A human Tumor Panel Blot V (Invitrogen Inc., San Diego, CA) and a human lymphoma blot (Invitrogen) were probed as described above either with a Br43x2/TACI probe (SEQ ID NO:21) or a BCMA probe (SEQ ID NO:24). A 1.5 kb transcript corresponding to TACI was found in non-Hodgkin's lymphoma and parotid tumor. A 1.2 kb transcript corresponding to BCMA was found in adenolymphoma, non-Hodgkins lymphoma, and parotid tumor.

[0120] Total RNA from CD4+, CD8+, CD19+ and mixed lymphocyte reaction cells (CellPro, Bothell, WA) was prepared using guanidine isothiocyanate (Chirgwin et al., *Biochemistry* 18:52-94, 1979), followed by a CsCl centrifugation step. Poly(A)+ RNA was isolated using oligo d(T) cellulose chromatography (Aviv and Leder, *Proc. Natl. Acad. Sci. USA*, 69:1408-12, 1972). Northern blot analysis was then performed as follows.

[0121] About 2 mg of each of the poly A+ RNAs was denatured in 2.2 M formaldehyde/phosphate buffer (50 mM Na₂HPO₄, 50 mM NaH₂PO₄, 50 mM NaOAc, 1 mM EDTA and 2.2 M formaldehyde) and separated by 1.5% agarose mini gel (Stratagene Cloning Systems, La Jolla, CA) electrophoresis in formaldehyde/phosphate buffer. The RNA was blotted overnight onto a nytran filter (Schleicher & Schuell, Keene, NH), and the filter was UV crosslinked (1,200 mJoules) in a STRATALINKER^a UV crosslinker (Stratagene Cloning Systems) and then baked at 80°C for 1 hour.

[0122] The blots were probed with either a TACI (SEQ ID NO:21) or BCMA (SEQ ID NO: 24) probe. A 1.5 kb band representing TACI was detected only in CD 19⁺ cells. A 1.2 kb transcript representing BCMA was detected faintly in CD 8⁺, CD 19⁺ and MLR cells.

[0123] Additional Northern Blot analysis was carried out on blots made with poly(A) RNA from K-562 cells (erythroid, ATCC CCL 243), HUT78 cells (T cell, ATCC TIB-161), Jurkat cells (T cell), DAUDI (Burkitt's human lymphoma, Clontech, Palo Alto, CA), RAJI (Burkitt's human lymphoma, Clontech) and HL60 (Monocyte) as described above. The blots were probed with either a TACI (SEQ ID NO:21) or BCMA (SEQ ID NO:24) probe. A transcript of 1.5 kb corresponding to TACI was detected in Raji cells. A transcript of 1.2 kb corresponding to BCMA was detected in Daudi, Raji and Hut 78 cells.

[0124] A PCR-based screen was used to identify tissues which expressed human or murine TACI and human BCMA. Human and Murine Rapid-Scan™ Gene Expression Panels (OriGene Technologies, Inc., Rockville, MD), were screened according to manufacturer's instructions. Oligonucleotide primers ZC24200 (SEQ ID NO:27) and ZC24201 (SEQ ID NO:28) were designed to span an exon junction and produce a 272 bp fragment corresponding to murine TACI. Expression was detected in spleen, thymus, lung, breast, heart, muscle, skin, adrenal gland, stomach, small intestine, brain, ovary, prostate gland and embryo. Additional bands of ~500 and 800bp were detected in many tissues.

[0125] Oligonucleotide primers ZC24198 (SEQ ID NO:29) and ZC24199 (SEQ ID NO:30) were designed to span an exon junction and produce a 204 bp fragment corresponding to human TACI. Expression was detected in spleen, brain,

heart, liver, colon, lung, small intestine, muscle, stomach, testis, placenta, salivary gland, adrenal gland, pancreas, prostate, peripheral blood lymphocytes and bone marrow.

[0126] Oligonucleotide primers ZC24271 (SEQ ID NO:31) and ZC24272 (SEQ ID NO:32) were designed to span an exon junction and produce a 329 bp fragment corresponding to human BCMA. Expression was detected in brain, spleen, colon, lung, small intestine, stomach, ovary, testis, salivary gland, adrenal gland, prostate, peripheral blood lymphocytes, bone marrow and fetal liver.

[0127] Oligonucleotide primers ZC24495 (SEQ ID NO:33) and ZC24496 (SEQ ID NO:34) were designed to span an exon junction and produce a 436 bp fragment corresponding to murine BCMA. Expression was detected in liver.

10 Example 6

Preparation of TACI-Ig and BCMA-Ig Fusion Vectors

Ig Gammal Fc4 Fragment Construction

[0128] To prepare the TACI-Ig fusion protein, the Fc region of human IgG1 (the hinge region and the CH2 and CH3 domains) was modified so as to remove Fc receptor (FcγRI) and complement (C1q) binding functions. This modified version of human IgG1 Fc was called Fc4.

[0129] The Fc region was isolated from a human fetal liver library (Clontech) by PCR using oligo primers ZC10,134 (SEQ ID NO:43) and ZC10,135 (SEQ ID NO:44). PCR was used to introduce mutations within the Fc region to reduce FcγRI binding. The FcγRI binding site (Leu-Leu-gly-Gly) was mutated to Ala-Glu-gly-Ala (amino acid residues 38-41 of SEQ ID NO:45) according to Baum et al. (*EMBO J.* 13:3992-4001, 1994), to reduce FcR1 binding (Duncan et al., *Nature* 332:563-4, 1988). Oligonucleotide primers ZC15,345 (SEQ ID NO:46) and ZC15,347 (SEQ ID NO:47) were used to introduce the mutation. To a 50 µl final volume was added 570 ng IgFc template, 5 µl 10X Pfu reaction Buffer (Stratagene), 8 µl of 1.25 mM dNTPs, 31 µl dH₂O, 2 µl 20 mM ZC15,345 (SEQ ID NO:46) and ZC15,347 (SEQ ID NO:47). An equal volume of mineral oil was added and the reaction was heated to 94°C for 1 minute. Pfu polymerase (2.5 units, Stratagene) was added followed by 25 cycles at 94°C for 30 seconds, 55°C for 30 seconds, 72°C for 1 minute followed by a 7 minute extension at 72°C. The reaction products were electrophoresed and the band corresponding to the predicted size of ~676 bp was detected. The band was excised from the gel and recovered using a QIAGEN QIAquickTM Gel Extraction Kit (Qiagen) according to the manufacturers instructions.

[0130] PCR was also used to introduce a mutation of Ala to Ser (amino acid residue 134 of SEQ ID NO:45) and Pro to Ser (amino acid residue 135 of SEQ ID NO:45) to reduce complement C1q binding and/or complement fixation (Duncan and Winter, *Nature* 332:788, 1988) and the stop codon TAA. Two, first round reactions were done using the FcγRI binding side-mutated IgFc sequence as a template. To a 50 µl final volume was added 1 µl FcγRI binding site mutated IgFc template, 5 µl 10X Rfu Reaction Buffer (Stratagene), 8 µl 1.25 mM dNTPs, 31 µl dH₂O, 2 µl 20 mM ZC15,517 (SEQ ID NO:48), a 5' primer beginning at nucleotide 26 of SEQ ID NO:45 and 2 µl 20 mM ZC15,530 (SEQ ID NO:49), a 3' primer beginning at the complement of nucleotide 405 of SEQ ID NO:45. The second reaction contained 2 µl each of 20 mM stocks of oligonucleotide primers ZC15,518 (SEQ ID NO:50), a 5' primer beginning at nucleotide 388 of SEQ ID NO:45 and ZC15,347 (SEQ ID NO:47), a 3' primer, to introduce the Ala to Ser mutation, Xba I restriction site and stop codon. An equal volume of mineral oil was added and the reactions were heated to 94°C for 1 minute. Pfu polymerase (2.5 units, Stratagene) was added followed by 25 cycles at 94°C for 30 seconds, 55°C for 30 seconds, 72°C for 2 minutes followed by a 7 minute extension at 72°C. The reaction products were electrophoresed and bands corresponding to the predicted sizes, ~370 and ~395 bp respectively, were detected. The bands were excised from the gel and extracted using a QIAGEN QIAquickTM Gel Extraction Kit (Qiagen) according to the manufacturers instructions. A second round reaction was done to join the above fragments and add the 5' Bam HI restriction site. To a 50 µl final volume was added 30 µl dH₂O, 8 µl 1.25 mM dNTPs, 5 µl 10X Pfu polymerase reaction buffer (Stratagene) and 1 µl each of the two first two PCR products. An equal volume of mineral oil was added and the reaction was heated to 94°C for 1 minute. Pfu polymerase (2.5 units, Stratagene) was added followed by 5 cycles at 94°C for 30 seconds, 55 °C for 30 seconds, and 72°C for 2 minutes. The temperature was again brought to 94°C and 2 µl each of 20 mM stocks of ZC15,516 (SEQ ID NO:51), a 5' primer beginning at nucleotide 1 of SEQ ID NO:45, and ZC15,347 (SEQ ID NO:47) were added followed by 25 cycles at 94°C for 30 seconds, 55°C for 30 seconds and 72°C for 2 minutes, and a final 7 minute extension at 72°C. A portion of the reaction was visualized using gel electrophoresis. A 789 bp band corresponding the predicted size was detected.

55 TACI-Fc4 and BCMA-Fc4 Expression Vector Construction

[0131] Expression plasmids containing TACI-Fc4 and BCMA-Fc4 fusion proteins were constructed via homologous recombination in yeast. A fragment of TACI cDNA was isolated using PCR that included the polynucleotide sequence

from nucleotide 15 to nucleotide 475 of SEQ ID NO:5. The two primers used in the production of the TACI fragment were: (1) a primer containing 40 bps of the 5' vector flanking sequence and 17 bps corresponding to the amino terminus of the TACI fragment (SEQ ID NO:52); (2) 40 bps of the 3' end corresponding to the flanking Fc4 sequence and 17 bp corresponding to the carboxyl terminus of the TACI fragment (SEQ ID NO:53). To an 100 μ l final volume was added 5 10 ng TACI template, 10 μ l 10X Taq polymerase Reaction Buffer (Perkin Elmer), 8 μ l 2.5 nM dNTPs, 78 μ l dH₂O, 2 μ l each of 20 mM stocks of oligonucleotide primers SEQ ID NO:52 and SEQ ID NO:53, and taq polymerase (2.5 units, Life Technology). An equal volume of mineral oil was added and the reaction was heated to 94°C for 2 minutes, followed by 25 cycles at 94°C for 30 seconds, 65 °C for 30 seconds, 65°C for 30 seconds, 72°C for 1 minute followed by a 5 minute extension at 72°C.

10 [0132] A fragment of BCMA cDNA was isolated using PCR that includes the polynucleotide sequence from nucleotide 219 to nucleotide 362 of SEQ ID NO:7. The two primers used in the production of the BCMA fragment were an oligonucleotide primer containing 40 bps of the 5' vector flanking sequence and 17 bps corresponding to the amino terminus of the BCMA fragment (SEQ ID NO:54); and an oligonucleotide primer containing 40 bps of the 3' end corresponding to the flanking Fc4 sequence and 17 bps corresponding to the carboxyl terminus of the BCMA fragment (SEQ ID NO:55). To a 100 μ l final volume was added 10 ng BCMA template, 10 μ l 10X Taq polymerase Reaction Buffer (Perkin Elmer), 8 μ l 2.5 mM dNTPs, 78 μ l H₂O, 2 μ l each of 20 mM stock solutions of oligonucleotide primers SEQ ID NO:54 and SEQ ID NO:55. An equal volume of mineral oil was added and the reaction was heated to 94°C for 2 minutes, followed by 25 cycles at 94°C for 30 seconds, 65°C for 30 seconds, 72°C for 1 minute followed by a 5 minute extension at 72°C.

15 [0133] The fragment containing the cDNA encoding the Fc4 fragment was constructed in a similar manner, one for each of the TACI and BCMA fusion constructs. For TACI the two primers used in the production of the Fc4 fragment were (upstream and downstream), an oligonucleotide primer containing 40 bps of the 5' TACI flanking sequence and 17 bps corresponding to the amino terminus of the Fc4 fragment (SEQ ID NO:56); and an oligonucleotide primer containing 40 bps of the 3' end corresponding to the flanking vector sequence and 17 bps corresponding to the carboxyl terminus of the Fc4 fragment (SEQ ID NO:57). For BCMA, the upstream primer in the production of the Fc4 fragment was an oligonucleotide primer containing 40 bps of the 5' BCMA flanking sequence and 17 bps corresponding to the amino terminus of the Fc4 fragment (SEQ ID NO:58). The downstream primer for the Fc4 for the BCMA construct was the same as that described above for TACI-Fc4 (SEQ ID NO:57).

20 [0134] To a 100 μ l final volume was added 10 ng Fc4 template described above, 10 μ l 10X Taq polymerase Reaction Buffer (Perkin Elmer), 8 μ l 2.5 nM dNTPs, 78 μ l dH₂O, 2 μ l each of 20 mM stocks of oligonucleotides SEQ ID NO:56 and SEQ ID NO:57 for TACI and oligonucleotides SEQ ID NO:58 and SEQ ID NO:57 for BCMA, and taq polymerase (2.5 units, Life Technology). An equal volume of mineral oil was added and the reaction was heated to 94°C for 2 minutes, then 25 cycles at 94°C for 30 seconds, 65°C for 30 seconds, 72°C for 1 minute followed by a 5 minute extension at 72°C.

25 [0135] Ten microliters of each of the 100 μ l PCR reactions described above was run on a 0.8% LMP agarose gel (Seaplaque GTG) with 1 x TBE buffer for analysis. The remaining 90 μ l of each PCR reaction was precipitated with the addition of 5 μ l 1 M NaCl and 250 μ l of absolute ethanol. The plasmid pZMP6 was cut with SmaI to linearize it at the polylinker. Plasmid pZMP6 was derived from the plasmid pCZR199 (American Type Culture Collection, Manassas, VA, ATCC# 98668) and is a mammalian expression vector containing an expression cassette having the CMV immediate early promoter, a consensus intron from the variable region of mouse immunoglobulin heavy chain locus, multiple restriction sites for insertion of coding sequences, a stop codon and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene and the SV40 terminator. The vector pZMP6 was constructed from pCZR199 by replacement of the metallothionein promoter with the CMV immediate early promoter, and the Kozac sequences at the 5' end of the open reading frame.

30 [0136] One hundred microliters of competent yeast cells (*S. cerevisiae*) were combined with 10 μ l containing approximately 1 μ g each of either the TACI or the BCMA extracellular domain and the Fc4 PCR fragments appropriate for recombination with each, and 100 ng of SmaI digested pZMP6 vector and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixtures were electropulsed at 0.75 kV (5 kV/cm), \sim ohms, 25 μ F. To each cuvette was added 600 μ l of 1.2 M sorbitol and the yeast were plated in two 300 μ l aliquots onto to URA-D plates and incubated at 30 °C.

35 [0137] After about 48 hours, the Ura+ yeast transformants from a single plate were resuspended in 1 ml H₂O and spun briefly to pellet the yeast cells. The cell pellet was resuspended in 1 ml of lysis buffer (2% Triton X-100, 1% SDS, 100 mM NaCl, 10 mM Tris, pH 8.0, 1 mM EDTA). Five hundred microliters of the lysis mixture was added to an Eppendorf tube containing 300 μ l acid washed glass beads and 200 μ l phenol-chloroform, vortexed for 1 minute intervals two or three times, followed by a 5 minute spin in a Eppendorf centrifuge at maximum speed. Three hundred microliters of the aqueous phase was transferred to a fresh tube, and the DNA precipitated with 600 μ l ethanol (EtOH), followed by centrifugation for 10 minutes at 4°C. The DNA pellet was resuspended in 100 μ l H₂O.

40 [0138] Transformation of electrocompetent *E. coli* cells (DH10B, GibcoBRL) was done with 0.5-2 ml yeast DNA prep

and 40 µl of DH10B cells. The cells were electropulsed at 2.0 kV, 25 mF and 400 ohms. Following electroporation, 1 ml SOC (2% Bacto[®] Tryptone (Difco, Detroit, MI), 0.5% yeast extract (Difco), 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose) was plated in 250 µl aliquots on four LB AMP plates (LB broth (Lennox), 1.8% Bacto[®] Agar (Difco), 100 mg/L Ampicillin).

- 5 [0139] Individual clones harboring the correct expression construct for TACI-Fc4 or BCMA-Fc4 were identified by restriction digest to verify the presence of the insert and to confirm that the various DNA sequences have been joined correctly to one another. The insert of positive clones were subjected to sequence analysis. Larger scale plasmid DNA is isolated using the Qiagen Maxi kit (Qiagen) according to manufacturer's instruction

10 Example 7

Mammalian Expression of TACI-Fc4 and BCMA-Fc4

- 15 [0140] BHK 570 cells (ATCC NO: CRL-10314) were plated in 10 cm tissue culture dishes and allowed to grow to approximately 50 to 70% confluence overnight at 37°C , 5% CO₂, in DMEM/FBS media (DMEM, Gibco/BRL High Glucose, (Gibco BRL, Gaithersburg, MD), 5% fetal bovine serum (Hyclone, Logan, UT), 1 mM L-glutamine (JRH Bio-sciences, Lenexa, KS), 1 mM sodium pyruvate (Gibco BRL)). The cells were then transfected with either the plasmid TACI-Fc4/pZMP6 or BCMA-Fc4/pZMP6, using Lipofectamine™ (Gibco BRL), in serum free (SF) media formulation (DMEM, 10 mg/ml transferrin, 5 mg/ml insulin, 2 mg/ml fetauin, 1% L-glutamine and 1% sodium pyruvate). TA-20 Cl-Fc4/pZMP6 or BCMA-Fc4/pZMP6 was diluted into 15 ml tubes to a total final volume of 640 µl with SF media. 35 µl of Lipofectamine™ (Gibco BRL) was mixed with 605 µl of SF medium. The Lipofectamine™ mix was added to the DNA mix and allowed to incubate approximately 30 minutes at room temperature. Five milliliters of SF media was added to the DNA:Lipofectamine™ mixture. The cells were rinsed once with 5 ml of SF media, aspirated, and the DNA:Lipofectamine™ mixture is added. The cells were incubated at 37°C for five hours, then 6.4 ml of DMEM/10% FBS, 25 1% PSN media was added to each plate. The plates were incubated at 37°C overnight and the DNA:Lipofectamine™ mixture was replaced with fresh 5% FBS/DMEM media the next day. On day 5 post-transfection, the cells were split into T-162 flask in selection medium (DMEM/ 5% FBS, 1% L-GLU, 1% NaPyr). Approximately 10 days post-transfection, two 150 mm culture dishes of methotrexate resistant colonies from each transfection were trypsinized and the cells are pooled and plated into a T-162 flask and transferred to large scale culture.

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Example 9

Transgenic Expression of Ztnf4

- 35 [0141] Transgenic animals expressing ztnf4 genes were made using adult, fertile males (B6C3f1), prepubescent fertile females (B6C3f1), vasectomized males (B6D2f1), and adult fertile females (B6D2f1) (all from Taconic Farms, Germantown, NY). The prepubescent fertile females were superovulated using Pregnant Mare's Serum gonadotrophin (Sigma, St. Louis, MO) and human Chorionic Gonadotropin (hCG (Sigma)). The superovulated females were subsequently mated with adult, fertile males, and copulation was confirmed by the presence of vaginal plugs.
- 40 [0142] Fertilized eggs were collected under a surgical scope (Leica MZ12 Stereo Microscope, Leica, Wetzlar, Germany). The eggs were then washed in hyaluronidase and Whitten's W640 medium (Table 8; all reagents available from Sigma Chemical Co.) that has been incubated with 5% CO₂, 5% O₂, and 90% N₂ at 37°C. The eggs were stored in a 37°C/5% CO₂ incubator until microinjection.

45

Table 8

WHITTEN'S 640 MEDIA		
	mgs/200 ml	mgs/500 ml
NaCl	1280	3200
KCl	72	180
KH ₂ PO ₄	32	80
MgSO ₄ ·7H ₂ O	60	150
Glucose	200	500
Ca ²⁺ Lactate	106	265
Benzylpenicillin	15	37.5
Streptomycin SO ₄	10	25

Table 8 (continued)

WHITTEN'S 640 MEDIA		
	mgs/200 ml	mgs/500 ml
NaHCO ₃	380	950
Na Pyruvate	5	12.5
H ₂ O	200 ml	500 ml
500 mM EDTA	100 µl	250 µl
5% Phenol Red	200 µl	500 µl
BSA	600	

[0143] The 858 bp open reading frame encoding full length human TACI ligand Biys (SEQ ID NO:35) was amplified by PCR so as to introduce an optimized initiation codon and flanking 5' *Pmel* and 3' *Ascl* sites using the oligonucleotide primers of SEQ ID NO:36 and SEQ ID NO:37. This *Pmel*/*Ascl* fragment was subcloned into pKFO24, a B and/or T cell-restricted transgenic vector containing the Ig Em enhancer (690bp *NotI*/*Xba*I from pEmSR; (Bodrug et al., *EMBO J.* 13:2124-30, 1994), the Ig V_h promoter (536 bp *HincII*/*Xba*I fragment from pJH1X(-) Hu et al., *J. Exp. Med.* 177: 1681-90, 1993), the SV40 16S intron (171 bp *Xba*I/*HindIII* fragment from pEmSR), a *Pmel*/*Ascl* polylinker, and the human growth hormone gene polyadenylation signal (627 bp *Smal*/*EcoRI* fragment; Seeburg, *DNA* 1:239-49, 1982). The transgene insert was separated from plasmid backbone by *NotI* digestion and agarose gel purification, and fertilized ova from matings of B6C3F1Tac mice described above were microinjected and implanted into pseudopregnant females essentially as previously described (Malik et al., *Molec. Cell. Biol.* 15:2349-58, 1995)

[0144] The recipients were returned to cages in pairs, and allowed 19-21 days gestation. After birth, 19-21 days postpartum was allowed before sexing and weaning, and a 0.5 cm biopsy (used for genotyping) was snipped off the tail with clean scissors.

[0145] Genomic DNA was prepared from the tail snips using a commercially available kit (DNeasy 96 Tissue Kit; Qiagen, Valencia, CA) following the manufacturer's instructions. Genomic DNA was analyzed by PCR using primers designed to the human growth hormone (hGH) 3' UTR portion of the transgenic vector. Primers ZC17251 (SEQ ID NO:38) and ZC17252 (SEQ ID NO:39) amplify a 368-base-pair fragment of hGH. The use of a region unique to the human sequence (identified from an alignment of the human and mouse growth hormone 3' UTR DNA sequences) ensured that the PCR reaction did not amplify the mouse sequence. In addition, primers ZC17156 (SEQ ID NO:40) and ZC17157 (SEQ ID NO:41), which hybridize to vector sequences and amplify the cDNA insert, may be used along with the hGH primers. In these experiments, DNA from animals positive for the transgene generated two bands, a 368-base-pair band corresponding to the hGH 3' UTR fragment and a band of variable size corresponding to the cDNA insert.

[0146] Once animals were confirmed to be transgenic (TG), they are back-crossed into an inbred strain by placing a TG female with a wild-type male, or a TG male with one or two wild-type female(s). As pups were born and weaned, the sexes were separated, and their tails snipped for genotyping.

[0147] To check for expression of a transgene in a live animal, a survival biopsy is performed. Analysis of the mRNA expression level of each transgene was done using an RNA solution hybridization assay or real-time PCR on an ABI Prism 7700 (PE Applied Biosystems, Inc., Foster City, CA) following the manufacturer's instructions.

Cell Preparation and Flow Cytometry

[0148] Founder mice were analyzed at various ages. For flow cytometric (FACS) analysis of lymphoid tissues, bone marrow (BM) cells were isolated from femurs and tibias by careful disruption in phosphate-buffered saline (PBS) using a mortar and pestle. Cells were resuspended, depleted of bone fragments by passive sedimentation, and pelleted at 1000 x g. Splenocytes, thymocytes, or lymph node cells were obtained by crushing intact tissues between glass slides, then resuspending and pelleting the cells as for BM. Cells were resuspended in FACS wash buffer (FACS WB) (Hank's balanced salt solution, 1% BSA, 10mM Hepes, pH 7.4) at a concentration of 20 x 10⁶ cells/ml prior to staining. To stain, 1 x 10⁶ cells were transferred to 5 ml tubes and washed with 1 ml of FACS WB, then pelleted at 1000 x g. Cells were then incubated on ice for 20 minutes in the presence of saturating amounts of the appropriate FITC-, PE- and/or TriColor (TC)-conjugated mAbs in a total volume of 100 ml in FACS WB. Cells were washed with 1.5 ml of WB, pelleted, then resuspended in 400 ml WB and analyzed on a FACSCalibur flow cytometer using CellQuest software (Becton Dickinson, Mountain View, CA). Detectors for forward (FSC) and side (SSC) light scatter were set on a linear scale, whereas logarithmic detectors were used for all three fluorescence channels (FL-1, FL-2, and FL-3).

[0149] Compensation for spectral overlap between FL channels was performed for each experiment using single color stained cell populations. All cells were collected ungated to disk and data were analyzed using CellQuest software. RBC and dead cells were excluded by electronically gating data on the basis of FSC vs. SSC profiles.

Antibodies

[0150] Fluorescein isothiocyanate (FITC)-conjugated anti-CD8 monoclonal antibody (mAb) (clone 53-6.7) and phycoertherin (PE)-conjugated anti-CD4 (clone RM4-5), anti-CD5 (clone 53-7.3), anti-CD19 (clone 1D3), and anti-syndecan (clone 281-2) mAbs were purchased from PharMingen (San Diego, CA). Tricolor(TC)-conjugated anti-CD45R/B220 mAb (clone RA3-6B2) was purchased from Caltag.

[0151] Transgenic mice over expressing ztnf4 in the lymphoid compartment develop increased numbers of peripheral B cells, increased plasma cells and elevated levels of serum immunoglobulin. These transgenic animals have an increased number of B200+ cells in the spleen, lymph nodes and thymus. The increased number of splenic B cells includes both conventional B-2 cells, and the normally rare population of B-1 cells. In general, B-1 cells are largely confined to the peritoneal and other body cavities, produce low affinity self-reactive antibodies, and have often been associated with the development of autoimmune diseases such as systemic lupus erythematosus SLE.

[0152] Older transgenic animals produce autoantibodies, develop proteinurea and sclerotic glomeruli, characteristics of systemic lupus erythematosus.

[0153] Figure 5A shows single cell suspensions of spleen (top panel), mesenteric lymph node (middle panel), and bone marrow (lower panel) prepared as described below, stained with anti-B220-TC and analyzed by flow cytometry. The number of B220+ cells in each tissue was calculated by multiplying the percent B220+ cells by the total number of live (trypan blue excluding) cells counted on a hemocytometer. Each bar represents data from individual ztnf4 transgenic (Tg, shaded bars) or nonTG littermate (open bars) control mice.

[0154] Figure 5B shows cells isolated from ztnf4 TG (right-hand panels) or nonTG littermate (left-hand panels) lymph node (top row), spleen (middle rows), and thymus (bottom row) were stained with mAbs to the molecules indicated (DC5, CD4 and CD8), then analyzed by flow cytometry. Data shown were gated to exclude dead cells and RBCs.

[0155] Figure 5C shows total IgG, IgM, and IgE levels in serum from ztnf4 transgenic mice ranging in age from 6 to 23 weeks old.

[0156] Figure 5D shows the amyloid deposition and thickened mesangium of the glomeruli identified in H&E stained kidney sections from ztnf4 transgenic mice compared to normal glomeruli from control littermates.

[0157] Figure 5E shows an increase in effector T cells in ztnf4 transgenic mice, similar to that reported by Mackay et al. (*J. Exp. Med.* 190:1697-1710, 1999).

[0158] Soluble TACI(BR43x2) or BCMA-Ig fusions are injected (IP, IM or IV) into ztnf4 over expressing transgenic animals. Flow cytometric (FACS) analysis of lymphoid tissues will be used to identify any change in the number of B220+ B cells in the spleen, lymph nodes and thymus.

Example 10**35 Direct Binding ELISA**

[0159] A direct binding ELISA was developed to characterize the ability of either soluble TACI-Ig or soluble BCMA-Ig to bind and inhibit the biological activity of ztnfr4 *in vitro*.

[0160] A 96 well plate was coated with 1 µg/ml Goat-anti-Human Ig (Jackson Labs, Bar Harbor, MA) in ELISA A buffer (0.1 M Na₂HCO₃, pH 9.6, 0.02% NaN₃) and incubated overnight at 4°C. TACI, BCMA, and an unrelated TNF receptor such as ztnfr10 (SEQ ID NO:42) as a control were titrated from 10 µg/ml through 5 fold dilutions to 320 ng/ml plus a zero and co-incubated with 2.5, 0.5, or 0.1 µg/ml biotinylated ztnf4 or ovalbumin as a negative control, and incubated 1 hour at room temperature.

[0161] The co-incubated receptor-biotinylated ligand mixture was then added to the goat-anti-human Ig coated 96 well plates. The plates were then washed (ELISA C, 500 µl Tween 20 (Sigma Chemical Co., St. Louis, Mo.), 200 mg NaN₃, PBS to a final volume of 1 liter) and blocked with Superblock (Pierce, Rockford, IL). The plates were then incubated at 37°C for 2 hours.

[0162] The plates are once again washed with ELISA C followed by the addition of 100 µl/well of neutravidin-HRP at 1:10,000 in ELISA B (5 or 10 µg BSA (Sigma) for 1% or 2% BSA, respectively, 250 µl Tween 20 (Sigma), 100 mg NaN₃, phosphate-buffered saline pH 7.2 (PBS, Sigma) to a final volume of 500 ml. Alternatively, the buffer may be made up as 1% or 2% BSA in ELISA C Buffer). The plates are then developed with OPD for 10 minutes at room temperature and read at 492.

Example 11**55 Biological Activity Assay**

[0163] A biological activity assay was developed to measure soluble TACI-FC inhibition of human B cell the stimu-

lation by soluble ztnf4. B cells were isolated from peripheral blood mononuclear cells (PBMNC) using CD19 magnetic beads and the VarioMacs magnetic separation system (Miltenyi Biotec Auburn, CA) according to the manufacturer's instructions. Purified B cells were mixed with soluble ztnf4 (25 ng/ml) and recombinant human IL-4 (10 ng/ml Pharminogen) and were plated (in triplicate) on to round bottom 96 well plates at 1×10^5 cells per well.

5 [0164] Soluble TACI-FC was diluted from 5 µg/ml to 6 ng/ml and incubated with the B cell for 5 days, pulsing overnight on day 4 with 1 µCi 3 H Thymidine (Amersham) per well. As a control soluble TACI-FC was also incubated with B cells and IL-4 without ztnf4 present.

[0165] Plates were harvested using Packard plate harvester and counted using the Packard reader. The TACI-Ig soluble receptor inhibited the ability of soluble ztnf4 to stimulate B cell proliferation *in vitro* in a dose-dependent manner.

10 A 10-fold molar excess TACI-Ig completely inhibits the proliferation of human B cells in response to soluble ztnf4 in the presence of IL-4.

Example 12

15 ORIGIN Assay

[0166] Levels of ztnf4 in individuals with a disease condition (such as SLE, rheumatoid arthritis for example) relative to normal individuals were determined using an electrochemiluminescence assay. A standard curve prepared from soluble, human ztnf4 at 10 ng/ml, 1 ng/ml, 0.1 ng/ml, 0.01 ng/ml and 0 ng/ml was prepared in ORIGIN buffer (Igen, Gaithersburg, MD). Serum samples were diluted in ORIGIN buffer. The standards and samples were incubated at room temperature for 2 hours with biotinylated rabbit anti-human ztnf4-NF BV antibody diluted to 1 µg/ml in Origin Assay Buffer (IGEN) and rutenylated rabbit anti-human ztnf4-NF BV polyclonal antibody diluted to 1 µg/ml in Origin Assay Buffer (IGEN). Following the incubation the samples were vortexed and 0.4 mg/ml streptavidin Dynabeads (Dynal, Oslo, Norway) were added to each of the standards and samples at 50 µl/tube and incubated for 30 minutes at room temperature. Samples were then vortexed and samples were read on an Origin Analyzer (Igen) according to manufacturer's instructions. The Origin assay is based on electrochemiluminescence and produces a readout in ECL-what is this, how does it work and what does this tell you.

[0167] An elevated level of ztnf4 was detected in the serum samples from both NZBWF1/J, and MRL/Mpj-Fas^{lpr} mice which have progressed to advanced stages of glomerulonephritis and autoimmune disease.

30

Example 13

Soluble TACI-Ig in a Spontaneous Model of SLE

35 [0168] NZBW mice become symptomatic for spontaneous SLE at approximately 7-9 months of age. TACI-Fc was administered to NZBW mice to monitor its suppressive effect on B cells over the 5 week period when, on average, B-cell autoantibody production is thought to be at high levels in NZBW mice.

[0169] One hundred, 8-week old female (NZB x NZW)F₁ mice (Jackson Labs) were divided into 6 groups of 15 mice. Prior to treatment the mice were monitored once a month for urine protein and blood was drawn for CBC and serum 40 banking. Serum will be screened for the presence of autoantibodies. Because proteinuria is the hallmark sign of glomerulonephritis, urine protein levels were monitored by dipstick at regular intervals over the course of the study. Prior to treatment the animals were weighed. Dosing was started when mice were approximately 5 months of age. The mice received intraperitoneal injections of vehicle only (PBS) or human IgG-FC (control protein) or TACI-FC4 (test protein) three times a week for 5 weeks.

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Group (5 mice each)	Treatment	Dose
1	untreated control	
2	vehicle only	
3	human IgG-FC	20 µg
4	human IgG-FC	100 µg
5	human TACI-FC4	20 µg
6	human TACI-FC4	

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[0170] Blood was collected twice during dosing and will be collected at least twice following dosing. Urine dipstick values for proteinuria and body weights were made every two weeks after dosing begins. Blood, urine dipstick value

and body weight were collected at the time of euthanasia. Weight of spleen, thymus, liver with gall bladder, left kidney and brain were taken. The spleen and thymus were divided for FACS analysis and histology. Submandibular salivary glands, mesenteric lymph node chain, liver lobe with gall bladder, cecum and large intestine, stomach, small intestine, pancreas, right kidney, adrenal gland, tongue with trachea and esophagus, heart and lungs will also be collected for histology.

[0171] Figure 6 shows an elevated level of ztnf4 in serum from NZBWF1 and MRL/lpr/lpr mice that correlates with the development of SLE. Figure 6A upper panel shows the correlation of ztnf4 serum levels with age, 68 NZBWF1 mice ranging from 10 to 40 weeks old and 10 week and 30 week old NZB/B control mice. The middle panel shows the correlation with proteinuria at three ranges, trace to 20 mg/dl (T-30), 100-300 ng/dl and 2000 mg/dl in NZBWF1 mice compared to control NZB/B mice. The lower panel shows ztnf4 levels with various titers of anti-ds DNA antibody in NZBWF1 mice compared to control NZB/B mice.

[0172] Figure 6B shows the same correlations made on 23 MRL/lpr/lpr mice ranging from 18-24 weeks old and 10 control 11 week old MRL/MpJ mice.

[0173] Figure 7 shows urinalysis results. Mice were considered to have proteinuria if the dipstick reading was ≥100 mg/dl. (A) PBS, (B) human IgG FC, 100 mg, (C) human IgG FC, 20 mg, (D) human TACI-IgG, 100 mg, and (E) human TACI-IgG, 20 mg. Mice treated with the soluble TACI-IgG fusion showed a reduction in proteinuria.

[0174] Analysis of peripheral blood from treated animals revealed that white blood cell and lymphocyte counts were reduced in TACI-FC treated mice (20 and 100 mg) when compared to FC (20 and 100 mg) and PBS treated mice, 2 weeks after the start of treatment. FAC analysis (lymphocyte gate) of peripheral blood drawn six weeks after treatment began (two weeks after last treatment was administered) and showed a dramatic decrease in percentage of B cells present in the samples. B cell levels were still in decline at five weeks after last treatment was administered, but not as dramatic. Table 9 provides the average (and standard deviation) for the mice in each treatment group (Table 9). The decline in the percent of B cells in peripheral blood was also observed two weeks into treatment.

Table 9

Treatment	Week 2		Week 5
	% B cells	% T cells	% B cells
PBS	26.05 (6.52)	67.05 (6.80)	20.83 (3.14)
100 mg FC	23.34 (5.77)	68.23 (7.30)	25.04 (8.07)
20 mg FC	24.09 (6.26)	65.27 (7.18)	18.96 (6.42)
100 mg TACI-FC	11.07 (5.03)	79.06 (6.71)	14.79 (4.76)
20 mg TACI-FC	16.37 (7.27)	69.72 (8.90)	19.14 (5.27)

Example 14

Soluble TACI-Ig in normal mice

[0175] TACI-FC was administered to Blab/C mice to monitor its effect on normal mice. Sixty, 8-week old female Balb/C mice (HSD) were divided into 12 groups of 5 mice. Prior to treatment the mice were weighed and blood was drawn for CBC and serum banking. Groups 1-9 received intraperitoneal injections (IP) of vehicle only (PBS) or human IgG-FC (control protein) or TACI-FC4 (test protein) daily for 12 days and were sacrificed on day 14. Groups 10 and 11 received IP injections three times per week for two weeks and were sacrificed on day 14.

Group (5 mice each)	Treatment	Dose
1	human TACI-FC4	200 mg
2	human TACI-FC4	100 mg
3	human TACI-FC4	20 µg
4	human TACI-FC4	5 µg
5	human FC4	200 µg
6	human FC4	100 mg
7	human FC4	20 mg
8	human FC4	5 mg

(continued)

Group (5 mice each)	Treatment	Dose
9	vehicle only	as used
10	human TACI-FC4	100 mg
11	human FC4	100 mg
12	untreated control	

[0176] Blood was collected on days 7 and 12. Blood and body weight were collected at the time of euthanasia. Weight of spleen, thymus, and brain were taken. The spleen and thymus were divided for FACS analysis and histology. Skin, spleen, mesenteric LN chain, submandibular salivary glands, ovary, uterus, cervix, bladder, mesenteric lymph node chain, liver lobe with gall bladder, cecum and large intestine, stomach, small intestine, pancreas, right kidney, adrenal gland, tongue with trachea and esophagus, heart, thymus, thigh muscle, left and right femur, brain will also be collected for histology.

[0177] As described above in Example 13, a significant reduction in percent B cells was seen on days 7 (by CBC) and 12 (using FACS) in peripheral blood cells taken from all TACI-FC4 treated samples compared to those treated with FC4 or PBS alone and analyzed by CBC or FACS. Additionally, there was nearly a 50% decrease in B cells in the spleens taken from animals treated with TACI-FC4 as compared to those from FC4 treated mice day 14.

Example 15

Anti-dsDNA ELISA

[0178] Autoimmunity is characterized by high levels of anti-double stranded DNA antibodies. To measure the levels anti-dsDNA antibodies in both the over expressing ztnf4 transgenic mice and the NZBW mice an ELISA assay was developed. A 96 well microtiter plate (Nunc) was coated with poly-L-lysine (Sigma) (20 µl/ml in 0.1 M Tris buffer pH 7.3) at 75 µl/well and incubated overnight at room temperature. The plates were then washed in dH₂O and coated with poly dAdT (Sigma) (20 µl/ml in 0.1 M Tris buffer pH 7.3) at 75 µl/well and incubated at room temperature for 60 minutes. The plates were then washed with dH₂O and blocked with 2%BSA (Sigma) in Tris Buffer for 30 minutes at room temperature followed by a final wash in dH₂O.

[0179] Serum samples were taken from the ztnf4 transgenic mice described in Example 10 and the NZBW mice described in Example 11. The serum samples were diluted 1:50 in 1% BSA/2% BGG (Calbiochem) in Tris Buffer. The diluted samples were then titrated into the coated plate at 1:50, 1:100, 1:200, 1:400, 1:800, 1:1600, 1:3200 and 1:6400 (50 µl/well) and incubated for 90 minutes at room temperature.

[0180] Plates were then washed in dH₂O and goat antimouse IgG-Fc-HRP (Cappel) diluted to 1:1000 in 1% BSA/2% BGG was added at 50µl/well. The plates were incubated for 60 minutes at room temperature. The plates were washed 5X in dH₂O and developed with OPD, 1 tablet/10 ml Novo D and plated at 100 µl/well. The developer was stopped with 1N H₂SO₄, 100 µl/well, and the OD read at 492 nm.

[0181] Figure 8 shows the anti-ds DNA levels in two ztnf4 transgenic mice (23 week old), two non-transgenic litter mates compared with the levels detected in serum from NZBWF1 (32 week old) and MRL/lpr/lpr (19 week old) mice.

Example 16

Soluble TACI-Ig in a Spontaneous Model of EAE

[0182] Twenty five female PLxSJL F1 mice (12 weeks old, Jackson Labs) are given a subcutaneous injection of 125 µg/mouse of antigen (myelin Proteolipid Protein, PLP, residues 139-151), formulated in complete Freund's Adjuvant. The mice are divided into 5 groups of 5 mice. Intraperitoneal injections of pertussis toxin (400 ng) are given on Day 0 and 2. The groups will be given a x, 10x, or 100x dose of TACI, BCMA or BR43x2, one group will receive vehicle only and one group will receive no treatment. Prevention therapy will begin on Day 0, intervention therapy will begin on day 7, or at onset of clinical signs. Signs of disease, weight loss, and paralysis manifest in approximately 10-14 days, and last for about 1 week. Animals are assessed daily by collecting body weights and assigning a clinical score to correspond to the extent of their symptoms. Clinical signs of EAE appear within 10-14 days of inoculation and persist for approximately 1 week. At the end of the study all animals are euthanized by gas overdose, and necropsied. The brain and spinal column are collected for histology or frozen for mRNA analysis. Body weight and clinical score data is plotted by individual and by group.

Clinical Score

[0183]

5	0 Normal
	0.5 Weak, tail tone may be reduced but not absent
10	1 Limp tail (cannot lift tail when mouse is picked up at base of tail)
	2 Limp tail, weak legs (cannot lift tail, can stay upright on hind legs but legs are shaky)
	3 Paresis (cannot sit with legs under body, walk in a paddling motion with legs behind)
	4 Paralysis (cannot move back legs, drags legs when trying to walk)
	5 Quadriplegia (paralysis in front legs or walking in a circular pattern, may have head tilt)
	6

15 Example 17

TACI-FC and the CIA model for Rheumatoid Arthritis

[0184] Eight week old male DBA/1J mice (Jackson Labs) are divided into groups of 5 mice/group and are given two subcutaneous injections of 50-100 μ l of 1mg/ml collagen (chick or bovine origin), at 3 week intervals. One control will not receive collagen injections. The first injection is formulated in Complete Freund's Adjuvant and the second injection is formulated in Incomplete Freund's Adjuvant. TACI-FC will be administered prophylactically at or prior to the second injection, or after the animal develops a clinical score of 2 or more that persists at least 24 hours. Animals begin to show symptoms of arthritis following the second collagen injection, usually within 2-3 weeks. Extent of disease is evaluated in each paw by using a caliper to measure paw thickness and assigning a clinical score (0-3) to each paw. Clinical Score, 0 Normal, 1 Toe(s) inflamed, 2 Mild paw inflammation, 3 Moderate paw inflammation, and 4 Severe paw inflammation. Animals will be euthanized after having established disease for a set period of time, usually 7 days. Paws are collected for histology or mRNA analysis, and serum is collected for immunoglobulin and cytokine assays.

30 Example 18

Neutralizing TACI antibodies

[0185] Polyclonal anti-peptide antibodies were prepared by immunizing 2 female New Zealand white rabbits with the peptide, huztnf4-1 SAGIAKLEEGPELQLAIPRE (SEQ ID NO:59) or huztnf4-2 SFKRGSALEEKENKELVKET (SEQ ID NO:60). The peptides were synthesized using an Applied Biosystems Model 431A peptide synthesizer (Applied Biosystems, Inc., Foster City, CA) according to manufacturer's instructions. The peptides were then conjugated to the carrier protein keyhole limpet hemocyanin (KLH) with maleimide-activation. The rabbits were each given an initial intraperitoneal (ip) injection of 200 μ g of peptide in Complete Freund's Adjuvant followed by booster ip injections of 100 μ g peptide in Incomplete Freund's Adjuvant every three weeks. Seven to ten days after the administration of the second booster injection (3 total injections), the animals were bled and the serum was collected. The animals were then boosted and bled every three weeks.

[0186] The ztnf4 peptide-specific rabbit seras were characterized by an ELISA titer check using 1 μ g/ml of the peptides used to make the antibody (SEQ ID NOs:59 and 60) as an antibody target. The 2 rabbit seras to the huztnf4-1 peptide (SEQ ID NO:59) have titer to their specific peptide at a dilution of 1:1E5 (1:100000). The 2 rabbit seras to the huztnf4-2 peptide (SEQ ID NO:60) had titer to their specific peptide at a dilution of 1:5E6 and to recombinant full-length proteins (N-terminal FLAG-tagged ztnf4 made in baculovirus (huztnf4s-NF-Bv) and C-terminally FLAG-tagged ztnf4 made in BHK cells) at a dilution of 1:5E6.

[0187] The ztnf4 peptide-specific polyclonal antibodies were affinity purified from the rabbit serum using CNBr-SEPHAROSE 4B protein columns (Pharmacia LKB) that were prepared using 10 mgs of the specific peptides (SEQ. ID. NOs.59 or 60) per gram CNBr-SEPHAROSE, followed by 20X dialysis in PBS overnight. Ztnf4-specific antibodies were characterized by an ELISA titer check using 1 μ g/ml of the appropriate peptide antigen or recombinant full-length protein (huztnf4s-NF-Bv) as antibody targets. The lower limit of detection (LLD) of the rabbit anti-huztnf4-1 affinity purified antibody on its specific antigen (huztnf4-1 peptide, SEQ ID NO:59) is a dilution of 5 ng/ml. The lower limit of detection (LLD) of the rabbit anti-huztnf4-2 affinity purified antibody on its specific antigen (huztnf4-2 peptide, SEQ ID NO:60) is a dilution of 0.5 ng/ml. The lower limit of detection (LLD) of the rabbit anti-huztnf4-2 affinity purified antibody on the recombinant protein huztnf4s-NF-Bv is a dilution of 5 ng/ml.

[0188] Mouse monoclonal antibodies were also generated and selected for inhibition of biotin-labeled

soluble ztnf4. None of the TACI monoclonal antibodies (248.14, 248.23, 248.24, or 246.3) block ztnf4 binding on BCMA. Monoclonal 248.23 reduces binding of 10 ng/ml ztnf4-biotin to about 50% when conditioned media is diluted to 1:243 and reduces binding to about 2X in undiluted media. Monoclonal 246.3 reduces binding of 10 ng/ml ztnf4-biotin to about 50% between a 1:243 and 1:181 dilution of conditioned media and reduces binding 5X in undiluted media.

5 [0189] The invention is not limited except as by the appended claims.

SEQUENCE LISTING

[0190]

10

<110> ZymoGenetics, Inc.

<120> SOLUBLE RECEPTOR BR43X2 AND METHODS OF USING

15

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<151> 1999-01-07

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<170> FastSEQ for Windows Version 3.0

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<212> DNA

<213> Homo sapiens

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<222> (6)...(746)

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5 1 5 10 15	
 gac cag gag gag cgc tgg tca ctc agc tgc cgc aag gag caa ggc aag Asp Gln Glu Glu Arg Trp Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys	98
10 20 25 30	
 ttc tat gac cat ctc ctg agg gac tgc atc agc tgt gcc tcc atc tgt Phe Tyr Asp His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile Cys	146
15 35 40 45	
 gga cag cac cct aag caa tgt gca tac ttc tgt gag aac aag ctc agg Gly Gln His Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu Arg	194
50 55 60	
 20 agc cca gtg aac ctt cca cca gag ctc agg aga cag cgg agt gga gaa Ser Pro Val Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly Glu	242
65 70 75	
 25	
 30	
 35	
 40	
 45	
 50	
 55	

	gtt gaa aac aat tca gac aac tcg gga agg tac caa gga ttg gag cac		290
5	Val Glu Asn Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu His		
	80	85	95
	aga ggc tca gaa gca agt cca gct ctc ccg ggg ctg aag ctg agt gca		338
	Arg Gly Ser Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser Ala		
10	100	105	110
	gat cag gtg gcc ctg gtc tac agc acg ctg ggg ctc tgc ctg tgt gcc		386
	Asp Gln Val Ala Leu Val Tyr Ser Thr Leu Gly Leu Cys Leu Cys Ala		
15	115	120	125
	gtc ctc tgc tgc ttc ctg gtg gcg gtg gcc tgc ttc ctc aag aag agg		434
	Val Leu Cys Cys Phe Leu Val Ala Val Ala Cys Phe Leu Lys Lys Arg		
20	130	135	140
	ggg gat ccc tgc tcc tgc cag ccc cgc tca agg ccc cgt caa agt ccg		482
	Gly Asp Pro Cys Ser Cys Gln Pro Arg Ser Arg Pro Arg Gln Ser Pro		
	145	150	155
25	gcc aag tct tcc cag gat cac gcg atg gaa gcc ggc agc cct gtg agc		530
	Ala Lys Ser Ser Gln Asp His Ala Met Glu Ala Gly Ser Pro Val Ser		
	160	165	170
30	aca tcc ccc gag cca gtg gag acc tgc agc ttc tgc ttc cct gag tgc		578
	Thr Ser Pro Glu Pro Val Glu Thr Cys Ser Phe Cys Phe Pro Glu Cys		
	180	185	190
35	agg gcg ccc acg cag gag agc gca gtc acg cct ggg acc ccc gac ccc		626
	Arg Ala Pro Thr Gln Glu Ser Ala Val Thr Pro Gly Thr Pro Asp Pro		
	195	200	205
40	act tgt gct gga agg tgg ggg tgc cac acc agg acc aca gtc ctg cag		674
	Thr Cys Ala Gly Arg Trp Gly Cys His Thr Arg Thr Thr Val Leu Gln		
	210	215	220
45	cct tgc cca cac atc cca gac agt ggc ctt ggc att gtg tgt gtg cct		722
	Pro Cys Pro His Ile Pro Asp Ser Gly Leu Gly Ile Val Cys Val Pro		
	225	230	235
50	gcc cag gag ggg ggc cca ggt gca taaatggggg tcagggaggg aaaggaggag		776
	Ala Gln Glu Gly Gly Pro Gly Ala		
	240	245	

5	ggagagagat ggagaggagg ggagagagaa agagaggtgg ggagagggga gagagatatg	836
	aggagagaga gacagaggag gcagagaggg agagaaacag aggagacaga gagggagaga	896
	gagacagagg gagagagaga cagagggaa gagagggcaga gagggaaaga ggcagagaag	956
	gaaagaggca gagagggaga gaggcagaga gggagagagg cagagagaca gagagggaga	1016
	gagggacaga gagagataga gcaggaggtc ggggactct gagtcccagt tcccagtgc	1076
10	gctgttaggtc gtcatcacct aaccacacgt gcaataaaagt cctcgtgcct gctgctcaca	1136
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<400> 2

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 Gln Glu Glu Arg Trp Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe
 5 20 25 30
 Tyr Asp His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile Cys Gly
 35 40 45
 Gln His Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu Arg Ser
 10 50 55 60
 Pro Val Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly Glu Val
 65 70 75 80
 Glu Asn Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu His Arg
 15 85 90 95
 Gly Ser Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser Ala Asp
 100 105 110
 Gln Val Ala Leu Val Tyr Ser Thr Leu Gly Leu Cys Leu Cys Ala Val
 20 115 120 125
 Leu Cys Cys Phe Leu Val Ala Val Ala Cys Phe Leu Lys Lys Arg Gly
 130 135 140
 Asp Pro Cys Ser Cys Gln Pro Arg Ser Arg Pro Arg Gln Ser Pro Ala
 25 145 150 155 160
 Lys Ser Ser Gln Asp His Ala Met Glu Ala Gly Ser Pro Val Ser Thr
 165 170 175
 Ser Pro Glu Pro Val Glu Thr Cys Ser Phe Cys Phe Pro Glu Cys Arg
 30 180 185 190
 Ala Pro Thr Gln Glu Ser Ala Val Thr Pro Gly Thr Pro Asp Pro Thr
 195 200 205
 Cys Ala Gly Arg Trp Gly Cys His Thr Arg Thr Thr Val Leu Gln Pro
 35 210 215 220
 Cys Pro His Ile Pro Asp Ser Gly Leu Gly Ile Val Cys Val Pro Ala
 225 230 235 240
 Gln Glu Gly Gly Pro Gly Ala

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10	cag gag gag cg ^c tgg tca ctc agc tgc cg ^c aag gag caa ggc aag ttc Gln Glu Glu Arg Trp Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe 20 25 30	96
15	tat gac cat ctc ctg agg gac tgc atc agc tgt gcc tcc atc tgt gga Tyr Asp His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile Cys Gly 35 40 45	144
20	cag cac cct aag caa tgt gca tac ttc tgt gag aac aag ctc agg agc Gln His Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu Arg Ser 50 55 60	192
25	cca gtg aac ctt cca cca gag ctc agg aga cag cg ^g agt gga gaa gtt Pro Val Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly Glu Val 65 70 75 80	240
30	gaa aac aat tca gac aac tcg gga agg tac caa gga ttg gag cac aga Glu Asn Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu His Arg 85 90 95	288
35	ggc tca gaa gca agt cca gct ctc ccg ggg ctg aag ctg agt gca gat Gly Ser Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser Ala Asp 100 105 110	336
40	cag gtg gcc ctg gtc tac agc acg Gln Val Ala Leu Val Tyr Ser Thr 115 120	360
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 5 Gln Glu Glu Arg Trp Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe
 20 25 30
 Tyr Asp His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile Cys Gly
 35 40 45
 10 Gln His Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu Arg Ser
 50 55 60
 Pro Val Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly Glu Val
 65 70 75 80
 15 Glu Asn Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu His Arg
 85 90 95
 Gly Ser Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser Ala Asp
 100 105 110
 20 Gln Val Ala Leu Val Tyr Ser Thr
 115 120

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 1 5 10
 40 agc cgt gtg gac cag gag gag cgc ttt cca cag ggc ctg tgg acg ggg 97
 Ser Arg Val Asp Gln Glu Glu Arg Phe Pro Gln Gly Leu Trp Thr Gly
 15 20 25
 45 gtg gct atg aga tcc tgc ccc gaa gag cag tac tgg gat cct ctg ctg 145
 Val Ala Met Arg Ser Cys Pro Glu Glu Gln Tyr Trp Asp Pro Leu Leu
 30 35 40

50

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	ggt acc tgc atg tcc tgc aaa acc att tgc aac cat cag agc cag cgc Gly Thr Cys Met Ser Cys Lys Thr Ile Cys Asn His Gln Ser Gln Arg 45	50	55	60	193
5	acc tgt gca gcc ttc tgc agg tca ctc agc tgc cgc aag gag caa ggc Thr Cys Ala Ala Phe Cys Arg Ser Leu Ser Cys Arg Lys Glu Gln Gly 65	70	75		241
10	aag ttc tat gac cat ctc ctg agg gac tgc atc agc tgt gcc tcc atc Lys Phe Tyr Asp His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile 80	85	90		289
15	tgt gga cag cac cct aag caa tgt gca tac ttc tgt gag aac aag ctc Cys Gly Gln His Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu 95	100	105		337
20	agg agc cca gtg aac ctt cca cca gag ctc agg aga cag cgg agt gga Arg Ser Pro Val Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly 110	115	120		385
25	gaa gtt gaa aac aat tca gac aac tcg gga agg tac caa gga ttg gag Glu Val Glu Asn Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu 125	130	135	140	433
30	cac aga ggc tca gaa gca agt cca gct ctc ccg ggg ctg aag ctg agt His Arg Gly Ser Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser 145	150	155		481
35	gca gat cag gtg gcc ctg gtc tac agc acg ctg ggg ctc tgc ctg tgt Ala Asp Gln Val Ala Leu Val Tyr Ser Thr Leu Gly Leu Cys Leu Cys 160	165	170		529
40	gcc gtc ctc tgc tgc ttc ctg gtg gcg gtg gcc tgc ttc ctc aag aag Ala Val Leu Cys Cys Phe Leu Val Ala Val Ala Cys Phe Leu Lys Lys 175	180	185		577
45	agg ggg gat ccc tgc tcc tgc cag ccc cgc tca agg ccc cgt caa agt Arg Gly Asp Pro Cys Ser Cys Gln Pro Arg Ser Arg Pro Arg Gln Ser 190	195	200		625
50	ccg gcc aag tct tcc cag gat cac gcg atg gaa gcc ggc agc cct gtg Pro Ala Lys Ser Ser Gln Asp His Ala Met Glu Ala Gly Ser Pro Val 205	210	215	220	673

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	agc aca tcc ccc gag cca gtg gag acc tgc agc ttc tgc ttc cct gag Ser Thr Ser Pro Glu Pro Val Glu Thr Cys Ser Phe Cys Phe Pro Glu	721
5	225 230 235	
	tgc agg gcg ccc acg cag gag agc gca gtc acg cct ggg acc ccc gac Cys Arg Ala Pro Thr Gln Glu Ser Ala Val Thr Pro Gly Thr Pro Asp	769
10	240 245 250	
	ccc act tgt gct gga agg tgg ggg tgc cac acc agg acc aca gtc ctg Pro Thr Cys Ala Gly Arg Trp Gly Cys His Thr Arg Thr Thr Val Leu	817
15	255 260 265	
	cag cct tgc cca cac atc cca gac agt ggc ctt ggc att gtg tgt gtg Gln Pro Cys Pro His Ile Pro Asp Ser Gly Leu Gly Ile Val Cys Val	865
20	270 275 280	
	cct gcc cag gag ggg ggc cca ggt gca taa atgggggtca gggagggaaa Pro Ala Gln Glu Gly Pro Gly Ala *	915
	285 290	
25	ggaggaggga gagagatgga gaggaggggga gagagaaaaga gaggtggggga gaggggagag agatatgagg agagagagac agaggaggca gaaagggaga gaaacagagg agacagagag	975
	ggagagagag acagagggag agagagacag aggggaagag aggcagagag ggaaagaggc	1035
30	agagaaggaa agagacaggc agagaaggag agaggcagag agggagagag gcagagaggg agagaggcag agagacagag agggagagag ggacagagag agatagagca ggaggtcggg	1095
	gcactctgag tcccagttcc cagtgcagct gtaggtcgtc atcacctaac cacacgtgca ataaaagtctt cgtgcctgtct gtcacagcc cccgagagcc ctcctcctg gagaataaaa	1155
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5	Gln Glu Glu Arg Phe Pro Gln Gly Leu Trp Thr Gly Val Ala Met Arg		
	20	25	30
	Ser Cys Pro Glu Glu Gln Tyr Trp Asp Pro Leu Leu Gly Thr Cys Met		
	35	40	45
10	Ser Cys Lys Thr Ile Cys Asn His Gln Ser Gln Arg Thr Cys Ala Ala		
	50	55	60
	Phe Cys Arg Ser Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe Tyr Asp		
	65	70	75
			80

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	His Leu Leu Arg Asp Cys Ile Ser Cys Ala Ser Ile Cys Gly Gln His		
	85	90	95
20	Pro Lys Gln Cys Ala Tyr Phe Cys Glu Asn Lys Leu Arg Ser Pro Val		
	100	105	110
	Asn Leu Pro Pro Glu Leu Arg Arg Gln Arg Ser Gly Glu Val Glu Asn		
	115	120	125
25	Asn Ser Asp Asn Ser Gly Arg Tyr Gln Gly Leu Glu His Arg Gly Ser		
	130	135	140
	Glu Ala Ser Pro Ala Leu Pro Gly Leu Lys Leu Ser Ala Asp Gln Val		
	145	150	155
30	Ala Leu Val Tyr Ser Thr Leu Gly Leu Cys Leu Cys Ala Val Leu Cys		
	165	170	175
	Cys Phe Leu Val Ala Val Ala Cys Phe Leu Lys Lys Arg Gly Asp Pro		
	180	185	190
35	Cys Ser Cys Gln Pro Arg Ser Arg Pro Arg Gln Ser Pro Ala Lys Ser		
	195	200	205
	Ser Gln Asp His Ala Met Glu Ala Gly Ser Pro Val Ser Thr Ser Pro		
	210	215	220
40	Glu Pro Val Glu Thr Cys Ser Phe Cys Phe Pro Glu Cys Arg Ala Pro		
	225	230	235
	Thr Gln Glu Ser Ala Val Thr Pro Gly Thr Pro Asp Pro Thr Cys Ala		
	245	250	255
45	Gly Arg Trp Gly Cys His Thr Arg Thr Thr Val Leu Gln Pro Cys Pro		
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	His Ile Pro Asp Ser Gly Leu Gly Ile Val Cys Val Pro Ala Gln Glu		
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50	Gly Gly Pro Gly Ala		
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<211> 995

<212> DNA

<213> Homo sapiens

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<222> (219)...(773)

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<400> 7

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agctgcttctt gctgcatttg ctctgaaatt cttgttagaga tattacttgt ccttccaggc 180
tgttcttctt gtagctccct ttgtgatc atg ttg cag atg gct ggg 236
Met Leu Gln Met Ala Gly
1 5

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50

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	cag tgc tcc caa aat gaa tat ttt gac agt ttg ttg cat gct tgc ata Gln Cys Ser Gln Asn Glu Tyr Phe Asp Ser Leu Leu His Ala Cys Ile	284
5	10 15 20	
	cct tgt caa ctt cga tgt tct tct aat act cct cct cta aca tgt cag Pro Cys Gln Leu Arg Cys Ser Ser Asn Thr Pro Pro Leu Thr Cys Gln	332
10	25 30 35	
	cgt tat tgt aat gca agt gtg acc aat tca gtg aaa gga acg aat gcg Arg Tyr Cys Asn Ala Ser Val Thr Asn Ser Val Lys Gly Thr Asn Ala	380
15	40 45 50	
	att ctc tgg acc tgt ttg gga ctg agc tta ata att tct ttg gca gtt Ile Leu Trp Thr Cys Leu Gly Leu Ser Leu Ile Ile Ser Leu Ala Val	428
20	55 60 65 70	
	ttc gtg cta atg ttt ttg cta agg aag ata agc tct gaa cca tta aag Phe Val Leu Met Phe Leu Leu Arg Lys Ile Ser Ser Glu Pro Leu Lys	476
25	75 80 85	
	gac gag ttt aaa aac aca gga tca ggt ctc ctg ggc atg gct aac att Asp Glu Phe Lys Asn Thr Gly Ser Gly Leu Leu Gly Met Ala Asn Ile	524
30	90 95 100	
	gac ctg gaa aag agc agg act ggt gat gaa att att ctt ccg aga ggc Asp Leu Glu Lys Ser Arg Thr Gly Asp Glu Ile Ile Leu Pro Arg Gly	572
35	105 110 115	
	ctc gag tac acg gtg gaa gaa tgc acc tct gaa gac tgc atc aag agc Leu Glu Tyr Thr Val Glu Glu Cys Thr Cys Glu Asp Cys Ile Lys Ser	620
40	120 125 130	
	aaa ccg aag gtc gac tct gac cat tgc ttt cca ctc cca gct atg gag Lys Pro Lys Val Asp Ser Asp His Cys Phe Pro Leu Pro Ala Met Glu	668
45	135 140 145 150	
	gaa ggc gca acc att ctt gtc acc acg aaa acg aat gac tat tgc aag Glu Gly Ala Thr Ile Leu Val Thr Thr Lys Thr Asn Asp Tyr Cys Lys	716
50	155 160 165	
	agc ctg cca gct gct ttg agt gct acg gag ata gag aaa tca att tct Ser Leu Pro Ala Ala Leu Ser Ala Thr Glu Ile Glu Lys Ser Ile Ser	764
55	170 175 180	

gct agg taa ttaaccattt cgactcgagc agtgccactt taaaaatctt 813
 Ala Arg *

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ttgtcagaat agatgatgtg tcagatctct ttaggatgac tgtattttc agttgccat 873
 acagctttt gtcctctaac tgtggaaact ctttatgtta gatatatttc tctaggttac 933
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 ga. 995

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 <212> PRT
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<400> 8

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Leu	Leu	His	Ala	Cys	Ile	Pro	Cys	Gln	Leu	Arg	Cys	Ser	Ser	Asn	Thr	
															30	
20																
Pro	Pro	Leu	Thr	Cys	Gln	Arg	Tyr	Cys	Asn	Ala	Ser	Val	Thr	Asn	Ser	
															45	
35																
Val	Lys	Gly	Thr	Asn	Ala	Ile	Leu	Trp	Thr	Cys	Leu	Gly	Leu	Ser	Leu	
															60	
50																
Ile	Ile	Ser	Leu	Ala	Val	Phe	Val	Leu	Met	Phe	Leu	Leu	Arg	Lys	Ile	
65															80	
70																
30	Ser	Ser	Glu	Pro	Leu	Lys	Asp	Glu	Phe	Lys	Asn	Thr	Gly	Ser	Gly	Leu
															95	
85																
Leu	Gly	Met	Ala	Asn	Ile	Asp	Leu	Glu	Lys	Ser	Arg	Thr	Gly	Asp	Glu	
100															110	
40	Ile	Ile	Leu	Pro	Arg	Gly	Leu	Glu	Tyr	Thr	Val	Glu	Glu	Cys	Thr	Cys
															125	
115																
Glu	Asp	Cys	Ile	Lys	Ser	Lys	Pro	Lys	Val	Asp	Ser	Asp	His	Cys	Phe	
130															140	
45	Pro	Leu	Pro	Ala	Met	Glu	Glu	Gly	Ala	Thr	Ile	Leu	Val	Thr	Lys	
145															160	
150																
Thr	Asn	Asp	Tyr	Cys	Lys	Ser	Leu	Pro	Ala	Ala	Leu	Ser	Ala	Thr	Glu	
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170																
50	Ile	Glu	Lys	Ser	Ile	Ser	Ala	Arg								
180																

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<210> 9

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 Phe Pro Gln Gly Leu Trp Thr Gly Val Ala Met Arg Ser Cys Pro Glu
 20 25 30
 Glu Gln Tyr Trp Asp Pro Leu Leu Gly Thr Cys Met Ser Cys Lys Thr
 10 35 40 45
 Ile Cys Asn His Gln Ser Gln Arg Thr Cys Ala Ala Phe Cys Arg Ser
 50 55 60
 Leu Ser Cys Arg Lys Glu Gln Gly Lys Phe Tyr Asp His Leu Leu Arg
 15 65 70 75 80
 Asp Cys Ile Ser Cys Ala Ser Ile Cys Gly Gln His Pro Lys Gln Cys
 85 90 95
 Ala Tyr Phe Cys Glu Asn Lys Leu Arg Ser Pro Val Asn Leu Pro Pro
 20 100 105 110
 Glu Leu Arg Arg Gln Arg Ser Gly Glu Val Glu Asn Asn Ser Asp Asn
 115 120 125
 Ser Gly Arg Tyr Gln Gly Leu Glu His Arg Gly Ser Glu Ala Ser Pro
 25 130 135 140
 Ala Leu Pro Gly Leu Lys Leu Ser Ala Asp Gln Val Ala Leu Val Tyr
 145 150 155 160
 Ser Thr Leu Gly Leu Cys Leu Cys Ala Val Leu Cys Cys Phe Leu Val
 30 165 170 175
 Ala Val Ala Cys Phe Leu Lys Lys Arg Gly Asp Pro Cys Ser Cys Gln
 180 185 190
 Pro Arg Ser Arg Pro Arg Gln Ser Pro Ala Lys Ser Ser Gln Asp His
 35 195 200 205
 Ala Met Glu Ala Gly Ser Pro Val Ser Thr Ser Pro Glu Pro Val Glu
 210 215 220
 Thr Cys Ser Phe Cys Phe Pro Glu Cys Arg Ala Pro Thr Gln Glu Ser
 40 225 230 235 240
 Ala Val Thr Pro Gly
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<223> Motif describing the cysteine-rich pseudo-repeat domain
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 <222> (1)...(2)
 <223> Each Xaa is independently any amino acid residue except cysteine, or absent.
 <221> VARIANT

<222> (4)...(4)

<223> Xaa is any amino acid residue except cysteine.

5 <221> VARIANT

<222> (5)...(5)

<223> Xaa is glutamine, glutamic acid, or lysine.

10 <221> VARIANT

<222> (6)...(6)

<223> Xaa is glutamine, glutamic acid, lysine, asparagine, arginine, aspartic acid, histidine, or serine.

15 <221> VARIANT

<222> (7)...(7)

<223> Xaa is glutamine or glutamic acid.

20 <221> VARIANT

<222> (8)...(9)

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35 <221> VARIANT

<222> (19)...(19)

<223> Xaa is isoleucine, methionine, leucine, or valine.

40 <221> VARIANT

<222> (20)...(20)

<223> Xaa is any amino acid residue except cysteine.

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<222> (22)...(24)

<223> Each Xaa is independently any amino acid residue except cysteine.

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<222> (26)...(31)

<223> Each Xaa is independently any amino acid residue except cysteine.

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<222> (32)...(33)

<223> Each Xaa is independently any amino acid residue except cysteine, or absent.

<221> VARIANT

<222> (35)...(36)

<223> Each Xaa is independently any amino acid residue except cysteine.

55 <221> VARIANT

<222> (37)...(37)

<223> Xaa is tyrosine or phenylalanine.

<221> VARIANT

<222> (39)...(40)

<223> Each Xaa is independently any amino acid residue except cysteine, or absent.

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Xaa	Xaa	Cys	Xaa	Asp	Xaa	Leu	Leu	Xaa								
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10	Xaa	Cys	Xaa	Xaa	Cys	Xaa	Xaa	Xaa	Cys	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	
																30
	Xaa	Cys	Xaa	Xaa	Xaa	Cys	Xaa	Xaa								
	35															

15

<400> 11

<211> 360

<212> DNA

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<213> Artificial Sequence

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<223> Degenerate oligonucleotide sequence encoding the polypeptide of SEQ ID NO:4

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<221> variation

<222> (1)...(360)

<223> Each N is independently A, T, G, or C.

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<400> 11

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<210> 12

<211> 741

<212> DNA

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<213> Artificial Sequence

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<223> Degenerate oligonucleotide sequence encoding a polypeptide of SEQ ID NO:2

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<221> variation

<222> (1)...(741)

<223> Each N is independently A, T, G, or C.

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	athwsntgyg cnwsnathtg yggncarcay ccnaarcart gygcntaytt ytgyaraay	180
	aarytnmgnw snccngtnaa yytnccnccn garytnmgnm gncarmgnws nggngargtn	240
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	70 75 80	
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188-88

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45

50

55

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40	gtc tcc aac aaa gcc ctc cca tcc atc gag aaa acc atc tcc aaa Val Ser Asn Lys Ala Leu Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys 130 135 140	432
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50	gcc aaa ggg cag ccc cga gaa cca cag gtg tac acc ctg ccc cca tcc Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser 145 150 155	480
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95		
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 Ile Pro Arg Glu
 20

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Ser Phe Lys Arg Gly Ser Ala Leu Glu Glu Lys Glu Asn Lys Glu Leu
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 Val Lys Glu Thr
 20

15 Claims

1. Use of a compound selected from the group consisting of:

 - a) a polypeptide comprising the extracellular domain of BR43x2 (SEQ ID NO:2);
 - b) a soluble polypeptide comprising the extracellular domain of TACI;
 - c) a polypeptide comprising the extracellular domain of BCMA;
 - d) a polypeptide comprising the sequence of SEQ ID NO:10;
 - e) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:2;
 - f) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:4;
 - 25 g) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:6;
 - h) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:8;
 - i) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:10;
 - j) a polypeptide of SEQ ID NO:4;
 - 30 l) amino acid residues 1-166 of SEQ ID NO:6;
 - m) amino acid residues 8-37 of SEQ ID NO:8; and
 - n) amino acid residues 1-48 of SEQ ID NO:8,

in the manufacture of a medicament for inhibiting ztnf4 activity in a mammal.

- 35 2. Use according to claim 1, wherein said mammal is a primate.

3. Use of a compound selected from the group consisting of:

- 40 a) a polypeptide comprising the extracellular domain of BR43x2 (SEQ ID NO:2);
b) a soluble polypeptide comprising the extracellular domain of TACI;
c) a polypeptide comprising the extracellular domain of BCMA;
d) a polypeptide comprising the sequence of SEQ ID NO:10;
e) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:2;
f) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:4;
g) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:6;
h) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:8;
i) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:10;
j) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:18;
50 k) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:20;
l) a polypeptide of SEQ ID NO:4;
m) amino acid residues 1-166 of SEQ ID NO:6;
n) amino acid residues 8-37 of SEQ ID NO:8; and
o) amino acid residues 1-48 of SEQ ID NO:8,

55 in the manufacture of a medicament for inhibiting BR43x2, TACI or BCMA receptor-ztnf4 engagement.

4. Use according to claims 1-3, wherein said compound is a fusion protein consisting of a first portion and a second portion joined by a peptide bond, said first portion comprising a polypeptide selected from the group consisting of:

- a) a polypeptide comprising the sequence of SEQ ID NO:8;
- b) a polypeptide comprising amino acid residues 25-58 of SEQ ID NO:2;
- c) a polypeptide comprising amino acid residues 34-66 of SEQ ID NO:6;
- d) a polypeptide comprising amino acid residues 71-104 of SEQ ID NO:6;
- e) a polypeptide comprising amino acid residues 25-104 of SEQ ID NO:6;
- f) a polypeptide comprising amino acid residues 8-37 of SEQ ID NO:8;
- g) a polypeptide comprising amino acid residues 41-88 of SEQ ID NO:8;
- h) a polypeptide comprising amino acid residues 8-88 of SEQ ID NO:8; and

10 said second portion comprising another polypeptide.

- 5 5. Use according to claim 4, wherein said first portion further comprises a polypeptide selected from the group consisting of:

- 15 a) amino acid residues 59-120 of SEQ ID NO:2;
- b) amino acid residues 105-166 of SEQ ID NO:6; and
- c) amino acid residues 89-150 of SEQ ID NO:8.

- 20 6. Use according to claim 4, wherein said first portion is selected from the group consisting of:

- 25 a) a polypeptide comprising the extracellular domain of BR43x2 (SEQ ID NO:2);
- b) a soluble polypeptide comprising the extracellular domain of TACI; and
- c) a polypeptide comprising the extracellular domain of BCMA.

30 7. Use according to Claim 4, wherein said first portion is selected from the group consisting of:

- 35 a) a polypeptide of SEQ ID NO:4;
- b) amino acid residues 1-154 of SEQ ID NOS:6; and
- c) amino acid residues 1-48 of SEQ ID NO:8.

- 30 8. Use according to claims 4-7, wherein said second portion is an immunoglobulin heavy chain constant region.

- 35 9. Use according to claim 8, wherein said immunoglobulin heavy chain constant region is a human immunoglobulin heavy chain constant region.

- 40 10. Use according to claim 9, wherein said human immunoglobulin heavy chain constant region is a human immunoglobulin heavy chain constant region of IgG1.

- 45 11. Use according to claims 8 to 10, wherein said medicament comprises a multimer of said fusion proteins.

- 40 12. Use according to claim 11, wherein said medicament comprises an immunoglobulin heavy chain constant region which contains two constant region domains and lacks the variable region.

- 45 13. Use according to claims 1-12, wherein the medicament is for Treatment of B lymphocytes.

- 45 14. Use according to claim 13, wherein the B lymphocytes are activated B lymphocytes.

- 45 15. Use according to claim 13, wherein the said B lymphocytes are resting B lymphocytes.

50 16. Use according to claims 1-15, wherein the medicament is for inhibiting the antibody production.

- 50 17. Use according to claim 16, wherein said antibody production is associated with an autoimmune disease.

55 18. Use according to claim 17, wherein said autoimmune disease is systemic lupus erythematosys, myasthenia gravis, multiple sclerosis, or rheumatoid arthritis.

- 55 19. Use according to claims 1-18, for treatment of asthma, bronchitis, emphysema, and end stage renal failure.

20. Use according to claim 19, wherein said renal disease is glomerulonephritis, vasculitis, nephritis or pyelonephritis.
21. Use according to claims 1-20, for treatment of renal neoplasms, multiple myelomas, lymphomas, light chain neuropathy or amyloidosis.
- 5 22. Use according to claims 1-21, for inhibiting effector T cells.
23. Use according to claim 22, for moderating immune response.
- 10 24. Use according to claim 22, wherein said inhibition further comprises immunosuppression.
25. Use according to claim 24, wherein said immunosuppression is associated with graft rejection, graft versus host disease, autoimmune disease or inflammation.
- 15 26. Use according to claim 17, wherein said autoimmune disease is insulin dependent diabetes mellitus or Crohn's Disease.
27. Use according to claims 1-26, for treatment of inflammation.
- 20 28. Use according to claim 27, wherein said inflammation is associated with joint pain, swelling, anemia or septic shock.
29. An isolated polynucleotide molecule encoding a polypeptide of SEQ ID NO:2.
30. An isolated polynucleotide molecule of SEQ ID NO:1.
- 25 31. An expression vector comprising the following operably linked elements:
- 30 a transcription promoter;
 a polynucleotide molecule according to claim 29; and
 a transcription terminator.
32. A cultured cell into which has been introduced an expression vector according to claim 31, wherein said cultured cell expresses said polypeptide encoded by said polynucleotide molecule.
- 35 33. A method of producing a polypeptide comprising:
 culturing a cell into which has been introduced an expression vector according to claim 31;
 whereby said cell expresses said polypeptide encoded by said polynucleotide molecule; and
 recovering said expressed polypeptide.
- 40 34. An isolated polypeptide having the sequence of SEQ ID NO:2.
35. A polypeptide of claim 34, in combination with a pharmaceutically acceptable vehicle.
- 45 36. A pharmaceutical composition comprising:
 a) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:2;
 b) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:4;
 c) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:6;
 d) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:8; or
 e) an antibody or antibody fragment which specifically binds to a polypeptide of SEQ ID NO:10;
 and a pharmaceutically acceptable carrier.
- 50 37. A composition according to claim 36 wherein said antibody or antibody fragment is selected from the group consisting of:
 a) polyclonal antibody;

- b) murine monoclonal antibody;
- c) humanized antibody derived from b); and
- d) human monoclonal antibody.

5 38. A composition according to claims 36 and 37, wherein said antibody fragment is selected from the group consisting
of F(ab'), Fab, Fv, scFv.

Patentansprüche

- 10 1. Verwendung eines Verbindung, ausgewählt aus der Gruppe, bestehend aus:
- a) einem Polypeptid, umfassend die extrazelluläre Domäne von BR43x2 (SEQ ID Nr. 2);
 - b) einem löslichen Polypeptid, umfassend die extrazelluläre Domäne von TACI;
 - c) einem Polypeptid, umfassend die extrazelluläre Domäne von BCMA;
 - d) einem Polypeptid, umfassend die Sequenz der SEQ ID Nr. 10;
 - e) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 2 bindet;
 - f) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 4 bindet;
 - 20 g) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 6 bindet;
 - h) einem Antikörper oder Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 8 bindet;
 - i) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 10 bindet;
 - k) einem Polypeptid der SEQ ID NR. 4;
 - l) den Aminosäureresten 1-166 der SEQ ID Nr. 6;
 - m) den Aminosäureresten 8-37 der SEQ ID Nr. 8; und
 - 30 n) den Aminosäureresten 1-48 der SEQ ID Nr. 8,

bei der Herstellung eines Medikaments zum Inhibieren der ztnf4-Aktivität in einem Säuger.

2. Verwendung gemäß Anspruch 1, worin der Säuger ein Primat ist.

- 35 3. Verwendung eines Verbindung, ausgewählt aus der Gruppe, bestehend aus:

- a) einem Polypeptid, umfassend die extrazelluläre Domäne von BR43x2 (SEQ ID Nr. 2);
- b) einem löslichen Polypeptid, umfassend die extrazelluläre Domäne von TACI;
- c) einem Polypeptid, umfassend die extrazelluläre Domäne von BCMA;
- d) einem Polypeptid, umfassend die Sequenz der SEQ ID Nr. 10;
- e) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 2 bindet;
- f) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 4 bindet;
- 40 g) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 6 bindet;
- h) einem Antikörper oder Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 8 bindet;
- i) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 10 bindet;
- j) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 18 bindet;
- 45 k) einem Antikörper oder einem Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 20 bindet;
- l) einem Polypeptid der SEQ ID NR. 4;
- m) den Aminosäureresten 1-166 der SEQ ID Nr. 6;
- n) den Aminosäureresten 8-37 der SEQ ID Nr. 8; und

- o) den Aminosäureresten 1-48 der SEQ ID Nr. 8, bei der Herstellung eines Medikaments zum Inhibieren des ztnf4-Eingriffs in den BR43x2-, TACI- oder BCMA-Rezeptor.
5. Verwendung gemäß einem der Ansprüche 1-3, worin die Verbindung ein Fusionsprotein ist, bestehend aus einem ersten Teil und einem zweiten Teil, verbunden über eine Peptidbindung, wobei der erste Teil ein Polypeptid umfasst, ausgewählt aus der Gruppe, bestehend aus:
- einem Polypeptid, umfassend die Sequenz aus SEQ ID Nr. 8;
 - einem Polypeptid, umfassend Aminosäurereste 25-58 der SEQ ID Nr. 2;
 - 10 einem Polypeptid, umfassend Aminosäurereste 34-66 der SEQ ID Nr. 6;
 - einem Polypeptid, umfassend Aminosäurereste 71-104 der SEQ ID Nr. 6;
 - einem Polypeptid, umfassend Aminosäurereste 25-104 der SEQ ID Nr. 6;
 - f) einem Polypeptid, umfassend Aminosäurereste 8-37 der SEQ ID Nr. 8;
 - 15 g) einem Polypeptid, umfassend Aminosäurereste 41-88 der SEQ ID Nr. 8;
 - h) einem Polypeptid, umfassend Aminosäurereste 8-88 der SEQ ID Nr. 8; und wobei der zweite Teil ein anderes Polypeptid umfasst.
5. Verwendung gemäß Anspruch 4, worin der erste Teil zusätzlich ein Polypeptid umfasst, ausgewählt aus der Gruppe, bestehend aus:
- a) den Aminosäureresten 59-120 der SEQ ID Nr. 2;
 - b) den Aminosäureresten 105-166 der SEQ ID Nr. 6; und
 - c) den Aminosäureresten 89-150 der SEQ ID Nr. 8.
25. Verwendung gemäß Anspruch 4, worin der erste Teil ausgewählt ist aus der Gruppe, bestehend aus:
- a) einem Polypeptid, umfassend die extrazelluläre Domäne von BR43x2 (SEQ ID Nr. 2);
 - b) einem löslichen Polypeptid, umfassend die extrazelluläre Domäne von TACI; und
 - c) einem Polypeptid, umfassend die extrazelluläre Domäne von BCMA.
30. 7. Verwendung gemäß Anspruch 4, worin der erste Teil ausgewählt ist aus der Gruppe, bestehend aus:
- a) einem Polypeptid der SEQ ID Nr. 4;
 - b) den Aminosäureresten 1-154 der SEQ ID Nr. 6; und
 - c) den Aminosäureresten 1-48 der SEQ ID Nr. 8.
8. Verwendung gemäß den Ansprüchen 4-7, worin der zweite Teil der konstante Bereich einer schweren Kette eines Immunglobulins ist.
40. 9. Verwendung gemäß Anspruch 8, worin der konstante Bereich einer schweren Kette eines Immunglobulins ein konstanter Bereich einer schweren Kette eines humanen Immunglobulins ist.
10. Verwendung gemäß Anspruch 9, worin der konstante Bereich einer schweren Kette eines humanen Immunglobulins ein konstanter Bereich der schweren Kette eines humanen Immunglobulins vom Typ IgG1 ist.
45. 11. Verwendung gemäß den Ansprüchen 8 bis 10, worin das Medikament ein Multimer der Fusionsproteine umfasst.
12. Verwendung gemäß Anspruch 11, worin das Medikament einen konstanten Bereich einer schweren Kette eines Immunglobulins umfasst, der zwei Domänen des konstanten Bereichs enthält und dem der variable Bereich fehlt.
50. 13. Verwendung gemäß den Ansprüchen 1-12, worin das Medikament zur Behandlung von B-Lymphozyten bestimmt ist.
14. Verwendung gemäß Anspruch 13, worin die B-Lymphozyten aktivierte B-Lymphozyten sind.
55. 15. Verwendung gemäß Anspruch 13, worin die B-Lymphozyten ruhende B-Lymphozyten sind.
16. Verwendung gemäß den Ansprüchen 1-15, worin das Medikament zur Inhibition der Antikörperproduktion bestimmt

ist.

17. Verwendung gemäß Anspruch 16, worin die Antikörperproduktion mit einer Autoimmunerkrankung assoziiert ist.

5 18. Verwendung gemäß Anspruch 17, worin die Autoimmunerkrankung der systemische Lupus Erythematoses, die Myasthenia Gravis, die Multiple Sklerose oder die rheumatoide Arthritis ist.

19. Verwendung gemäß den Ansprüche 1-18 zur Behandlung von Asthma, Bronchitis, Emphysemen und dem Nie-
renversagen im Endstadium.

10 20. Verwendung gemäß Anspruch 19, worin es sich bei der Nierenerkrankung um die Glomerulonephritis, die Vasku-
litis, die Nephritis oder die Pyelonephritis handelt.

15 21. Verwendung gemäß den Ansprüchen 1-20 zur Behandlung von renalen Neoplasmen, multiplen Myelomen, Lym-
phomen, der leichten Ketten-Nurotherapie oder der Amyloidose.

22. Verwendung gemäß den Ansprüchen 1-21 zur Inhibition von Effektor-T-Zellen.

23. Verwendung gemäß Anspruch 22 zum Abmildern einer Immunantwort.

20 24. Verwendung gemäß Anspruch 22, worin die Inhibition zusätzlich eine Immunsuppression umfasst.

25 25. Verwendung gemäß Anspruch 24, worin die Immunsuppression mit der Transplantatabstoßung, den Graft-versus-
Host-Erkrankungen (GvH-Reaktionen), einer Autoimmunerkrankung oder einer Entzündung assoziiert ist.

26. Verwendung gemäß Anspruch 17, worin die Autoimmunerkrankung der insulinabhängige Diabetes Mellitus oder Morbus Crohn ist.

27. Verwendung gemäß den Ansprüchen 1-26 zur Behandlung von Entzündungen.

30 28. Verwendung gemäß Anspruch 27, worin die Entzündung mit Gelenkschmerzen, Schwellungen, Anämie oder dem septischen Schock assoziiert ist.

29. Isoliertes Polynukleotidmolekül, kodierend ein Polypeptid der SEQ ID Nr. 2.

35 30. Isoliertes Polynukleotidmolekül der SEQ ID Nr. 1.

31. Expressionsvektor, umfassend die folgenden, operativ miteinander verbundenen Elemente:

40 einen Transkriptionspromoter;
ein Polynukleotidmolekül gemäß Anspruch 29; und
einen Transkriptionsterminator.

45 32. Kultivierte Zelle, in die ein Expressionsvektor gemäß Anspruch 31 eingeführt worden ist, wobei die kultivierte Zelle das von dem Polynukleotidmolekül kodierte Polypeptid exprimiert.

33. Verfahren zum Produzieren eines Polypeptids, umfassend:

50 Kultivieren einer Zelle, in die ein Expressionsvektor gemäß Anspruch 31 eingeführt worden ist;
wodurch die Zelle das von dem Polynukleotidmolekül kodierte Polypeptid exprimiert; und
Wiedergewinnen des exprimierten Polypeptids.

34. Isoliertes Polypeptid mit der Sequenz der SEQ ID Nr. 2.

55 35. Polypeptid gemäß Anspruch 34, in Verbindung mit einem pharmazeutisch annehmbaren Träger.

36. Pharmazeutische Zubereitung, umfassend:

- a) einen Antikörper oder ein Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 2 bindet;
5 b) einen Antikörper oder ein Antikörperfragment, der oder das spezifisch an ein Polypeptid der SEQ ID Nr. 4 bindet;
c) einen Antikörper oder ein Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 6 bindet;
10 d) einen Antikörper oder ein Antikörperfragment, der bzw. das spezifisch an ein Polypeptid der SEQ ID Nr. 8 bindet;
e) einen Antikörper oder ein Antikörperfragment, der bzw. das spezifisch an ein Polypeptider SEQ ID Nr. 10 bindet; und

einen pharmazeutisch annehmbaren Träger.

- 15 37. Zubereitung gemäß Anspruch 36, worin der Antikörper oder das Antikörperfragment aus der Gruppe ausgewählt ist, bestehend aus:

- 20 a) einem polyclonalen Antikörper;
b) einem monoklonalen Antikörper der Maus;
c) einem aus b) abgeleiteten, humanisierten Antikörper; und
d) einem menschlichen monoklonalen Antikörper.

- 25 38. Zubereitung gemäß den Ansprüchen 36 und 37, worin das Antikörperfragment ausgewählt ist aus der Gruppe, bestehend aus F(ab'), Fab, Fv, scFv.

Revendications

1. Utilisation d'un composé choisi dans le groupe consistant en :

- 30 a) un polypeptide comprenant le domaine extracellulaire de BR43x2 (SEQ ID NO:2) ;
b) un polypeptide soluble comprenant le domaine extracellulaire de TACI ;
c) un polypeptide comprenant le domaine extracellulaire de BCMA ;
d) un polypeptide comprenant la séquence de SEQ ID NO:10 ;
e) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:2 ;
35 f) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:4 ;
g) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:6 ;
h) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:8 ;
i) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:10 ;
k) un polypeptide de SEQ ID NO:4 ;
40 l) les résidus d'acides aminés 1-166 de SEQ ID NO:6 ;
m) les résidus d'acides aminés 8-37 de SEQ ID NO:8 ; et
n) les résidus d'acides aminés 1-48 de SEQ ID NO:8 ,

dans la fabrication d'un médicament pour inhiber l'activité de ztnf4 chez un mammifère.

- 45 2. Utilisation selon la revendication 1 où ledit mammifère est un primate.

3. Utilisation d'un composé choisi dans le groupe consistant en :

- 50 a) un polypeptide comprenant le domaine extracellulaire de BR43x2 (SEQ ID NO:2) ;
b) un polypeptide soluble comprenant le domaine extracellulaire de TACI ;
c) un polypeptide comprenant le domaine extracellulaire de BCMA ;
d) un polypeptide comprenant la séquence de SEQ ID NO:10 ;
e) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:2 ;
55 f) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:4 ;
g) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:6 ;
h) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:8 ;
i) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:10 ;

- j) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:18 ;
k) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:20 ;
l) un polypeptide de SEQ ID NO:4 ;
5 m) les résidus d'acides aminés 1-166 de SEQ ID NO:6 ;
n) les résidus d'acides aminés 8-37 de SEQ ID NO:8 ; et
o) les résidus d'acides aminés 1-48 de SEQ ID NO:8,

dans la fabrication d'un médicament pour inhiber l'engagement de ztnf4 avec le récepteur BR43x2, TACI ou BCMA.

- 10 4. Utilisation selon les revendications 1-3 où ledit composé est une protéine de fusion consistant en une première partie et une seconde partie reliées par une liaison peptidique, ladite première partie comprenant un polypeptide choisi dans le groupe consistant en :

- 15 a) un polypeptide comprenant la séquence de SEQ ID NO:8 ;
b) un polypeptide comprenant les résidus d'acides aminés 25-58 de SEQ ID NO:2 ;
c) un polypeptide comprenant les résidus d'acides aminés 34-66 de SEQ ID NO:6 ;
d) un polypeptide comprenant les résidus d'acides aminés 71-104 de SEQ ID NO:6 ;
e) un polypeptide comprenant les résidus d'acides aminés 25-104 de SEQ ID NO:6 ;
f) un polypeptide comprenant les résidus d'acides aminés 8-37 de SEQ ID NO:8 ;
20 g) un polypeptide comprenant les résidus d'acides aminés 41-88 de SEQ ID NO :8 ;
h) un polypeptide comprenant les résidus d'acides aminés 8-88 de SEQ ID NO:8 ; et

ladite seconde partie comprenant un autre polypeptide.

- 25 5. Utilisation selon la revendication 4 où ladite première partie comprend en outre un polypeptide choisi dans le groupe consistant en :

- 30 a) les résidus d'acides aminés 59-120 de SEQ ID NO:2 ;
b) les résidus d'acides aminés 105-166 de SEQ ID NO:6 ; et
c) les résidus d'acides aminés 89-150 de SEQ ID NO:8.

6. Utilisation selon la revendication 4 où ladite première partie est choisie dans le groupe consistant en :

- 35 a) un polypeptide comprenant le domaine extracellulaire de BR43x2 (SEQ ID NO:2) ;
b) un polypeptide soluble comprenant le domaine extracellulaire de TACI ; et
c) un polypeptide comprenant le domaine extracellulaire de BCMA.

7. Utilisation selon la revendication 4 où ladite première partie est choisie dans le groupe consistant en :

- 40 a) un polypeptide de SEQ ID NO:4 ;
b) les résidus d'acides aminés 1-154 de SEQ ID NO:6 ; et
c) les résidus d'acides aminés 1-48 de SEQ ID NO:8.

- 45 8. Utilisation selon les revendications 4-7 où ladite seconde partie est une région constante de chaîne lourde d'immunoglobuline.

9. Utilisation selon la revendication 8 où ladite région constante de chaîne lourde d'immunoglobuline est une région constante de chaîne lourde d'immunoglobuline humaine.

- 50 10. Utilisation selon la revendication 9 où ladite région constante de chaîne lourde d'immunoglobuline humaine est une région constante de chaîne lourde d'immunoglobuline humaine de IgG1.

11. Utilisation selon les revendications 8 à 10 où ledit médicament comprend un multimère desdites protéines de fusion.

- 55 12. Utilisation selon la revendication 11 où ledit médicament comprend une région constante de chaîne lourde d'immunoglobuline qui contient deux domaines de région constante et qui est dépourvue de la région variable.

13. Utilisation selon les revendications 1-12 où le médicament est destiné au traitement de lymphocytes B.
14. Utilisation selon la revendication 13 où les lymphocytes B sont des lymphocytes B activés.
- 5 15. Utilisation selon la revendication 13 où lesdits lymphocytes B sont des lymphocytes B quiescents.
16. Utilisation selon les revendications 1-15 où le médicament est destiné à inhiber la production d'anticorps.
- 10 17. Utilisation selon la revendication 16 où ladite production d'anticorps est associée avec une maladie auto-immune.
18. Utilisation selon la revendication 17 où ladite maladie auto-immune est le lupus érythémateux aigu disséminé, la myasthénie grave, la sclérose en plaques ou la polyarthrite rhumatoïde.
- 15 19. Utilisation selon les revendications 1-18 pour le traitement de l'asthme, de la bronchite, de l'emphysème et de l'insuffisance rénale au stade final.
- 20 20. Utilisation selon la revendication 19 où ladite maladie rénale est la glomérulonéphrite, la vascularite, la néphrite ou la pyélonéphrite.
21. Utilisation selon les revendications 1-20 pour le traitement des néoplasmes rénaux, des myélomes multiples, des lymphomes, de la neuropathie à chaînes légères ou de l'amyloïdose.
22. Utilisation selon les revendications 1-21 pour inhiber les cellules T effectrices. ..
- 25 23. Utilisation selon la revendication 22 pour modérer la réponse immunitaire.
24. Utilisation selon la revendication 22 où ladite inhibition comprend en outre l'immunosuppression.
- 25 25. Utilisation selon la revendication 24 où ladite immunosuppression est associée avec un rejet de greffe, la maladie du greffon contre l'hôte, une maladie auto-immune ou une inflammation.
26. Utilisation selon la revendication 17 où ladite maladie auto-immune est le diabète sucré insulino-dépendant ou la maladie de Crohn.
- 35 27. Utilisation selon les revendications 1-26 pour le traitement d'une inflammation.
28. Utilisation selon la revendication 27 où ladite inflammation est associée à une douleur articulaire, un gonflement, une anémie ou un choc septique.
- 40 29. Molécule polynucléotidique isolée codant un polypeptide de SEQ ID NO: 2.
30. Molécule polynucléotidique isolée de SEQ ID NO:1.
31. Vecteur d'expression comprenant les éléments liés de manière active suivants :
45 un promoteur de transcription ;
 une molécule polynucléotidique selon la revendication 29 ; et
 un terminateur de transcription.
- 50 32. Cellule cultivée dans laquelle a été introduit un vecteur d'expression selon la revendication 31 où ladite cellule cultivée exprime ledit polypeptide codé par ladite molécule polynucléotidique.
33. Procédé de production d'un polypeptide comprenant :
55 la culture d'une cellule dans laquelle a été introduit un vecteur d'expression selon la revendication 31 ;
 de sorte que ladite cellule exprime ledit polypeptide codé par ladite molécule polynucléotidique ; et
 la récupération dudit polypeptide exprimé.

34. Polypeptide isolé ayant la séquence de SEQ ID NO:2.

35. Polypeptide selon la revendication 34 en combinaison avec un véhicule pharmaceutiquement acceptable.

5 36. Composition pharmaceutique comprenant :

- a) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:2 ;
- b) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:4 ;
- c) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:6 ;
- 10 d) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:8 ; ou
- e) un anticorps ou fragment d'anticorps qui se lie spécifiquement à un polypeptide de SEQ ID NO:10 ; et un support pharmaceutiquement acceptable.

15 37. Composition selon la revendication 36 où ledit anticorps ou fragment d'anticorps est choisi dans le groupe consistant en :

- a) un anticorps polyclonal ;
- b) un anticorps monoclonal murin ;
- c) un anticorps humanisé dérivé de b) ; et
- 20 d) un anticorps monoclonal humain.

38. Composition selon les revendications 36 et 37 où ledit fragment d'anticorps est choisi dans le groupe consistant en F(ab'), Fab, Fv, scFv.

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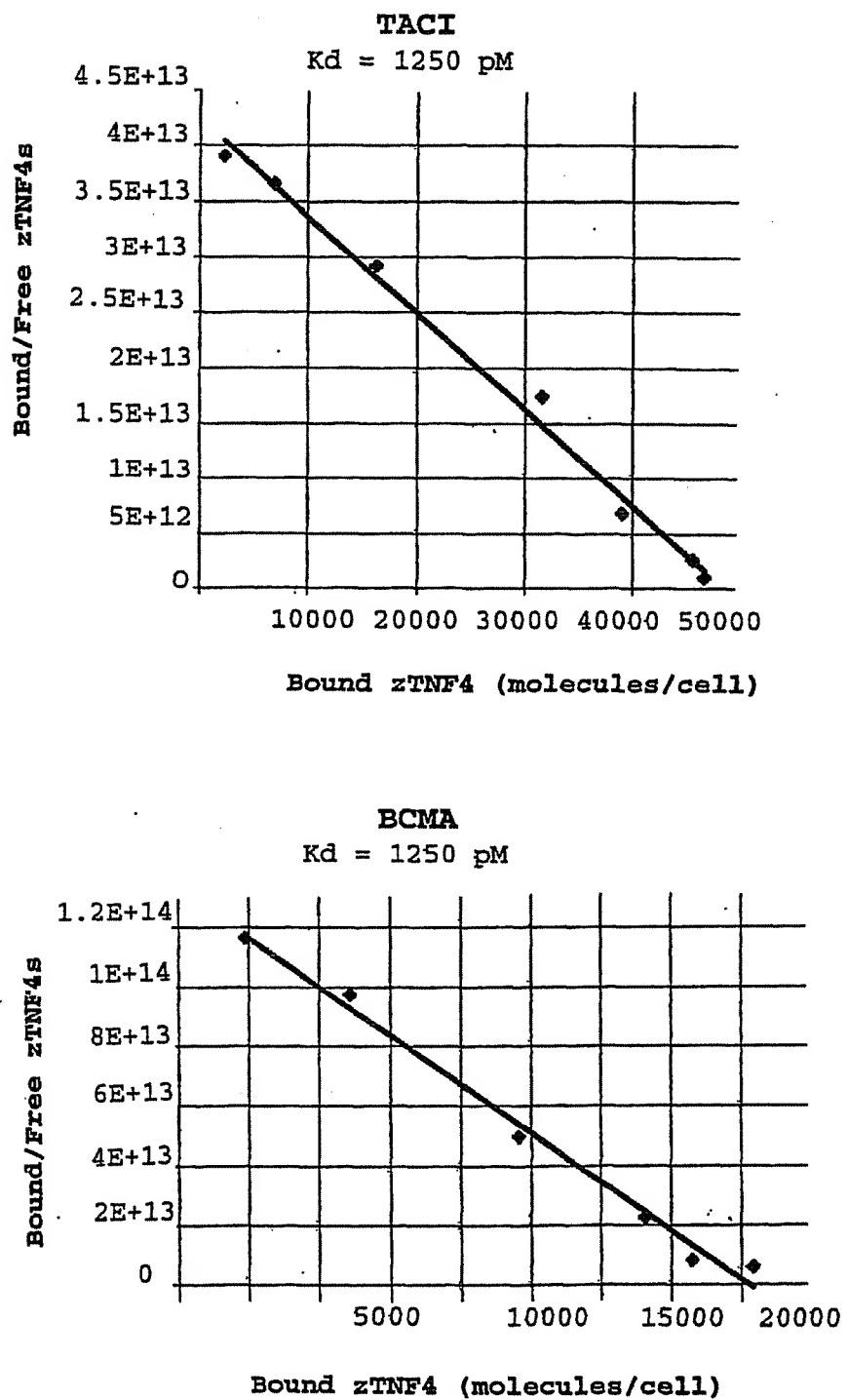
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TacI	-----	-----	-----	MSGLGRSRRG
BR43X1	-----	-----	-----	-GRSRRG
BR43X2	-----	-----	-----	MSGLGRSRRG
BCMA	-----	-----	-----	
 TacI	GRSRVDQEER	FPQGLWTGVA	MRSCPPEEQYW	DPLL-GTCMS CKTICNHQSQ
BR43X1	GRSRVDQEER	FPQGLWTGVA	MRSCPPEEQYW	DPLL-GTCMS CKTICNHQSQ
BR43X2	GRSRVDQEER	-----	-----	-----
BCMA	-----	-----	MLQM AGQCSQNEYF	DSLL-HACIP CQLRCSSNTP
				<--- 1st cys repeat -----
 TacI	-RTCAAFCRS	L-----SC	RKEQGKFYDH	LL-RD-CISC ASICGQHPKQ
BR43X1	-RTCAAFCRS	L-----SC	RKEQGKFYDH	LL-RD-CISC ASICGQHPKQ
BR43X2	-----WS	L-----SC	RKEQGKFYDH	LL-RD-CISC ASICGQHPKQ
BCMA	PLTCQRYCNA	SVTNNSVKGTN	AILWTCLGLS	LIISLAVFVL MFLLRKISSE
	----->			<--- 2nd cys repeat -----
 TacI	CAYFCENKLR	SPVNLPPPELR	RQRSGEVENN	SDNSGRYQGL EHRGSEASPA
BR43X1	CAYFCENKLR	SPVNLPPPELR	RQRSGEVENN	SDNSGRYQGL EHRGSEASPA
BR43X2	CAYFCENKLR	SPVNLPPPELR	RQRSGEVENN	SDNSGRYQGL EHRGSEASPA
BCMA	PLKDEFKNTG	SGLLGMANID	LEKSRTGDEI	ILPRGLEYTV EECTCEDCIK
	----->			
 TacI	LPGLKLSADQ	VALVYSTLGL	CLCAVLCCFL	VAVACFLKKR GDPCSCQPRS
BR43X1	LPGLKLSADQ	VALVYSTLGL	CLCAVLCCFL	VAVACFLKKR GDPCSCQPRS
BR43X2	LPGLKLSADQ	VALVYSTLGL	CLCAVLCCFL	VAVACFLKKR GDPCSCQPRS
BCMA	SKPKVDSDH	FPLPAMEEGA	TILVTTKTND	YCKSLPAALS ATEIEKSISA
				<--- TACI/BR43 TM --->
 TacI	RPRQSPAKSS	QDHAMEAGSP	VSTSPEPVET	CSFCFPECRA PTQESAVTPG
BR43X1	RPRQSPAKSS	QDHAMEAGSP	VSTSPEPVET	CSFCFPECRA PTQESAVTPG
BR43X2	RPRQSPAKSS	QDHAMEAGSP	VSTSPEPVET	CSFCFPECRA PTQESAVTPG
BCMA	R-----	-----	-----	-----
 TacI	TPDPTCAGRW	GCHTRTTVLQ	PCPHIPDSGL	GIVCVPAQEG GPGA-----
BR43X1	TPDPTCAGRW	GCHTRTTVLQ	PCPHIPDSGL	GIVCVPAQEG GPGA-----
BR43X2	TPDPTCAGRW	GCHTRTTVLQ	PCPHIPDSGL	GIVCVPAQEG GPGA-----
BCMA	-----	-----	-----	-----

FIGURE 1

**Figure 2**

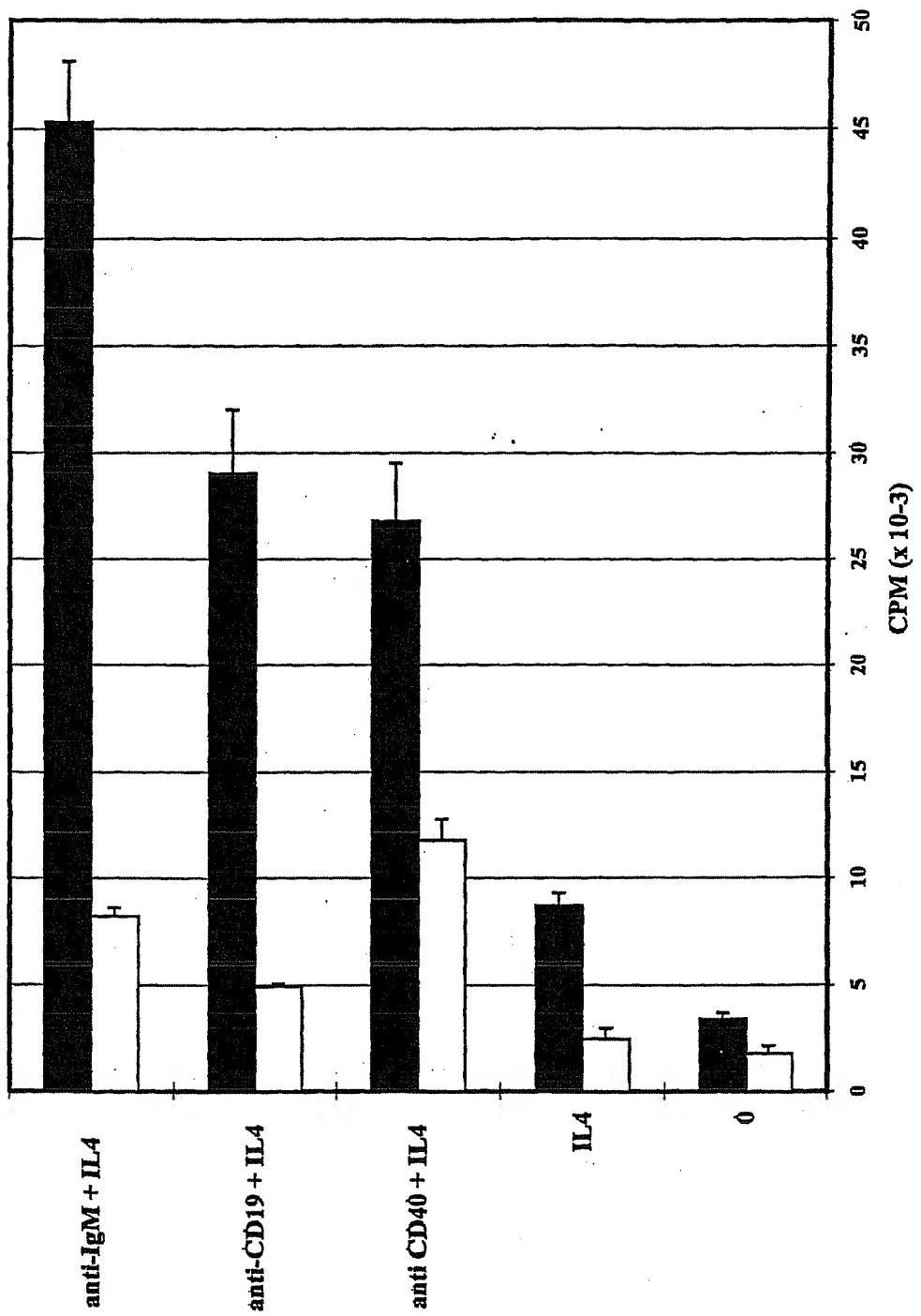


Figure 3A

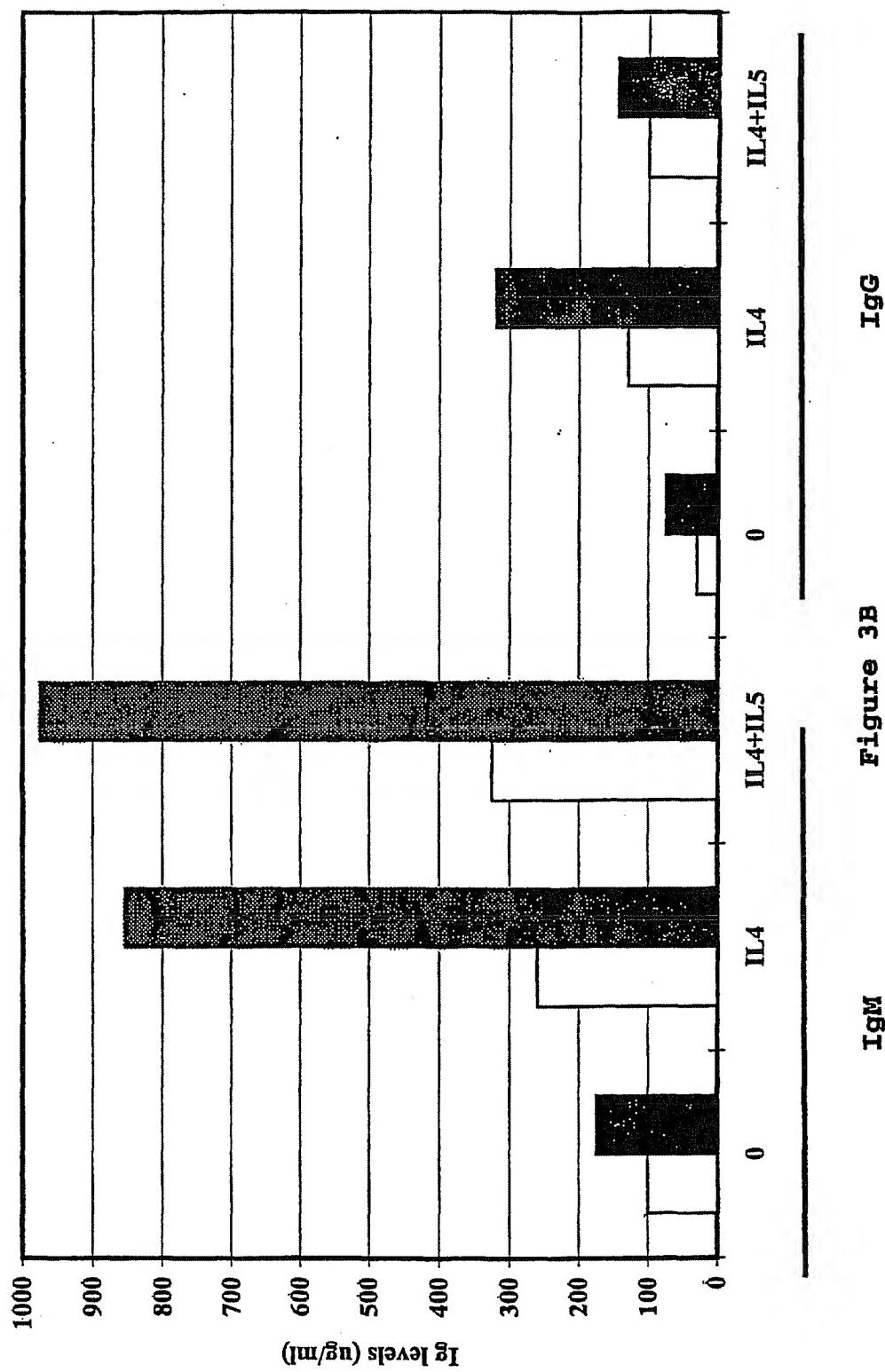


Figure 3B
IgM IgG

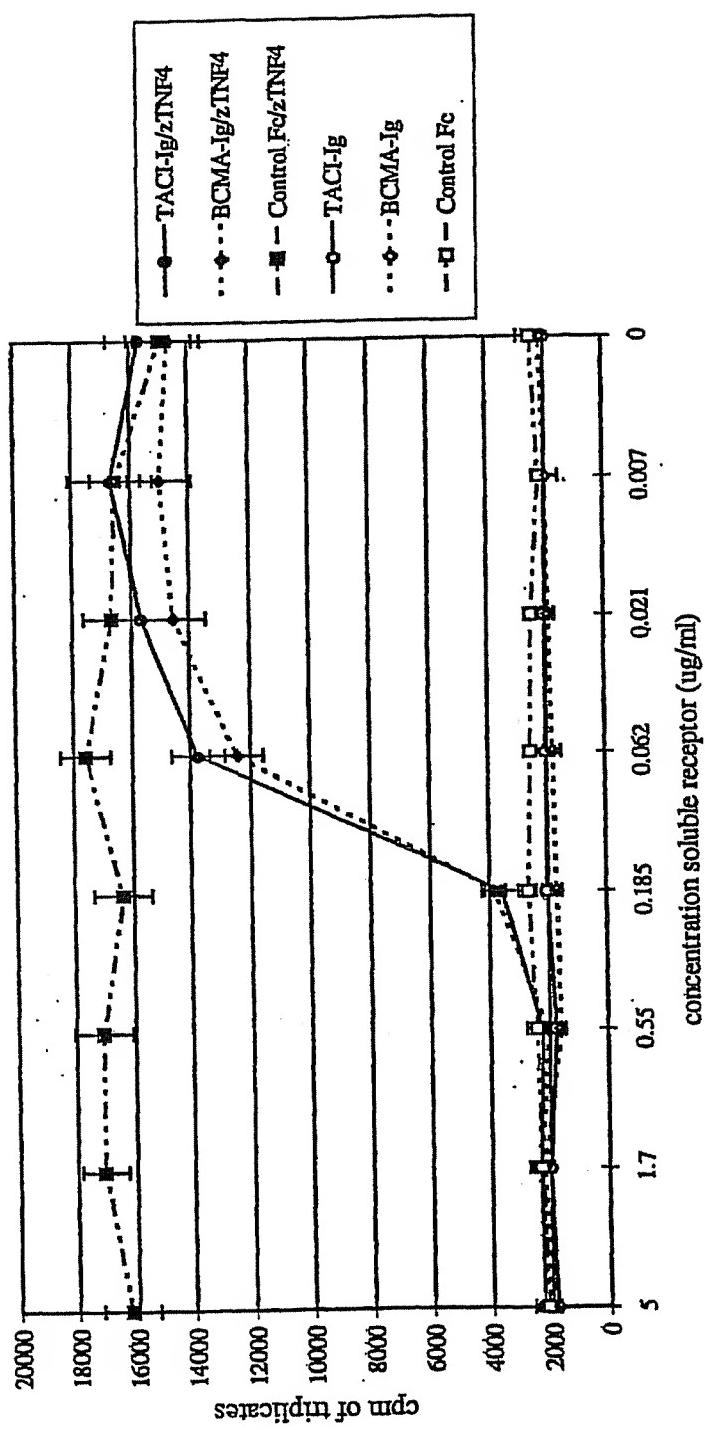
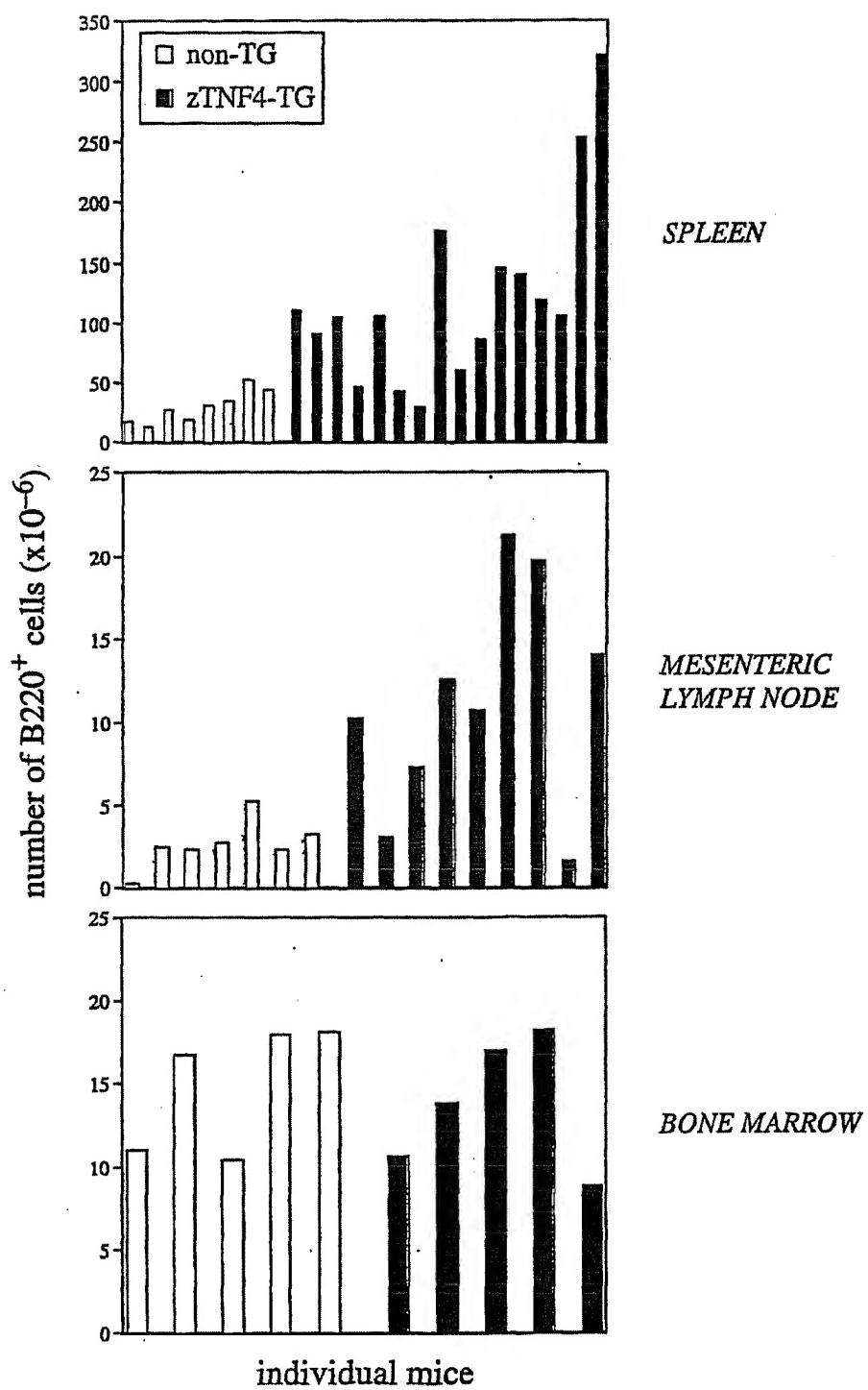


Figure 4



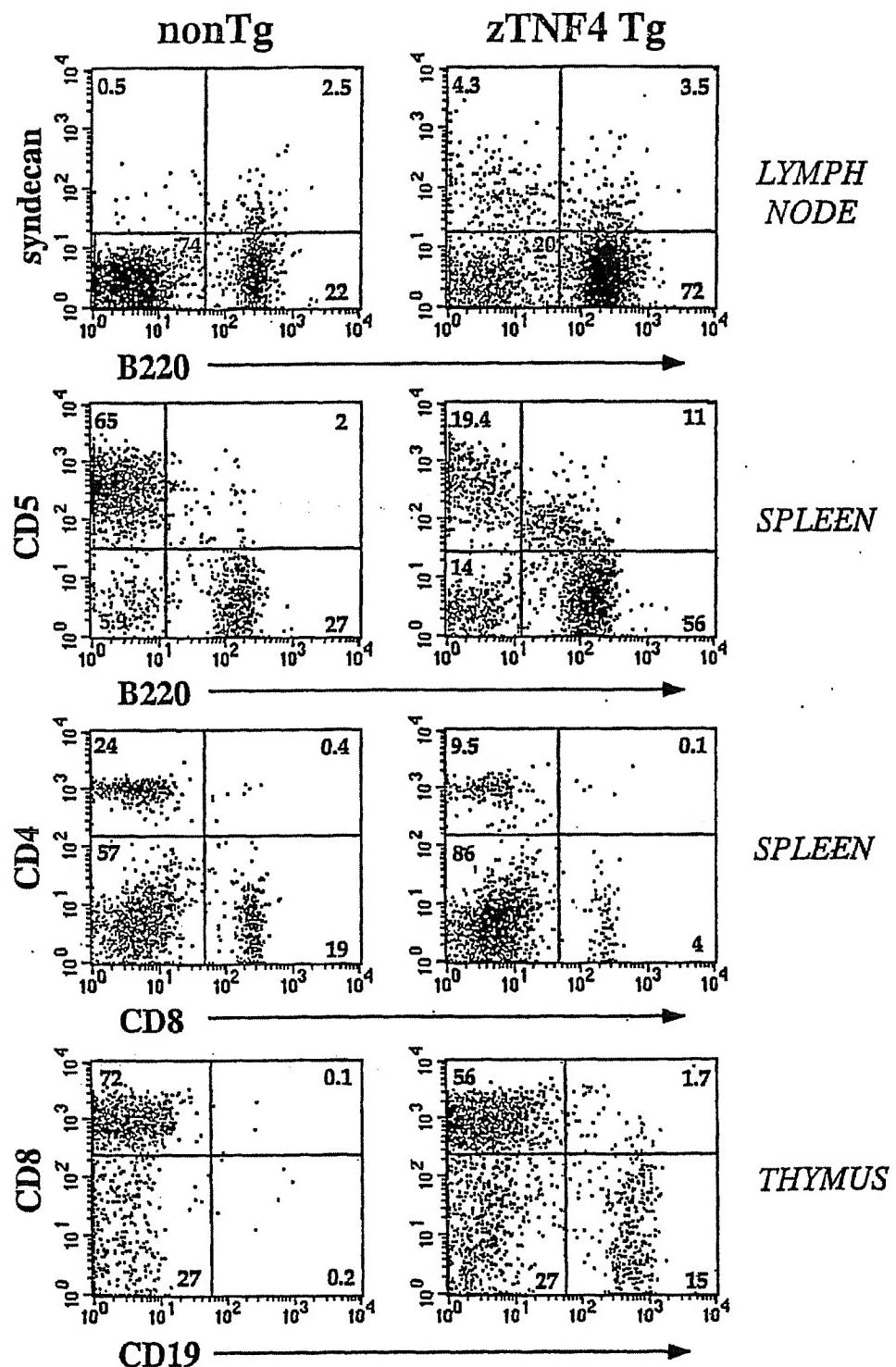


Figure 5B

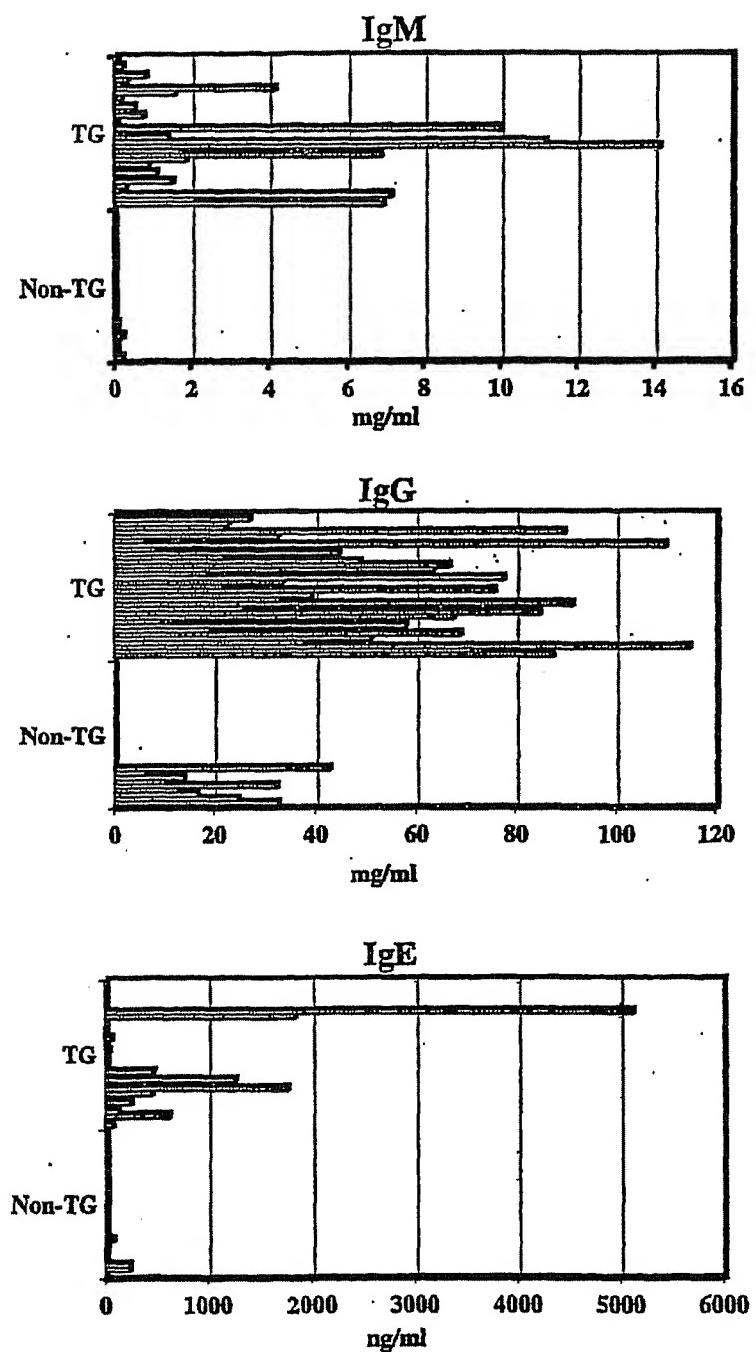


Figure 5C

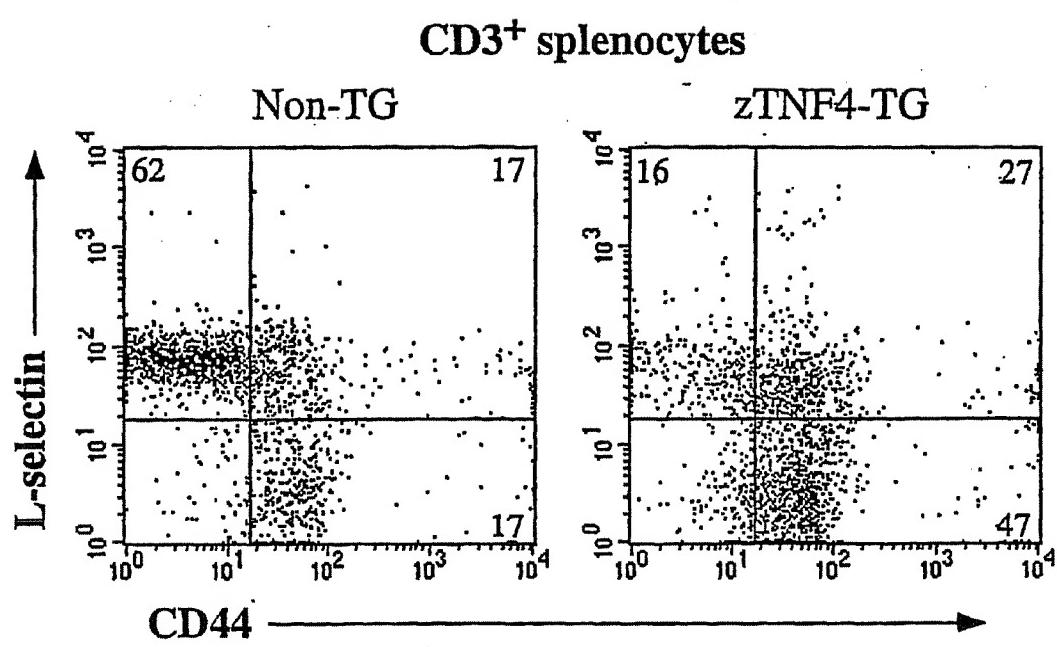


Figure 5E

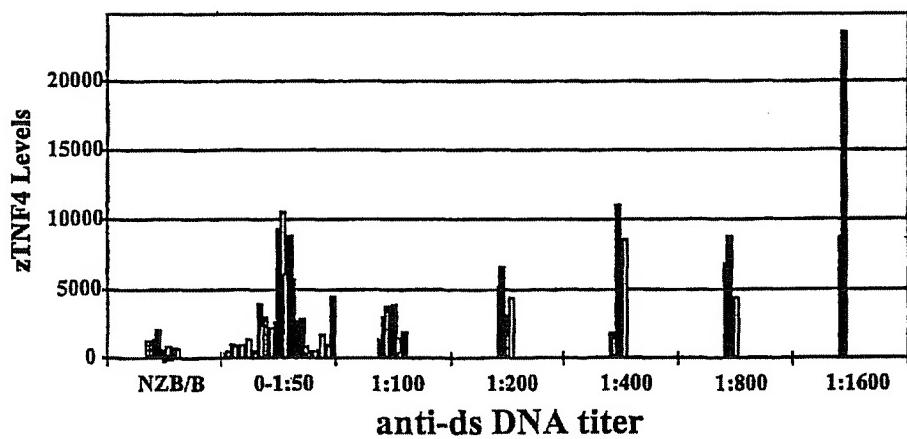
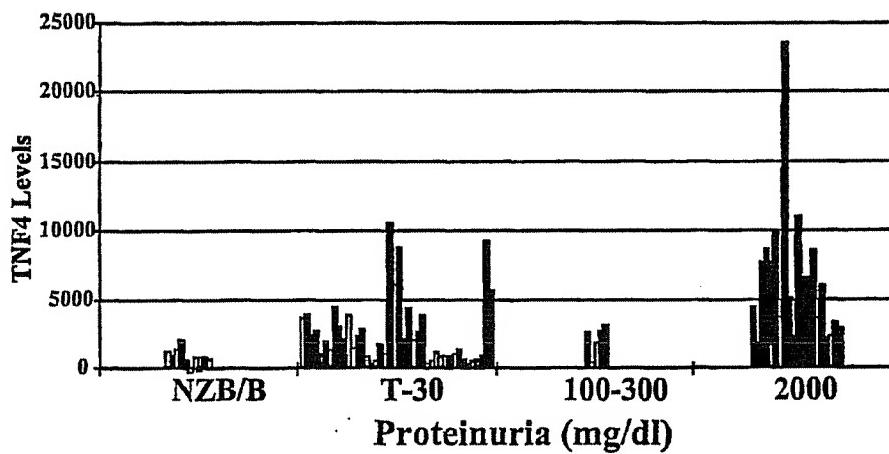
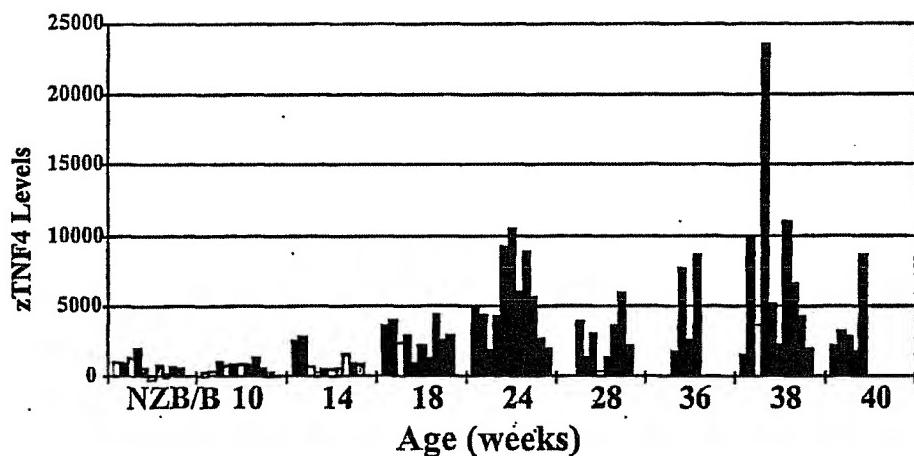
NZBWF1

Figure 6A

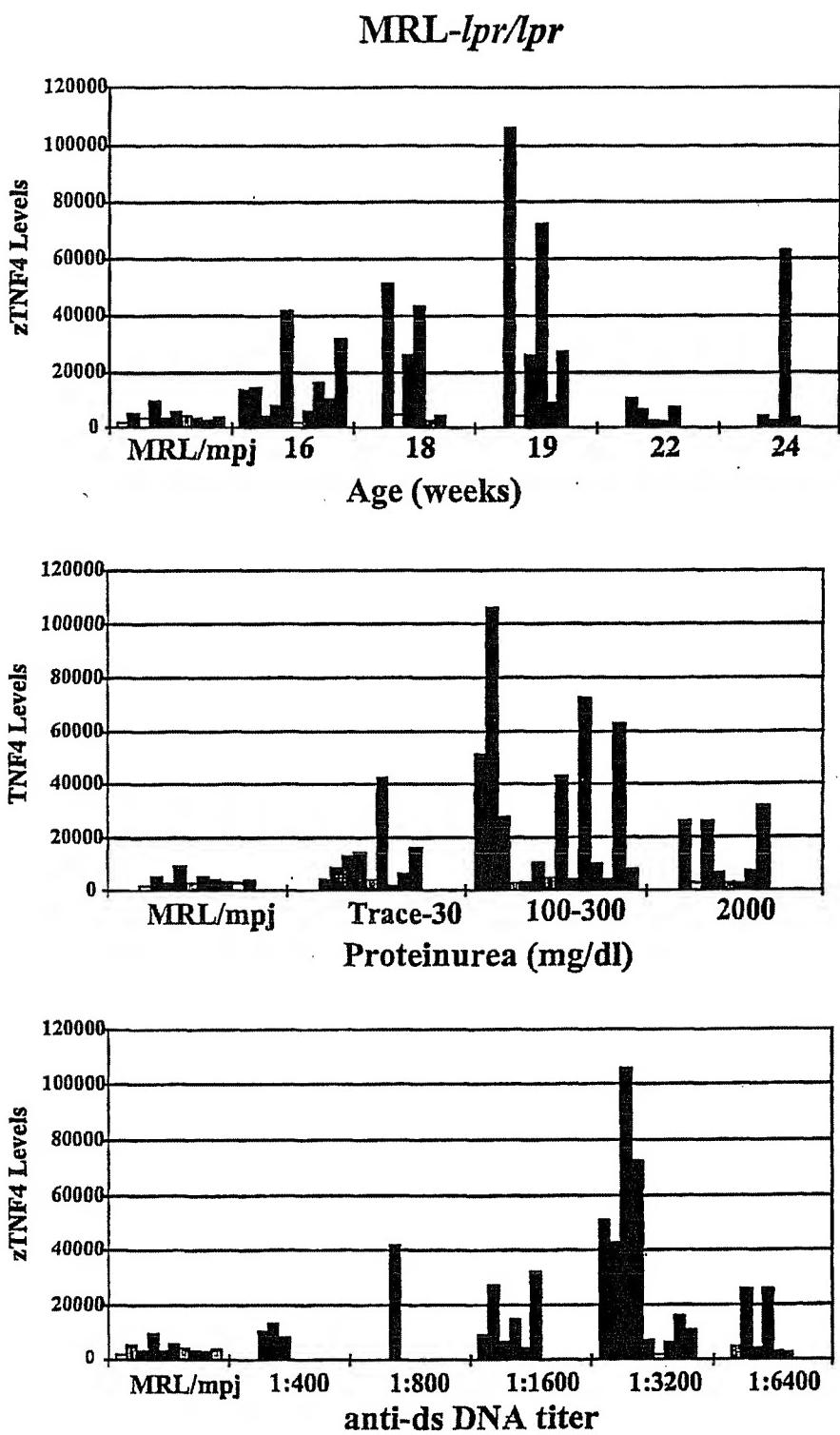


Figure 6B

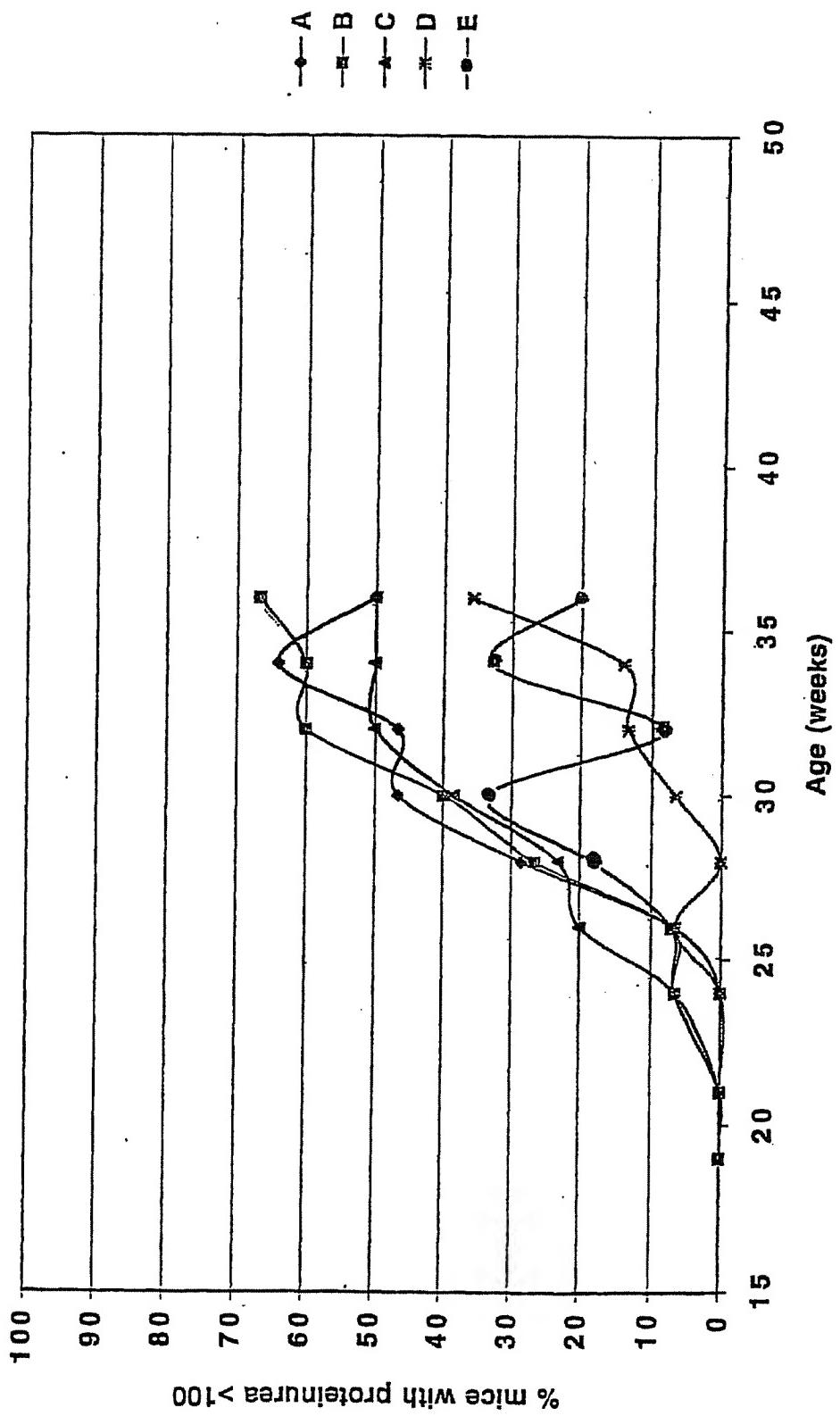


Figure 7

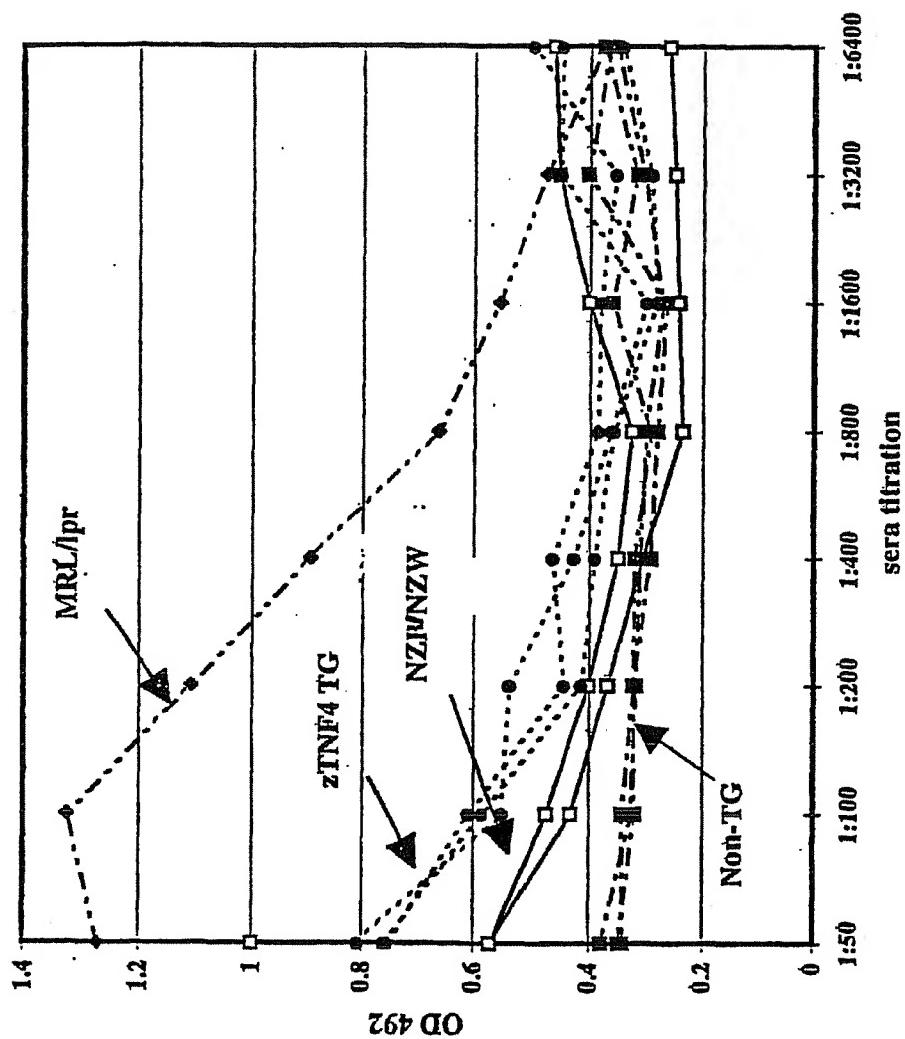


Figure 8